

AD \_\_\_\_\_

Award Number: DAMD17-97-1-7009

TITLE: Emergency Interventions After Severe Traumatic Brain  
Injury in Rats: Effect on Neuropathology and Functional  
Outcome

PRINCIPAL INVESTIGATOR: Patrick M. Kochanek, M.D.

CONTRACTING ORGANIZATION: University of Pittsburgh  
Pittsburgh, Pennsylvania 15260

REPORT DATE: August 2000

TYPE OF REPORT: Final Supplemental

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for public release;  
distribution unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

DTIC QUALITY INSPECTED 4

20001027 025

# REPORT DOCUMENTATION PAGE

Form Approved  
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE August 2000	3. REPORT TYPE AND DATES COVERED Final Supplemental (15 Dec 96 - 14 Jul 00)	
4. TITLE AND SUBTITLE Emergency Interventions After Severe Traumatic Brain Injury in Rats: Effect on Neuropathology and Functional Outcome			5. FUNDING NUMBERS DAMD17-97-1-7009	
6. AUTHOR(S) Patrick M. Kochanek, M.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Pittsburgh Pittsburgh, Pennsylvania 15260  E-MAIL: kochanekpm@anes.upmc.edu			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)  U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES Report contains color graphics.				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited				12b. DISTRIBUTION CODE
13. ABSTRACT (Maximum 200 Words)  Traumatic brain injury (TBI) contributes to combat morbidity/mortality. Studies in models of TBI have focused on novel mediators and mechanisms. We used controlled cortical impact (CCI), a contemporary model of TBI in rats to study <i>field-oriented treatments</i> . The following technical objectives were addressed: 1) What is the optimal ventilation strategy? 2) Is hypothermia beneficial? and 3) What is the optimal sedative/analgesic? A fourth objective, combining hypothermia plus other therapies was abandoned due to the limited efficacy of hypothermia. The most important findings/publications include: Objective #1) In a report published in the <i>Journal of Neurosurgery</i> , we demonstrated that early aggressive hyperventilation worsened neuronal death. Objective #2) We published the first report showing that hypothermia was ineffective in the combat-relevant scenario of CCI followed by secondary hypoxemia. That work is <i>in press</i> in <i>Critical Care Medicine</i> . Objective #3) We reported remarkably poor outcome in rats treated with narcotics (fentanyl) versus general anesthesia (isoflurane) after CCI. That work was presented at the 1999 meeting of the National Neurotrauma Society by research trainee Dr. Kimberly Statler, who received the Women in Neurotrauma Award. The paper is <i>in press</i> in the <i>Journal of Neurotrauma</i> . Since narcotics are the current field treatment after TBI, we then completed a seven anesthetic study in our CCI model. This is the first comprehensive assessment of the effect of anesthetic agents on outcome in experimental TBI. Notably, agents that are commonly used in the field (narcotics, benzodiazepines, and propofol) had detrimental effects on selective aspects of outcome. The results of this study will be presented at the National Neurotrauma Society meeting this fall.				
14. SUBJECT TERMS Head and Trauma			15. NUMBER OF PAGES 104	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)  
Prescribed by ANSI Std. Z39-18  
298-102

## FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

\_\_\_ Where copyrighted material is quoted, permission has been obtained to use such material.

\_\_\_ Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

\_\_\_ Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

X In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

N/A For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

N/A In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

N/A In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

N/A In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.



PI - Signature

Date

## TABLE OF CONTENTS

FRONT COVER .....	1
STANDARD FORM (SF), REPORT DOCUMENTATION PAGE .....	2
FOREWORD.....	3
TABLE OF CONTENTS .....	4
INTRODUCTION.....	5
BODY.....	7
KEY RESEARCH ACCOMPLISHMENTS.....	16
REPORTABLE OUTCOMES .....	16
CONCLUSIONS .....	18
REFERENCES .....	19
APPENDIX .....	20



## (5) INTRODUCTION

Please note that this publication represents a supplement to our final report, which was submitted in January 2000. There are still a number of manuscripts and abstracts that are either *in press*, *in submission* or *in preparation*. In addition to this revised final report, further supplements will follow for subsequent publications.

Traumatic brain injury (TBI) is an important contributor to combat casualty morbidity and mortality. Although progress has been made in the development of rodent models of TBI (such as the controlled cortical impact (CCI) model), most of the studies with these models have focused on molecular mechanisms of damage. Because there has been a paucity of application of **practical emergency interventions** in TBI models, we felt that it was important to address this deficiency since this could have important implications for field and emergency management of both soldiers and civilians. Our overall **hypothesis** is that optimal manipulation of practical interventions applicable in the emergency treatment of severe TBI (respiratory management, temperature control, and sedation) can reduce secondary brain injury in a rat model of brain contusion, and thereby improve functional and/or neuropathological outcome.

### *Funding year 1*

In yr. 1 of funding, we addressed the **first Technical Objective**, namely, to investigate the effects of mechanical ventilation strategies (as applied by the first responder in the field) on functional and neuropathological outcome in our model. **We found that aggressive, prophylactic hyperventilation (HV) applied for 4 h immediately after injury is detrimental (vs normal PaCO<sub>2</sub>), and leads to increased neuronal death in selectively vulnerable brain regions.** This study was published as a full manuscript in the *Journal of Neurosurgery* (1). The reviewers indicated that this was an important study that would be cited often. Dr. M. Forbes, a Critical Care Medicine fellow training in research with Dr. Kochanek, authored the study.

To set the stage for the evaluation of therapies after injury (as proposed studies in technical objectives 2-4), it appeared that it would be optimal to have TBI models both with and without a secondary insult-- since such insults are common in the field. This was accomplished by two studies assessing our model (2,3), including, adding a 30-min period of moderate hypoxemia to the CCI. Characterization of that model was described in the 1997 report and was presented in 1998 at the National Neurotrauma Society Meeting (3). During yrs. 2 and 3, we used both the CCI model and the CCI plus secondary hypoxemia model to test therapies.

### *Funding year 2*

In yr. 2 we performed three studies addressing Technical Objective 2 and part of Objective 3. These studies included: 1) assessment of the effect of transient (4 h), moderate hypothermia on outcome after TBI with a secondary insult, 2) assessment of the effect of prolonged (12-h) moderate hypothermia on outcome after TBI, and 3) assessment of the effect of anti-excitotoxic therapy (the NMDA-receptor antagonist MK-

801) early after TBI. **The results of these studies showed that hypothermia (32 °C, for either 4 h or 12 h) reduced DNA damage early after injury, but beneficial effects on long-term outcome could not be demonstrated in the model. It was particularly ineffective after the combined CCI plus hypoxemia. In contrast, we were surprised to find that MK-801 improved functional outcome. However, neither treatment improved brain histopathology after injury.** Three research fellows (Drs. C. Robertson, M. Whalen, and R. Ruppel) worked on these projects with the PI (Dr. Kochanek) during yr. 2. Dr. Robertson presented an abstract at the 1999 annual meeting of the Society of Critical Care Medicine (4). That work on hypothermia is *in press* as a full manuscript in the journal *Critical Care Medicine* (5). We also reported that 4 h of moderate hypothermia attenuates DNA damage after injury (6,7). That work was presented last year at the Society of Critical Care Medicine Meeting and the manuscript is in preparation. Our work on hypothermia in TBI was summarized in an invited review article published by the International Trauma, Anesthesia and Critical Care Medicine Society (ITACCS)(8). Dr. Randall Ruppel presented the work on MK-801 at the 1999 Meeting of the National Neurotrauma Society (9). It was one of 12 papers selected for oral presentation out of over 200 papers submitted. The manuscript is in preparation for submission to the *Journal of Neurotrauma*.

### *Funding year 3*

During yr. 3, we carried out a comprehensive study of sedation/analgesia comparing a narcotic (fentanyl) to a conventional general anesthetic (isoflurane). Fentanyl and morphine are the most commonly used narcotics after human head injury while isoflurane is the most commonly used anesthetic in rat models. **We reported remarkably poor outcome in rats treated with narcotics (fentanyl) versus general anesthesia (isoflurane) after CCI. That study is extremely relevant since narcotics are the current field treatment for combat casualties.** That work was presented by Critical Care Medicine fellow research trainee Dr. Kimberly Statler at the 1999 meetings of the National Neurotrauma Society, the Society for Neuroscience, and the Society of Critical Care Medicine (10-12). **Dr. Statler received the 1999 Women in Neurotrauma Award at the National Neurotrauma Meeting and an Educational Scholarship from the Society of Critical Care Medicine.** The full paper is *in press* in the *Journal of Neurotrauma* (13). The lack of beneficial effect of hypothermia in our model coupled with the remarkably powerful effect of isoflurane suggested the elimination of technical objective 4 in favor of a more comprehensive study of sedatives/analgesics early after CCI (i.e., expansion of proposed technical objective 3).

**We recently completed work on the final study in this proposal, namely a comparison of 7 sedative/analgesic treatments applied in a field relevant paradigm. This is the first comprehensive assessment of the effect of anesthetic agents on outcome in experimental TBI. This study showed detrimental effects of narcotics, diazepam, and propofol after TBI, and suggested powerful benefits of isoflurane at or near the time of injury. These findings suggest important implications for acute patient care, especially field application, as well as for experimental models of TBI.** Dr. Kimberly Statler will present the results of this study at the National Neurotrauma Society meeting this fall.

## **(6) BODY**

### **(a) Technical Objective 1: Mechanical ventilation strategies after severe TBI in rats (see summary for 1996-1997 [yr-1] and reference 1, both in appendix).**

We showed that aggressive, early HV after TBI augments neuronal death in CA3 hippocampus in a rat model of cerebral contusion. The further reduction of CBF with HV during the low cerebral blood flow state immediately after injury coupled with alkalosis may increase the vulnerability of selected neurons to damage. The results of this study reinforce the meticulous attention necessary to prevent secondary injury after TBI. A risk in the use of HV is demonstrated.

**Recommendation:** Unless signs of impending herniation (unilateral pupillary dilation, hypertension, bradycardia) are present, this study supports the targeting of normocarbica for mechanical ventilation in the emergency stabilization of the brain trauma victim.

### **(b) Technical Objectives 2: Testing of field-relevant therapies (notably hypothermia) in experimental models of severe TBI (with and without a secondary hypoxemic insult) in rats.**

**(b1)** *Effect of transient (4 h), moderate (32 °C) hypothermia on functional and histological outcome after experimental TBI with a superimposed secondary hypoxemic insult in rats. (see summary for 1998-1999 [yr-2] and reference 5, both in appendix).*

We tested the effect of 4 h of hypothermia in our model of TBI with a 30-min secondary hypoxemic insult. Hypothermia is effective in a variety of experimental models with transient application (1-4 h) and in a single-center study in humans (32°C applied for 24 h). However, in neither experimental nor clinical TBI has hypothermia been tested when applied after the combination of TBI with a secondary insult. We found no benefit from hypothermia after this field-relevant combined insult. This suggests three possibilities. First, the combination of TBI plus a secondary hypoxemic insult may be so severe that no single treatment will be effective. Second, the insult is so severe that no therapies will be effective. Third, based on our work in technical objective 3, it is possible that a beneficial effect of hypothermia is being masked by using isoflurane anesthesia (vida infra). This work is *in press* as a full paper in the journal *Critical Care Medicine* (5).

**Recommendation:** Even in centers where hypothermia was shown to be effective after TBI, this has not been the case for severely injured patients GCS 3-4. It is likely that severe injuries, such as that modeled by CCI with a 30-min hypoxemic secondary insult, will require combination therapies or may be refractory to all therapy. Also, based on the work by our group on technical objective #3, it will be important in future studies to test hypothermia using a sedative/analgesic approach that is similar to that used in the field or

clinic—i.e., with narcotics. To our knowledge, such a study has never been carried out in a rodent model of TBI.

**(b2)** *Effect of prolonged (12 h), moderate (32 °C) hypothermia on functional and histological outcome after experimental TBI in rats. (see summary for 1998-1999 [yr-2] in appendix for detailed methods).*

Based on the aforementioned study, in the CCI model, we sought to test, to our knowledge for the first time in any laboratory, the prolonged application of hypothermia in a rodent model of TBI. This included over 13-h of controlled mechanical ventilation and physiological monitoring. To date, only brief 1-4-h applications have been tested. In this study, we examined TBI without a secondary insult. **As described in last year's progress report, we failed to observe important beneficial effects of 12-h of hypothermia on functional or histopathological outcome after CCI.** This is a surprising finding which is discussed below.

**Recommendation:** Studies of 12-h of hypothermia in any experimental animal model are very labor intensive. The negative result of this study suggests one of two possibilities. First, there may be both beneficial and deleterious aspects to the use of hypothermia. Despite promising data from single clinical sites, recently, a randomized, controlled multi-center trial of hypothermia in human head injury failed to yield a positive result. **Second, once again based on the work by our group on technical objective #3, it will be important in future studies to test hypothermia using a sedative/analgesic approach that is similar to that used in the field or clinic—i.e., with narcotics.** It is our recommendation that this be tried first in a rodent model using either morphine or fentanyl anesthesia followed by either 1 or 4-h of hypothermia vs normothermia.

**(b3)** *Effect of the anti-excitotoxic NMDA-receptor antagonist MK-801 on functional and histological outcome after experimental TBI in rats. (see summary for 1998-1999 [yr. 2], in appendix, for detailed methods).*

In the third treatment trial in yr. 2, we sought to test the effect of the traditional NMDA-receptor antagonist MK-801 in our TBI model (without secondary insult). **The NMDA antagonist MK-801 was effective in improving both motor function and some aspects of cognitive function after CCI.** The motor effects were more dramatic than those seen with hypothermia. In a separate pilot study, MK-801 was not effective when tested in our TBI plus secondary hypoxemia model, again suggesting this insult may be too severe for any single therapy. Although this specific agent is not available for clinical use, it suggests that this category of agents—targeting excitotoxicity—represents a viable strategy.

**Recommendation:** This finding is particularly relevant since our studies addressing technical objective #3 suggested that an anti-excitotoxic general anesthesia strategy such as isoflurane produced a markedly better outcome than treatment with the narcotic analgesic fentanyl. Although there have been several negative clinical trials of anti-

excitotoxic therapy, there is frequently a delay in administration of treatment for as much as 6-h in these trials. Several reports have suggested that important components of the excitotoxic response may occur in the initial 1-2-h after injury. **Based on our findings, anti-excitotoxic strategies should not be abandoned, rather consideration should be given to the field application of these strategies. In addition, sedative/analgesics with anti-excitotoxic properties must be extensively studied in experimental TBI, in both small animal and large animal models.**

**(c) Technical Objective 3: Testing of the optimal field-relevant sedative/analgesic therapy in an experimental model of severe TBI in rats (see reference 13 in appendix).**

*(c1) Comparison of the effects of TBI on functional and histological outcome after experimental TBI in rats anesthetized with fentanyl or isoflurane (described below).*

Currently, in clinical practice, fentanyl is the most commonly used emergency sedative for intubated patients with severe TBI. Fentanyl, has little direct anti-excitotoxic properties. Thus, to begin investigating this area, we tested how fentanyl treatment compared to standard isoflurane anesthesia in our model.

#### *Outcome protocol*

Rats were initially anesthetized with N<sub>2</sub>O:O<sub>2</sub> (2:1) and 4% isoflurane and then endotracheally intubated and mechanically ventilated. Anesthesia was maintained for the preparatory surgery with 2 - 2.5% isoflurane and N<sub>2</sub>O:O<sub>2</sub> (2:1). Pancuronium bromide (0.1 mg/kg/h) was given iv for muscle relaxation. Femoral venous and arterial vessels were cannulated for continuous blood pressure measurement, blood sampling, and administration of medications. A rectal probe was inserted to monitor core temperature. The rat was then placed in a stereotaxic frame and a left parietal craniotomy was performed. The dura and bone flap were left in place until immediately before CCI. A burr hole was drilled into the left frontal bone for temperature probe placement into the frontal lobe. Continuously monitored physiologic parameters included arterial blood pressure and rectal and brain temperatures. Blood glucose, hematocrit, and arterial blood gas samples were assessed every 15 min for the initial hour and every 30 min thereafter. PaCO<sub>2</sub> was controlled at 35 - 45 mm Hg. This protocol produced a PaO<sub>2</sub> of greater than 70 mm Hg in all preparations. Both brain and rectal temperatures were maintained at 37.0 ± 0.5 °C.

Rats were allowed to stabilize for 5-min after completion of surgical preparation and then randomized to receive either fentanyl or isoflurane anesthesia. In the fentanyl group (n=9), isoflurane was discontinued and 10 µg/kg of fentanyl was administered iv, followed by a continuous iv infusion at 50 µg/kg/h. In the isoflurane group (n=9), inspired isoflurane concentration was reduced to 1%, and normal saline, the vehicle for fentanyl-treated rats, was administered to match the volume received by fentanyl infusion. Both anesthetic groups continue to receive N<sub>2</sub>O:O<sub>2</sub> (2:1). After 30-min equilibration, TBI was induced by CCI. In pilot studies comparing isoflurane and



fentanyl using our standard CCI model (6-mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.5-mm deformation depth), all of the fentanyl-treated rats developed pulmonary edema and died early after injury. Thus, to compare the effect of isoflurane vs fentanyl on long-term outcome in our model, our standard injury was reduced (2.0-mm deformation depth). After CCI, the bone flap was replaced and sealed with dental cement, and the scalp incision was closed. Anesthesia was continued for 4-h in the isoflurane group and 3.5-h in the fentanyl group to facilitate similar extubation times. At the end of the anesthetic period, rats received 100% oxygen, were allowed to awaken and resume spontaneous breathing, and were then extubated and returned to their cages. Sham rats underwent identical preparation/anesthesia, but no CCI ( $n=6$  per group).

Motor function, including beam balance and beam walking tasks, was tested by an observer blinded to group assignment on days 1-5 after injury. Morris Water Maze (MWM) testing was performed using an acquisition paradigm on days 14-20 after injury. Lesion volume and hippocampal neuron survival were assessed on day 21.

### ICP Protocol

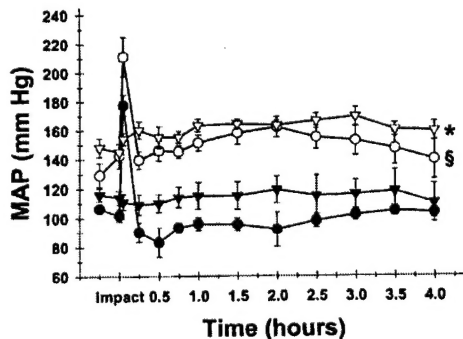


Fig 1: MAP vs time after injury. MAP in fentanyl-treated injured (open circles) and sham (open triangles) rats was ~50 mm Hg higher than in isoflurane-treated rats at all time points (injured shown by closed circles and shams by closed triangles). \*  $p < 0.05$ , isoflurane vs fentanyl at each time after injury, §  $p < 0.05$ , isoflurane vs fentanyl at all time points, including baseline, in shams.

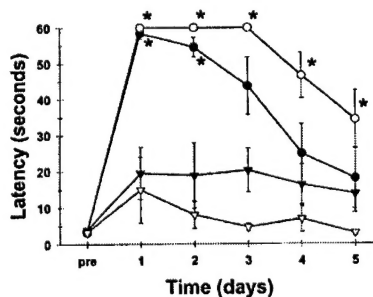


Fig 2: Beam walking latency vs d after injury. Isoflurane-treated rats recovered by post-injury d 3, fentanyl-treated rats failed to regain normal function by the end of the 5-d period. \*  $p < 0.05$ , injured vs sham. Beam balance latency showed similar benefit of isoflurane vs fentanyl.

Based both on results of the above protocol showing that fentanyl-treated rats had higher MAP throughout the experiment and on a recent report that increased MAP may exacerbate injury after TBI, ICP and percent brain water were monitored in a separate cohort of rats ( $n=9$  per anesthetic group) subjected to either fentanyl or isoflurane anesthesia and CCI in an identical paradigm to that used to assess functional outcome.

Surgical preparation, randomization, anesthetic administration, and CCI were identical to the outcome protocol, with minor exceptions. Specifically, an intraparenchymal ICP monitor (Codman microtransducer) was inserted through a burr hole in the frontal bone into the contralateral (right) frontal cortex at the time of craniotomy. After CCI, anesthesia was continued and ICP was monitored for 4-h in both anesthetic groups. Cerebral perfusion pressure (CPP) was calculated as the difference between MAP and ICP. Rats were killed by decapitation at the end of the anesthetic period. Brains were immediately removed and a 3-mm coronal slice was made through the center of the contusion. Per cent brain water was determined in the coronal

slice using the wet-dry weight method. Brain water content was determined in both the injured and the homologous region of the uninjured hemispheres.

As an added control, a separate cohort of rats ( $n=3$ ) was subjected to CCI and allowed to recover without anesthesia. These rats were prepared for CCI under isoflurane anesthesia as above, allowed to recover a tail-pinch response, and then subjected to CCI. Arterial MAP was monitored via a femoral arterial catheter for 4-h during recovery without anesthesia.

## Results

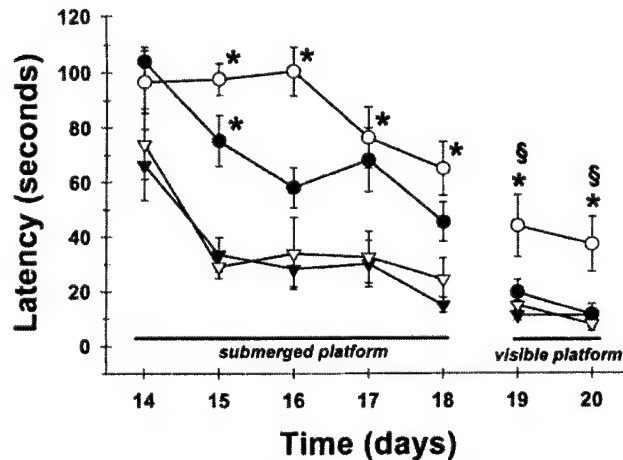


Fig 3: Latency to find platform vs time after injury in an acquisition paradigm of the MWM. Shams anesthetized with isoflurane (closed triangles) or fentanyl (open triangles) had similar performances. During the first few d of hidden platform testing, injured rats in fentanyl (open circles) and isoflurane (closed circles) groups had impaired performance vs sham. By d 3, latencies to find the hidden platform were similar in injured isoflurane-treated rats and shams. In contrast, longer latencies persisted in injured fentanyl-treated rats throughout the 5-d hidden platform testing. Latencies in all groups improved during visible platform testing; but, shams and injured isoflurane-treated rats performed better than injured fentanyl-treated rats during visible platform testing. \* $p < 0.05$ , injured vs sham; § $p < 0.05$ , isoflurane vs fentanyl.

compared to ~105 mm Hg in the isoflurane groups.

Rats anesthetized with isoflurane performed better on beam balance and beam walking

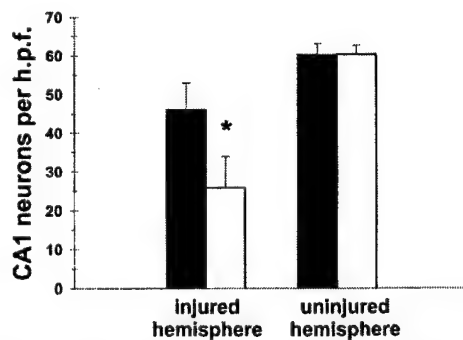


Fig 4: Neuron counts in injured CA1 hippocampus were greater in isoflurane- vs fentanyl-treated rats. Neuron counts in uninjured CA1 hippocampus were similar in both treatment groups. \* $p < 0.05$ , injured vs uninjured.

Time to extubation did not differ after injury between isoflurane and fentanyl treatment groups ( $269 \pm 7$  min vs  $275 \pm 15$  min,  $p = 0.29$ ). Physiologic values, including  $\text{PaCO}_2$ ,  $\text{PaO}_2$ , blood glucose and Hct did not differ between anesthetic groups. In contrast, MAP was higher in injured rats treated with fentanyl compared to their isoflurane counterparts ( $p < 0.05$ ) during the entire posttrauma period (Fig 1). Similarly, MAP was higher in shams treated with fentanyl vs isoflurane ( $p < 0.05$ ) during the entire duration of anesthesia (Fig 1). Fentanyl-treated rats had a MAP of ~150 mm Hg

tasks after TBI compared to their fentanyl counterparts ( $p < 0.05$ , Fig 2). Following injury, isoflurane-anesthetized rats also performed better than their fentanyl-treated counterparts during MWM testing with a hidden platform ( $p < 0.05$ , Fig 3). Motor and MWM performances did not differ between sham groups.

Lesion volume, expressed as  $\text{mm}^3$  or as percent of uninjured hemisphere, at 21-d did not differ between treatment groups (see reference 13 in appendix for details). In contrast, neuron counts in the injured CA1 hippocampus were markedly greater

in isoflurane-treated rats ( $p < 0.05$ , Fig 4). Neuron counts in the injured CA3 hippocampus, however, did not differ significantly between treatment groups (see reference 13 in appendix for details).

In the ICP protocol, again physiologic values, including  $\text{PaCO}_2$ ,  $\text{PaO}_2$ , glucose and Hct, did not differ between anesthetic groups. As in the outcome protocol, MAP was higher in rats treated with fentanyl compared to their isoflurane counterparts ( $p < 0.05$ ). ICP was similar in both anesthetic groups; however, there was a trend toward higher ICP in rats anesthetized with isoflurane by 3-4-h after TBI (Fig 5). This strongly suggests that the higher MAP in fentanyl vs isoflurane treated rats did not exacerbate intracranial hypertension. As expected from the difference in MAP, CPP was higher in the fentanyl treatment group (Fig 6).

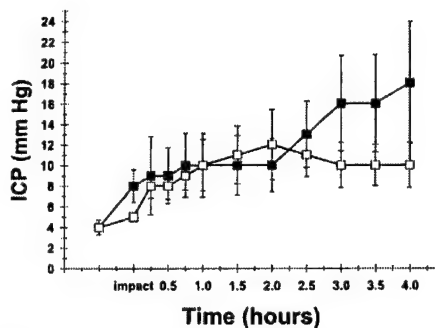


Fig 5: ICP vs time after injury. Initial ICP was ~4 mm Hg in both isoflurane (open squares) and fentanyl (closed squares) groups. ICP gradually increased, reaching 10-18 mm Hg by 4 h. ICP was similar between groups; but, isoflurane-treated rats showed a trend toward higher ICP after injury (vs fentanyl) that did not reach significance.

fentanyl-treated rats and rats recovering without anesthesia ( $p < 0.05$  vs both groups).

**Recommendation:** The theoretical advantages of isoflurane vs fentanyl are compelling; however, explanations for the observed improvement in neurologic outcome after TBI in isoflurane-anesthetized rats remain to be determined. What has become clearer is that

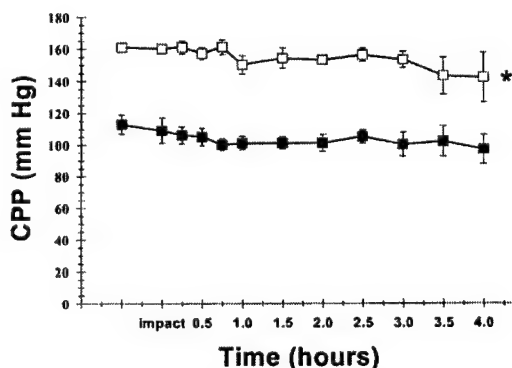


Fig 6: CPP vs time after injury. CPP was increased in rats treated with fentanyl (open squares) vs isoflurane (closed squares), \*  $p < 0.05$ , isoflurane vs fentanyl at all time points except 3.5 and 4h.

anesthetic agents may have considerable impact upon outcome following TBI. The results of this study suggest two important potential ramifications. First, isoflurane may not represent the optimal anesthetic in experimental TBI since it may mask potential benefits of novel therapies. Second, despite common clinical and field use, narcotics such as morphine or fentanyl may not be the optimal sedative/analgesic agent to administer to patients in the acute phase after severe head injury. We feel this has considerable relevance since narcotics



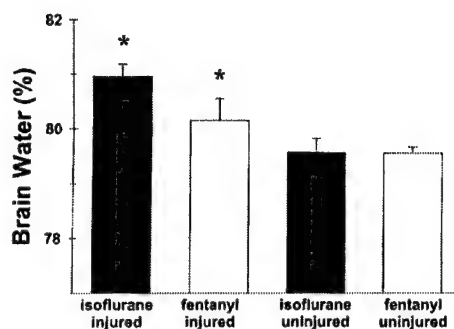


Fig 7: Percent brain Water 4 h after TBI. Percent brain water in the injured hemisphere was increased vs respective non-injured hemisphere in both isoflurane- and fentanyl-treated rats; however, brain water did not differ between anesthetic groups. \* $p < 0.05$ , isoflurane vs fentanyl.

(either fentanyl or morphine) are first line agents in field or emergency department. Consideration should also be given to the possibility that agents like isoflurane anesthesia could be provided in the field. Our suspicion was that narcotics are not deleterious, rather general anesthetics such as isoflurane are powerfully beneficial, possibly through anti-excitotoxic actions or cerebral blood flow promotion early after injury. Defining the factors responsible for improved outcome with isoflurane may help to direct the clinical application of more optimal sedative/analgesic agents and possibly to identify novel therapies. **Finally, since the**

**immediate post-trauma sedative/analgesic regimen had such a powerful effect on both functional and histopathological outcome, we completed a comprehensive comparison of field-relevant sedative/analgesic strategies (vida infra).**

## (C2) Randomized, blinded study in the rat model of CCI of seven different sedative/analgesic strategies for field use in TBI.

We have recently completed a nine group (seven anesthetic) study in our CCI model. Rats are prepared for TBI exactly as described in our protocol comparing isoflurane and fentanyl above. Anesthesia for surgical preparations is 2% isoflurane in nitrous oxide/oxygen (2:1). After surgical preparation, anesthesia is discontinued until tail-pinch response is obtained and then CCI is delivered. Rats are then randomized to one of the 8 groups below ( $n = 9$  per group, Table 1). There is also a sham group (thus, a total of 9 groups). The sedation or anesthesia is maintained for a period of 60 min and the rats are then weaned and extubated when recovered. Outcome parameters are identical to the isoflurane vs fentanyl outcome study (motor and MWM function; lesion volume, hippocampal CA1 and CA3 cell counts).

**Table 1. Sedation/analgesia study posttrauma**

Anesthetic/Sedative	Dose
Diazepam	3.4 mg/kg iv
Fentanyl	10 $\mu$ g/kg then 50 $\mu$ g/kg/h iv
Isoflurane	1% by inhalation
Ketamine	10mg/kg then 2.5 mg/kg/min iv
Morphine	10 mg/kg iv
Pentobarbital	50 mg/kg iv
Propofol	85 mg/kg/h iv
None <sup>†</sup>	NA
Sham	NA

<sup>†</sup>Isoflurane anesthesia discontinued and TBI immediately on return of tail pinch reflex.

Comment: First, pilot studies were conducted in our CCI model using each anesthetic. The selected doses were based on those commonly reported in the literature with minor adjustments based on our pilot data. As you are aware, our rat facility became infected with a virus midway through the

study, necessitating elimination of all potentially infected animals. The study was restarted in April 2000. Although the physiologic and functional studies are finished, we are still completing the histologic evaluation of brain tissue.

## Results

Physiologic parameters (brain temperature, blood gases, and glucose) did not differ between experimental groups, except for MAP (Fig 8). As seen in our prior study, MAP was higher in fentanyl vs isoflurane anesthetized rats (~135 mm Hg vs ~105 mm Hg,  $p < 0.05$ ). Although there was a trend toward higher MAP in rats treated with diazepam, morphine, or pentobarbital vs isoflurane, these did not reach statistical significance.

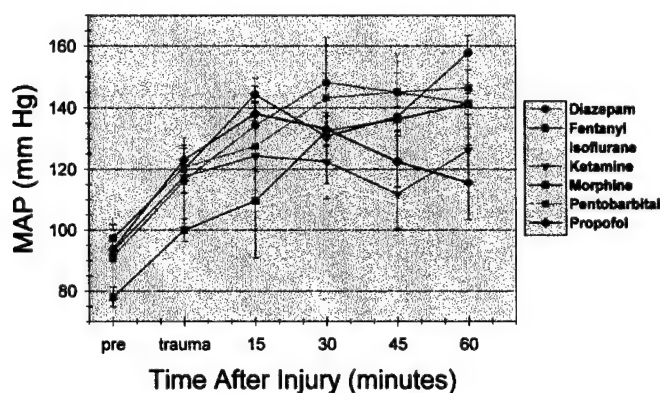


Fig 8: MAP vs time after injury. MAP is higher in fentanyl (red) vs isoflurane (yellow) anesthetized rats ( $p < 0.05$ ).

Motor function results showed that some agents had detrimental effects on selected outcome parameters. Rats anesthetized with morphine or propofol performed worse than sham on beam balance testing ( $p < 0.05$ , Fig 9). Beam balance performances were similar among all other experimental groups. On beam walking tasks, shams performed better than all other experimental groups ( $p < 0.05$ , Fig 10). The longest latencies (poorest performances)

were again seen with morphine or propofol anesthesia. Beam walking performances were similar for all other treatment groups. Although there were no significant differences between experimental groups in MWM testing with a submerged platform, rats treated with diazepam or fentanyl had a trend toward worse performance than shams ( $p < 0.10$ , Fig 11). Similarly, in the visible platform paradigm, rats treated with fentanyl

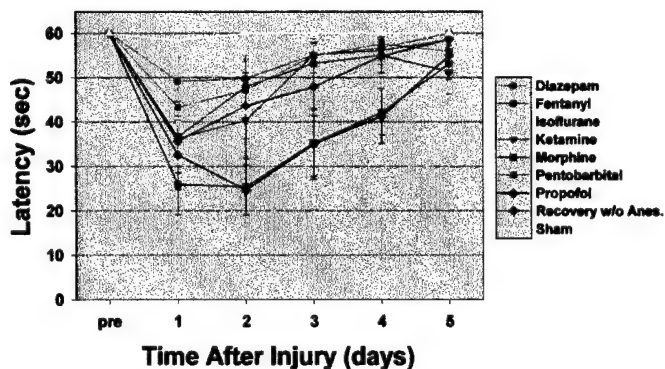


Fig 9: Beam balance latency vs time after injury. Rats treated with morphine (purple) or propofol (burgundy) performed worse than shams ( $p < 0.05$ ); however, there were no differences between other treatment groups.

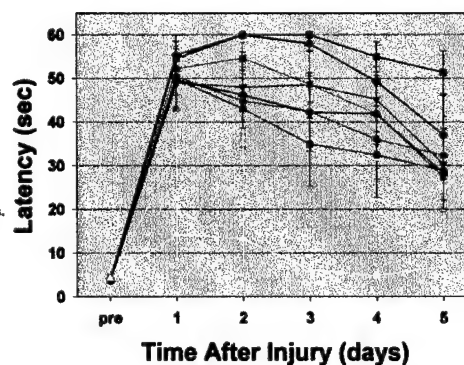


Fig 10: Beam walking latency vs time after injury. Sham rats performed better than all other experimental groups ( $p < 0.05$ ). There were no significant differences among treatment groups.

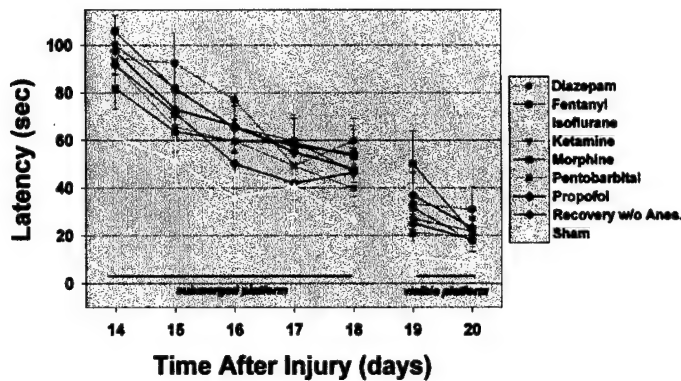


Fig 11: Latency to find platform vs time after injury in an acquisition paradigm of the MWM. With the submerged platform task, all treatment groups had similar performances; however, there was a trend toward worse performance in the diazepam or fentanyl treatment groups ( $p < 0.10$ ). Similarly, during visible platform testing, rats treated with fentanyl or morphine had a trend toward worse performance than sham ( $p < 0.10$ ). All other experimental groups had similar performances.

or morphine had a trend toward worse performance than sham ( $p < 0.10$ ); however, there were no significant differences between other experimental groups. Assessment of lesion volume and hippocampal cell counts is ongoing.

**Recommendation:** The results of this multi-sedation study show that rats anesthetized with narcotics, diazepam, or propofol had worse functional outcome than sham. **Surprisingly, rats treated with isoflurane only before injury performed as well as those given isoflurane both before injury and for 1 h post-**

trauma. Taken in context of our prior study (see C1) showing improved outcome with isoflurane vs fentanyl initiated *prior to injury*, these data suggest that powerful beneficial effects of isoflurane likely occur at or near the time of impact. Considering their common clinical use, the potential detrimental effects of treatment with narcotics, diazepam, or propofol deserve further characterization. Additionally, the use of isoflurane immediately before injury (despite the fact that rats were allowed to recover to tail pinch before injury) appears to mask the effects of the other sedative agents used in the study.

The dramatic influence of early anesthetic management after TBI in our model suggests important implications for the field management of TBI. Although isoflurane may not represent the most practical field agent, novel medications with similar anti-excitotoxic and/or cerebral blood flow promoting effects may well provide a breakthrough in the clinical management of TBI. Our study clearly demonstrates that the an optimal field anesthetic regimen after severe TBI needs to be defined. Clinical studies to evaluate the effects of different anesthetic strategies immediately after impact (i.e. those applied by a field medic) are needed. In addition, further animal studies are warranted to better define the optimal anesthetic properties early after TBI.

As in initial step, we are currently conducting studies to elucidate the potential neuroprotective mechanisms of isoflurane anesthesia by assessing cerebral glucose utilization, cerebral blood flow, and cerebral calcium accumulation during isoflurane vs fentanyl anesthesia in our CCI model of TBI. Finally, we are comparing anesthetic strategies in our CCI model using fentanyl (in lieu of isoflurane) as the preparatory anesthetic agent. We feel that this approach will more accurately simulate the clinical head injury, both in the military and civilian sectors.

## **(7) KEY RESEARCH ACCOMPLISHMENTS**

### **In order of importance**

#### **Sedation/Anesthesia**

- Narcotics, the standard field-treatment of victims of severe head injury (after intubation) and a front line treatment in emergency departments in the civilian sector had not been directly compared with general anesthesia in a contemporary rodent model of TBI. After experimental TBI, rats anesthetized at the time of injury with isoflurane vs fentanyl exhibited markedly better functional and histopathological outcomes. Additionally, it appears that powerful beneficial effects of isoflurane occur at or near the time of impact. Despite current clinical use, narcotics may not be the optimal sedative/analgesic early after TBI. Although isoflurane may not represent the most practical field agent, novel medications with similar anti-excitotoxic and/or cerebral blood flow promoting effects may well provide a breakthrough in the clinical management of TBI.

#### **Hyperventilation**

- Aggressive hyperventilation for 4-5 h early after TBI is associated with an exacerbation of hippocampal neuronal death in selectively vulnerable brain regions adjacent to the contusion site.

#### **Hypothermia**

- Transient, moderate hypothermia, effective after TBI alone in prior studies, was demonstrated to be ineffective after experimental TBI in rats subjected to TBI with a superimposed secondary insult (hypoxemia). This may be clinically important since hypoxemic patients have not been randomized in current clinical trials of hypothermia after TBI.

#### **Mechanisms**

- Moderate hypothermia reduces markers of injury (such as DNA damage) early after experimental TBI. However, sustained (12 h) of hypothermia was also surprisingly ineffective (on long-term outcome) after experimental TBI in rats. This suggests that although there are beneficial effects of hypothermia, there are potential side effects.

## **(8) REPORTABLE OUTCOMES**

### **Manuscripts<sup>s</sup>**

Forbes ML, Clark RSB, Dixon CE, Graham SH, Marion DW, DeKosky ST, Schiding JK, Kochanek PM: Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact. *Journal of Neurosurgery* 88:549-556, 1998.

Kochanek PM, Safar P, Marion DW, Tisherman SA, Clark RSB, DeKosky ST: Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock: From Mild Cooling To Suspended Animation In: Hypothermia in Trauma: Deliberate or Accidental. ITACCS Monograph, CE Smith and CM Grande, Editors, Baltimore, Maryland, 1997, pp 17-20.

Robertson CL, Clark RSB, Dixon CE, Graham ST, Alexander HL, Wisniewski SR, Marion DW, Safar PJ, Kochanek PM: No Long Term Benefit from Hypothermia after Severe Traumatic Brain Injury with Secondary Insult in Rats. *Critical Care Medicine* (in press).

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Graham SH, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome vs. fentanyl after traumatic brain injury in rats. *Journal of Neurotrauma* (in press).

<sup>§</sup>Full manuscripts from abstracts (see below) Whalen et al., and Ruppel et al. are also in preparation.

#### **Abstracts and Presentations**

Alexander HL, Robertson CL, Dixon CE, Clark RSB, Graham SH, Safar PJ, Kochanek PM: Vertical Versus Angled Controlled Cortical Impact in Rats. *Journal of Neurotrauma* 15:854, 1998.

Alexander H, Statler KD, Kochanek PM, Dixon CE, Clark RSB, Vagni V, Graham SH, Marion DW, Safar P: Effects of seven anesthetics on outcome in experimental traumatic brain injury. *Journal of Neurotrauma* (in press).

Clark RSB, Robertson CL, Dixon CE, Alexander HL, Graham SH, Safar PJ, Kochanek PM: Effect of Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Journal of Neurotrauma* 15:864, 1998.

Robertson CL, Clark R, Dixon CE, Graham S, Alexander H, Wisniewski S, Marion D, Safar P, Kochanek P: No Long-Term Benefit from Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Critical Care Medicine* 27(1):A52, 1999.

Ruppel RA, Kochanek PM, Dixon CE, Alexander HL, Graham SH, Clark RSB, Wisniewski SR, Marion DW, Safar PJ: MK-801 improves functional outcome in rats after controlled cortical impact. *Journal of Neurotrauma* 16:986, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Society for Neuroscience Abstract* 25:537, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.

## AWARDS

(1999) Women in Neurotrauma Award to Dr. Kimberly Statler for her presentation to the 1999 Meeting of the National Neurotrauma Society entitled, "Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats."

(2000) Educational Scholarship to Dr. Kimberly Statler from the Society of Critical Care Medicine for her abstract entitled, "Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats."

## (9) CONCLUSIONS

1. Based on the important finding in this study where the use of fentanyl in our CCI model produced deleterious effects on outcome after TBI, animal models should utilize clinically relevant sedative/analgesic treatments. The beneficial mechanisms of isoflurane (possibly promotion of cerebral blood flow or reduction of excitotoxicity) should be investigated for the development of novel treatments. Isoflurane anesthesia likely provides powerful beneficial effects at or near the time of injury. Sedative agents with anti-excitotoxic and/or cerebral blood flow promoting actions similar to isoflurane may thus represent better alternatives early after TBI. **Narcotics may not be the optimal sedative/analgesic early after TBI. Better field therapy than narcotics must be developed.**
2. Based on studies in rats using the CCI model, we have demonstrated tangible risk to aggressive, indiscriminate hyperventilation early after injury—specifically—augmentation of neuronal death in selectively vulnerable brain regions. This suggests that aggressive hyperventilation should not be indiscriminately used in the field



treatment of TBI, rather it should be applied if there are signs and/or symptoms of herniation. Mild hyperventilation (used in our control group) or normocapnia may be preferable.

3. Based on our studies in rats, hypothermia, although showing some beneficial effects, particularly early after TBI (such as a reduction in DNA damage, etc), may have some deleterious effects which result in only modest overall beneficial effects on long-term outcome. This is particularly true in the setting of severe injury (such as TBI plus secondary hypoxemic insults) where it is possible that there is little to gain except side effects. Also based on our narcotic (fentanyl) vs isoflurane study and our multi-sedation study, hypothermia should be re-examined in future studies with narcotic anesthesia, since beneficial effects of isoflurane may be masking any benefit from hypothermia.

## (10) REFERENCES

1. Forbes ML, Clark RSB, Dixon CE, Graham SH, Marion DW, DeKosky ST, Schiding JK, Kochanek PM: Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact. *Journal of Neurosurgery* 88:549-556, 1998.
2. Alexander HL, Robertson CL, Dixon CE, Clark RSB, Graham SH, Safar PJ, Kochanek PM: Vertical Versus Angled Controlled Cortical Impact in Rats. *Journal of Neurotrauma* 15:854, 1998.
3. Clark RSB, Robertson CL, Dixon CE, Alexander HL, Graham SH, Safar PJ, Kochanek PM: Effect of Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Journal of Neurotrauma* 15:864, 1998.
4. Robertson CL, Clark R, Dixon CE, Graham S, Alexander H, Wisniewski S, Marion D, Safar P, Kochanek P: No Long-Term Benefit from Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Critical Care Medicine* 27(1):A52, 1999.
5. Robertson CL, Clark RSB, Dixon CE, Graham ST, Alexander HL, Wisniewski SR, Marion DW, Safar PJ, Kochanek PM: No Long Term Benefit from Hypothermia after Severe Traumatic Brain Injury with Secondary Insult in Rats. *Critical Care Medicine* (in press).
6. Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.
7. Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.

8. Kochanek PM, Safar P, Marion DW, Tisherman SA, Clark RSB, DeKosky ST: Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock: From Mild Cooling To Suspended Animation In: Hypothermia in Trauma: Deliberate or Accidental. ITACCS Monograph, CE Smith and CM Grande, Editors, Baltimore, Maryland, 1997, pp 17-20.
9. Ruppel RA, Kochanek PM, Dixon CE, Alexander HL, Graham SH, Clark RSB, Wisniewski SR, Marion DW, Safar PJ: MK-801 improves functional outcome in rats after controlled cortical impact. *Journal of Neurotrauma* 16:986, 1999.
10. Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Society for Neuroscience Abstract* 25:537, 1999.
11. Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.
12. Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.
13. Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Graham SH, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome vs. fentanyl after traumatic brain injury in rats. *Journal of Neurotrauma* (in press).

## **(11) APPENDIX**

### **1. 1997 Report**

### **2. 1998 Report**

### **3. Curriculum Vitae**

### **4. Manuscripts:**

Forbes ML, Clark RSB, Dixon CE, Graham SH, Marion DW, DeKosky ST, Schiding JK, Kochanek PM: Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact. *Journal of Neurosurgery* 88:549-556, 1998.



Kochanek PM, Safar P, Marion DW, Tisherman SA, Clark RSB, DeKosky ST: Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock: From Mild Cooling To Suspended Animation In: Hypothermia in Trauma: Deliberate or Accidental. ITACCS Monograph, CE Smith and CM Grande, Editors, Baltimore, Maryland, 1997, pp 17-20.

Robertson CL, Clark RSB, Dixon CE, Graham ST, Alexander HL, Wisniewski SR, Marion DW, Safar PJ, Kochanek PM: No Long Term Benefit from Hypothermia after Severe Traumatic Brain Injury with Secondary Insult in Rats. *Critical Care Medicine* (in press).

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Graham SH, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome vs. fentanyl after traumatic brain injury in rats. *Journal of Neurotrauma* (in press).

#### **5. Abstracts:**

Alexander HL, Robertson CL, Dixon CE, Clark RSB, Graham SH, Safar PJ, Kochanek PM: Vertical Versus Angled Controlled Cortical Impact in Rats. *Journal of Neurotrauma* 15:854, 1998.

Clark RSB, Robertson CL, Dixon CE, Alexander HL, Graham SH, Safar PJ, Kochanek PM: Effect of Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Journal of Neurotrauma* 15:864, 1998.

Robertson CL, Clark R, Dixon CE, Graham S, Alexander H, Wisniewski S, Marion D, Safar P, Kochanek P: No Long-Term Benefit from Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Critical Care Medicine* 27(1):A52, 1999.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.

Ruppel RA, Kochanek PM, Dixon CE, Alexander HL, Graham SH, Clark RSB, Wisniewski SR, Marion DW, Safar PJ: MK-801 improves functional outcome in rats after controlled cortical impact. *Journal of Neurotrauma* 16:986, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Society for Neuroscience Abstract* 25:537, 1999

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.

Alexander H, Statler KD, Kochanek PM, Dixon CE, Clark RSB, Vagni V, Graham SH, Marion DW, Safar P: Post-traumatic application of seven different anesthetic agents in the controlled cortical impact model of traumatic brain injury: effect on outcome. *Journal of Neurotrauma* (in press).

**(12) BINDING (N/A)**

**(13) FINAL REPORTS**

**a) Bibliography of all publications and abstracts (see appendix)**

**b) List of personnel receiving pay from the research effort**

Patrick M. Kochanek, M.D.

Peter Safar, M.D.

Henry Alexander

Scott Heineman

Marci Provins

Linda Amick

## Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact

MICHAEL L. FORBES, M.D., ROBERT S. B. CLARK, M.D., C. EDWARD DIXON, PH.D., STEVEN H. GRAHAM, M.D., PH.D., DONALD W. MARION, M.D., STEVEN T. DEKOSKY, M.D., JOANNE K. SCHIDING, B.S., AND PATRICK M. KOCHANNEK, M.D.

*Safar Center for Resuscitation Research, The University of Pittsburgh Brain Trauma Research Center, and Departments of Anesthesiology/Critical Care Medicine, Pediatrics, Neurology, Neurosurgery, and Psychiatry, University of Pittsburgh, Pittsburgh, Pennsylvania*

Minimizing secondary injury after severe traumatic brain injury (TBI) is the primary goal of cerebral resuscitation. For more than two decades, hyperventilation has been one of the most often used strategies in the management of TBI. Laboratory and clinical studies, however, have verified a post-TBI state of reduced cerebral perfusion that may increase the brain's vulnerability to secondary injury. In addition, it has been suggested in a clinical study that hyperventilation may worsen outcome after TBI.

**Object.** Using the controlled cortical impact model in rats, the authors tested the hypothesis that aggressive hyperventilation applied immediately after TBI would worsen functional outcome, expand the contusion, and promote neuronal death in selectively vulnerable hippocampal neurons.

**Methods.** Twenty-six intubated, mechanically ventilated, isoflurane-anesthetized male Sprague-Dawley rats were subjected to controlled cortical impact (4 m/second, 2.5-mm depth of deformation) and randomized after 10 minutes to either hyperventilation ( $\text{PaCO}_2 = 20.3 \pm 0.7$  mm Hg) or normal ventilation groups ( $\text{PaCO}_2 = 34.9 \pm 0.3$  mm Hg) containing 13 rats apiece and were treated for 5 hours. Beam balance and Morris water maze (MWM) performance latencies were measured in eight rats from each group on Days 1 to 5 and 7 to 11, respectively, after controlled cortical impact. The rats were killed at 14 days postinjury, and serial coronal sections of their brains were studied for contusion volume and hippocampal neuron counting (CA1, CA3) by an observer who was blinded to their treatment group.

Mortality rates were similar in both groups (two of 13 in the normal ventilation compared with three of 13 in the hyperventilation group, not significant [NS]). There were no differences between the groups in mean arterial blood pressure, brain temperature, and serum glucose concentration. There were no differences between groups in performance latencies for both beam balance and MWM or contusion volume ( $27.8 \pm 5.1$  mm<sup>3</sup> compared with  $27.8 \pm 3.3$  mm<sup>3</sup>, NS) in the normal ventilation compared with the hyperventilation groups, respectively. In brain sections cut from the center of the contusion, hippocampal neuronal survival in the CA1 region was similar in both groups; however, hyperventilation reduced the number of surviving hippocampal CA3 neurons (29.7 cells/hpf, range 24.2–31.7 in the normal ventilation group compared with 19.9 cells/hpf, range 17–23.7 in the hyperventilation group [25th–75th percentiles]; \* $p < 0.05$ , Mann-Whitney rank-sum test).

**Conclusions.** Aggressive hyperventilation early after TBI augments CA3 hippocampal neuronal death; however, it did not impair functional outcome or expand the contusion. These data indicate that CA3 hippocampal neurons are selectively vulnerable to the effects of hyperventilation after TBI. Further studies delineating the mechanisms underlying these effects are needed, because the injudicious application of hyperventilation early after TBI may contribute to secondary neuronal injury.

**KEY WORDS** • head injury • hyperventilation • alkalosis • hippocampus • rat

**T**RAUMATIC brain injury (TBI) is often complicated by malignant intracranial hypertension,<sup>32</sup> which is associated with high mortality rates and has been managed using a combination of therapies including osmotherapy, diuretics, sedation, neuromuscular blockade, optimization of cerebral perfusion pressure, and hyperventilation.<sup>6,12,32,38,51</sup> Hyperventilation therapy has been an integral part of the clinical armamentarium in the management of severe TBI for more than 20 years:<sup>11</sup> this ther-

apy rapidly reduces cerebral blood flow (CBF) and cerebral blood volume in areas of the brain with intact CO<sub>2</sub> autoregulation, providing one option in the management of TBI complicated by malignant intracranial hypertension.<sup>1,34,42</sup>

In recent studies, however, researchers have defined a state of reduced CBF early after TBI in humans<sup>3,31</sup> and animals,<sup>5,20,25,46,56,57</sup> particularly in the first 8 hours after TBI. Some authors have hypothesized that the brain is more

vulnerable to secondary injury during this period and that additional reduction of CBF by hyperventilation may attenuate the delivery of important energy substrates.<sup>7,11,30,39,47,48</sup> Yoshida and Marmarou<sup>58</sup> reported that hyperventilation produced relative ischemia in cat brain after fluid-percussion injury and demonstrated an increase in brain lactate and inhibition of recovery of the ratio of phosphocreatine to inorganic phosphate. Muizelaar, et al.,<sup>40</sup> also demonstrated a loss of brain interstitial bicarbonate buffer after sustained prophylactic hyperventilation in rabbits. It has been reported that hyperventilation after TBI in animals and humans can reduce CBF to what traditionally have been considered ischemic levels.<sup>10,24,42</sup> However, defining the ischemic threshold in injured tissue is problematic.<sup>22,33</sup> Muizelaar, et al.,<sup>39</sup> reported that prolonged hyperventilation after TBI in humans may worsen functional outcome, raising questions regarding the appropriate indications and timing for the optimum application of hyperventilation after TBI. Recently published guidelines for the management of severe head injury<sup>6</sup> state that "in the absence of intracranial hypertension, hyperventilation ( $\text{PaCO}_2 \leq 35$  mm Hg) therapy should be avoided during the first 24 hours after severe TBI. . .," although "hyperventilation therapy may be necessary for brief periods where there is acute neurologic deterioration. . ." Consistent with these guidelines, in the setting of acute neurological deterioration, aggressive hyperventilation is used by both emergency and critical care personnel. In addition, in the initial stabilization of the brain-injured patient, aggressive hyperventilation (appropriate in the setting of impending herniation, or iatrogenic) occasionally occurs in both the prehospital and acute care settings. The specific impact of hyperventilation during this early low-flow period remains to be determined. Despite the availability of well-characterized rodent models of TBI, which reproduce the early posttraumatic reduction in CBF, the effect of aggressive hyperventilation on histopathological and functional outcome has not, to our knowledge, been investigated.

Using a rat model of focal percussive contusion, we hypothesized that aggressive hyperventilation, beginning immediately after TBI and continuing for 5 hours, would worsen functional outcome, expand the contusion, and promote neuronal death in selectively vulnerable hippocampal neurons.

## Materials and Methods

### Animals and Study Groups

All experimental protocols used in this report were approved by the Animal Care and Use Committee of the University of Pittsburgh. Twenty-six virus-free Sprague-Dawley rats weighing  $346 \pm 5$  g were studied. Food and water were continuously available in their home cages. After TBI the rats were randomly assigned to one of two groups of 13 animals, one receiving normal ventilation ( $\text{PaCO}_2 = 30\text{--}40$  mm Hg) and one receiving hyperventilation ( $\text{PaCO}_2 = 15\text{--}25$  mm Hg).

### Surgery and Brain Trauma Model

Anesthesia was induced using 4% isoflurane in  $\text{N}_2\text{O}/\text{O}_2$  (2:1). The rats were endotracheally intubated and mechanically ventilated. The isoflurane concentration was reduced to 2% followed by sterile surgical placement of a femoral arterial catheter for continuous mean arterial blood pressure (MABP) and arterial blood gas monitoring.

Intramuscular injections of penicillin (100,000 U) and gentamicin (10 mg/kg) were given to minimize the risk of infection. Pancuronium bromide was administered at dosages of 0.1 mg/kg/hour via the arterial line to control ventilation. The rats' core temperature was monitored using a rectal probe.

After stereotactically guided head positioning, an incision was made and the scalp was retracted, exposing the left parietal bone. A craniotomy was made using a high-speed dental drill aided by a binocular operating microscope. A burr hole was made 5 mm anterior and 2 mm lateral to the bregma in the left side of the skull and a temperature probe (0.009-in outer diameter) was inserted through the burr hole and 2 mm into the left parietal cortex. The bone flap was left in place and the isoflurane was reduced to 1% followed by a 30-minute equilibration period. The brain temperature was maintained at  $37 \pm 0.5^\circ\text{C}$ . Normal arterial blood gas levels were achieved in all rats and  $\text{PaO}_2$  was maintained at greater than 70 mm Hg.

The TBIs were produced using a controlled cortical impact device as recently described<sup>9,25</sup> with minor modifications. Fifteen minutes before controlled cortical impact, an arterial blood sample was obtained for measurement of arterial blood gas levels, glucose concentration, and hematocrit. The bone flap was then removed and a vertical controlled cortical impact (4 m/second impactor velocity, 2.5-mm deformation depth) was delivered onto the exposed dura overlying the left parietal cortex. The bone flap was replaced and sealed with dental cement and the scalp was sutured.

### Study Design

The study protocol was designed to mimic the aggressive use of hyperventilation (as opposed to normal ventilation) in the immediate posttrauma period in the prehospital as well as early hospital setting. Ten minutes after controlled cortical impact, rats were randomized to either the normal ventilation group (13 animals,  $\text{PaCO}_2$  range 30–40 mm Hg) or the hyperventilation group (13 animals,  $\text{PaCO}_2$  range 15–25 mm Hg). The ventilator was adjusted to maintain normocarbida or hypocarbida for 5 hours after controlled cortical impact. Arterial blood gas readings were obtained at 30 minutes post-controlled cortical impact, then hourly. The MABP was recorded every 30 minutes after controlled cortical impact. Brain and rectal temperatures were recorded every 15 minutes.

At 5 hours after controlled cortical impact, anesthesia was discontinued. Temperature probes and the femoral artery catheter were removed and the rat was weaned from mechanical ventilation in the course of 1 hour and underwent extubation. The time to extubation was recorded. After extubation, supplemental  $\text{O}_2$  was administered for 30 minutes. When it had fully recovered, the rat was returned to its cage with full access to food and water.

### Functional Outcome and Behavior Assessment

**Beam Balance.** Vestibulomotor function was tested using the beam balance test<sup>14</sup> in eight rats from each group. One hour before surgery, the rat was placed lengthwise on a 1.5-cm-wide beam suspended above the ground. The time the rat remained on the beam was recorded (up to 60 seconds). The rat was then removed from the beam and the procedure was repeated. Rats were considered trained when they remained on the beam for three consecutive periods of 60 seconds. Beam balance tests were also performed daily on Days 1 to 5 postinjury. Three trials were recorded and averaged each day for each rat.

**Morris Water Maze.** Cognitive function was tested in the same eight rats from each group using a standard variation of the Morris water maze (MWM) paradigm.<sup>15,35</sup> A pool 180 cm in diameter and 60 cm deep was painted black and filled with water to a depth of 28 cm. A clear Plexiglas platform 10 cm in diameter and 26 cm high (2 cm below the water surface) was used as the hidden goal platform. The pool was located in a  $2.5 \times 2.5$ -m room with numerous extra-maze cues (for example, posters, pipes, bookcase) that remained constant throughout the experiment. Testing started 7 days after controlled cortical impact to avoid confounding effects of motor deficits. The rats underwent four trials per day for 5 consecutive days to assess spatial memory performance. The rats started each trial once from each of the four possible start locations

# Augmented neuronal death following hyperventilation post-TBI

TABLE 1

Physiological values in two groups of rats treated with hyperventilation or normal ventilation after TBI\*

Value	Normal Ventilation		Hyperventilation	
	Baseline	Postrandomization	Baseline	Postrandomization
pH	7.39 ± 0.01	7.37 ± 0.01	7.38 ± 0.01	7.53 ± 0.01†
PaCO <sub>2</sub> (mm Hg)	36.7 ± 1.1	34.9 ± 0.3	37.2 ± 0.9	20.3 ± 0.7†
PaO <sub>2</sub> (mm Hg)	165 ± 6	167 ± 4	168 ± 4	180 ± 3†
base deficit (mmol/L)	2.7 ± 3.4	4.2 ± 0.7	-0.6 ± 0.9	4.8 ± 0.6
serum glucose (mg%)	189 ± 9	174 ± 6	158 ± 10	152 ± 9
hct (%)	36 ± 2.3	35 ± 0.6	32.3 ± 1.5	35 ± 0.6
time to extubate (min)	NA	28 ± 6	NA	29 ± 5
brain temperature (°C)	36.7 ± 0.1	37 ± 0	36.6 ± 0.1	37 ± 0
rectal temperature (°C)	36.5 ± 0.6	37 ± 0	37.1 ± 0.1	37.1 ± 0.1
MABP (mm Hg)	129 ± 4	123 ± 4	129 ± 8	128 ± 3

\* All values are expressed as mean ± SEM. Abbreviations: hct = hematocrit; NA = not applicable.

† p < 0.05 at 30 minutes postrandomization compared with baseline.

(north, south, east, and west); the order of the starting location was randomized. The goal platform was positioned 45 cm from the outside wall and was placed in either the northeast, southeast, southwest, or northwest quadrant of the maze. The location of the platform was kept constant for each rat. Rats were manually placed in the pool facing the wall and were given a maximum of 120 seconds to find the hidden platform. If the rats failed to find the platform within 120 seconds, they were placed there by the researcher. All rats were allowed to remain on the platform for 30 seconds before being placed in a heated incubator between trials. There was a 4-minute intertrial interval. All data were recorded by means of a video tracking system.

## Histopathological Studies

At 14 days after controlled cortical impact (after completion of all of the functional outcome testing), the rats were anesthetized with 5% isoflurane and killed by perfusion fixation using 10% buffered formalin. Their brains were removed and postfixed at 4°C for a minimum of 1 week, and then cryoprotected in sucrose and cut with a cryotome into 10-μ coronal sections at 1-mm increments from the occipital to the frontal lobe and stained with Cresyl violet.

**Contusion Volume.** We used a computerized image analysis system to outline the margin of the contusion and the sectional area of the contusion at each 1-mm increment was calculated by an observer (M.L.F.) who was blinded to the treatment group. Contusion volume in each rat was calculated as the sum of these sections.

**Hippocampal Cell Counting.** Neuronal loss in hippocampal regions CA1 and CA3 pyramidal layers was quantified.<sup>8</sup> A coronal section cut from the dorsal hippocampus underlying the area of contusion, approximately 2.6 mm posterior to the bregma, was used for analysis in each rat. The regions were visualized at × 100 magnification, then localized and counted at × 400 by an observer (R.S.B.C.) blinded to treatment group. Only complete cells with a clearly defined body and nucleus were counted. Surviving pyramidal CA1 and CA3 hippocampal neurons were counted in six separate × 400 fields for each region in both hemispheres. Sections were excluded if the boundary of the contusion extended into the pyramidal layers of the hippocampus or if fixation artifacts precluded accurate counting. Data are reported as the average number of surviving neurons per high-power field for the CA1 and CA3 hippocampal regions in both the ipsilateral and contralateral hemispheres.

## Statistical Analysis

Survival was compared between groups using Fisher's exact test. Between group comparisons of physiological parameters, beam balance, and MWM latencies were made using one- or two-way analysis of variance (ANOVA) for repeated measures where appropriate and post-hoc tests with appropriate correction for multiple compar-

isons. Contusion volume was normally distributed and was compared between groups using Student's t-test. Hippocampal neuronal survival in CA1 and CA3 was not normally distributed and was compared between groups using the Mann-Whitney rank-sum test. Significance was defined at a probability level of less than 0.05.

## Sources of Supplies and Equipment

Pancuronium bromide and gentamicin were purchased from Elkins-Sinn, Cherry Hill, NJ, and penicillin was acquired from Upjohn, Kalamazoo, MI. The stereotactic head positioning system was obtained from David Kopf, Tjunga, CA. The temperature probe was purchased from Physitemp Corp., Clifton, NJ. The video tracking system (Poly-Trak) was acquired from San Diego Instrument, Inc., San Diego, CA, and the image analysis system (MCID) was from Imaging Research, St. Catharines, Ontario, Canada.

## Results

### Physiological Parameters

Baseline and 30-minute postrandomization physiological data are presented for all measured parameters in Table 1. After randomization, there was a marked increase in pH and decrease in PaCO<sub>2</sub> in the hyperventilation group (compared with baseline, p < 0.05). Hyperventilation was also associated with a small increase (12 mm Hg) in PaO<sub>2</sub> compared with baseline (p < 0.05). This difference was attributable to the increased minute ventilation and mean airway pressure in the hyperventilation group. At no time were any of the rats hypoxemic (PaO<sub>2</sub> < 100 mm Hg). The entire time course of PaCO<sub>2</sub>, arterial pH, MABP, and brain temperature after TBI is given for both groups in Fig. 1. The PaCO<sub>2</sub> and pH levels differed between groups at all time points after randomization (p < 0.05). The MABP and brain temperature were similar in both groups.

Five of 26 rats died during the 14-day study, with all deaths occurring on the day of injury. Two rats remained unresponsive postinjury and were unable to demonstrate any spontaneous respiratory effort for 1 hour after discontinuation of anesthesia and were therefore killed. Three rats developed pulmonary edema and/or respiratory distress and died soon after extubation. There were no differences in mortality between groups (two of 13 in the normal ventilation group compared with three of 13 in the hyperventilation group). There were no differences between groups in time to extubation (Table 1).

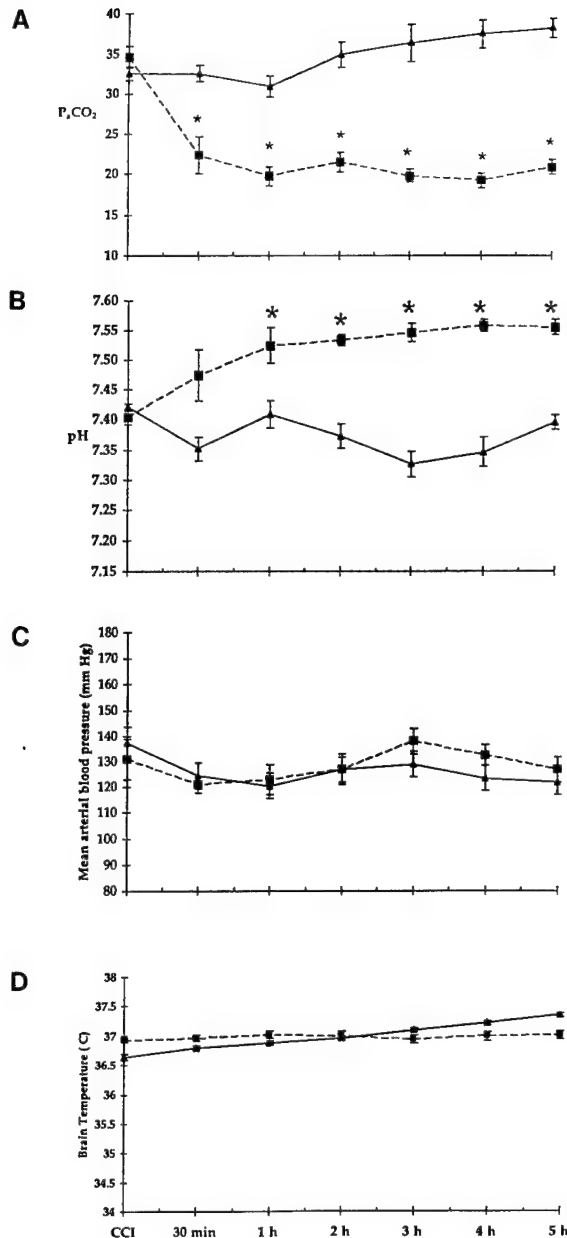


FIG. 1. Graphs showing time course of (A) PaCO<sub>2</sub> (mm Hg), (B) arterial pH, (C) MABP (mm Hg), and (D) brain temperature (°C) in all rats treated with either normal ventilation (triangles w/ solid line, 13 animals) or hyperventilation (squares w/ broken line, 13 animals) after controlled cortical impact. \* $p < 0.05$  for normal ventilation compared with hyperventilation. Data are expressed as the mean  $\pm$  standard error of the mean (SEM).

#### Functional Outcome Assessment

**Beam Balance.** There was no difference between groups in motor performance latencies over time ( $F_{1,15} = 0.17$ ,  $p < 0.69$ , Fig. 2). Maximum impairment of performance occurred on Days 1 or 2 in both groups, and eventually returned to baseline. Beam balance performance did not differ significantly between normal ventilation and hyperventilation groups.

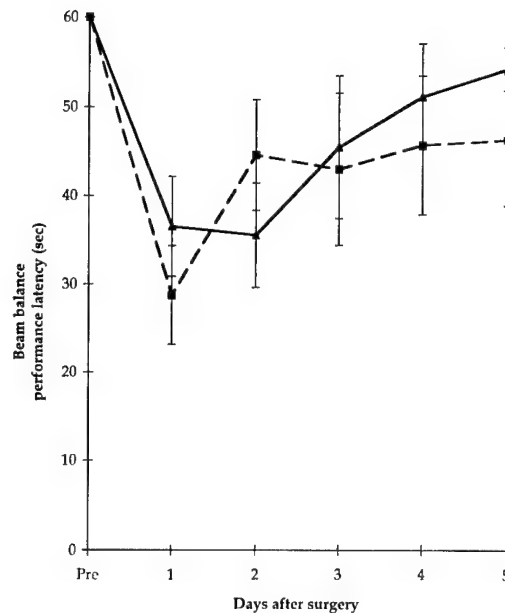


FIG. 2. Graph showing mean beam balance performance latencies (mean  $\pm$  SEM, in seconds) in rats before and on Days 1 to 5 after controlled cortical impact (4 m/second, 2.5-mm cortical deformation depth). Repeated-measures ANOVA revealed no difference in duration of balance maintained between the two groups (triangles = normal ventilation [eight rats]; squares = hyperventilation [eight rats]).

**Morris Water Maze.** There was no difference between normal ventilation and hyperventilation groups in the time needed to find the hidden platform in the MWM test ( $F_{1,15} = 0.50$ ,  $p < 0.50$ , Fig. 3). In addition, there was a statistically nonsignificant tendency ( $t_{13} = 1.77$ ,  $p < 0.065$ ) for the rats in the hyperventilation group to swim slower than the rats in the normal ventilation group ( $30.8 \pm 1.0$  compared with  $35.4 \pm 2.1$  cm/second).

#### Histopathological Studies

**Contusion Volume.** At the injury level selected for this study, the contusion was generally restricted to the parietal cortex beneath the impact site. Contusion volume in both groups is shown in Fig. 4. There was no difference between groups ( $27.8 \pm 5.1$  mm<sup>3</sup> in the normal ventilation group compared with  $27.8 \pm 3.3$  mm<sup>3</sup> in the hyperventilation group) in this outcome parameter.

**Hippocampal Cell Counting.** Figure 5 shows the number of surviving neurons/hpf in the CA1 and CA3 regions of the dorsal hippocampus ipsilateral to the contusion. There were no differences in the number of surviving CA1 hippocampal neurons between groups after controlled cortical impact. There was, however, a further reduction in the number of surviving CA3 neurons in the hyperventilation group after controlled cortical impact compared with the normal ventilation group (normal ventilation 29.7, range 24.2–31.7 neurons/hpf, compared with hyperventilation 19.9, range 17–23.7 neurons/hpf; median [25th–75th percentiles],  $p < 0.05$ ). Neuronal cell counts in the CA1 and CA3 regions of the hemisphere contralateral to the contusion did not differ in either the normal ventilation or hyperventilation groups (CA1 counts = 55.3, range 52.1–59



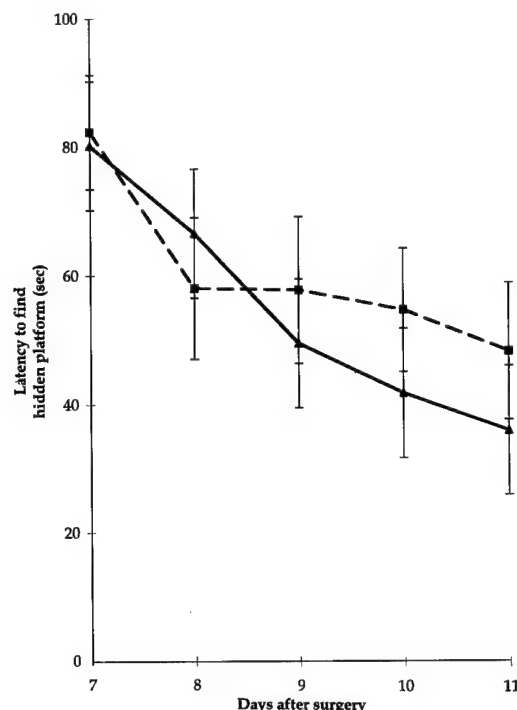


FIG. 3. Graph showing MWM performance latency to find a hidden platform (mean  $\pm$  SEM, in seconds) by rats on Days 7 to 11 after controlled cortical impact. There was no difference between groups (triangles = normal ventilation [eight animals]; squares = hyperventilation [eight animals]) when performances were compared using ANOVA with repeated measures.

[normal ventilation] and 57.3, range 51.3–59 [hyperventilation]; CA3 = 40, range 36.6–41.2 [normal ventilation] and 38, range 33–41.7 [hyperventilation]).

## Discussion

In a model of controlled cortical impact-induced focal contusion in rats, aggressive hyperventilation for 5 hours after TBI augments neuronal death in the CA3 region of the hippocampus ipsilateral to the contusion. However, hyperventilation did not worsen motor function or cognitive outcome, as assessed using standard beam balance and MWM paradigms, respectively, and did not increase contusion volume.

Hippocampal CA3 neurons are selectively vulnerable to delayed neuronal death after TBI.<sup>2,8,19,49,52,53</sup> Theories about the mechanisms underlying this process remain speculative. Potential mechanisms include ischemia, TBI-induced excitotoxicity, apoptosis, and inflammation.<sup>8,19,49</sup>

Yamakami and McIntosh<sup>56,57</sup> reported reduced CBF as early as 15 and 30 minutes after TBI. Using a piglet model of TBI, Pfenninger, et al.,<sup>46</sup> reported CBF reduction as early as 5 minutes post-TBI. Some flow levels were in the range consistent with ischemia. We have previously demonstrated that the hippocampus and cortex ipsilateral to the impact show marked flow reduction (at least 60%) at 2 hours after TBI in the controlled cortical impact model.<sup>25</sup> Cerebral blood flow approaches ischemic levels in the

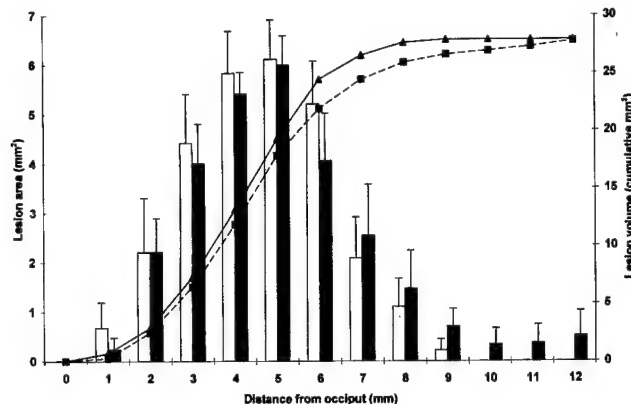


FIG. 4. Bar graph depicting mean lesion area (left y-axis, mm<sup>2</sup>) compared with distance from occiput (mm) measured 14 days after controlled cortical impact (open bars, normal ventilation [11 rats]; closed bars, hyperventilation [10 rats]). Contusion volume (mm<sup>3</sup>) was calculated as the sum of these areas in each group and is depicted as the cumulative volume (right y-axis) in the normal ventilation (triangles) and hyperventilation (squares) groups. There was no difference between groups in contusion volume (normal ventilation,  $27.8 \pm 5.1$  mm<sup>3</sup> compared with hyperventilation,  $27.8 \pm 3.1$  mm<sup>3</sup>, mean  $\pm$  SEM).

core of the contusion at 2 hours postinjury. Although we have not evaluated the reactivity status of the cerebral circulation to changes in PaCO<sub>2</sub> at 2 hours after TBI in this model, we have reported that CO<sub>2</sub> reactivity is impaired, although still present (62–71% of baseline) in and around the contusion at 24 hours after controlled cortical impact in rats.<sup>16</sup>

Hyperventilation rapidly reduces cerebral blood volume and intracranial pressure (ICP).<sup>11</sup> In some studies, this intervention has been associated with CBF values consistent with ischemia or brain tissue hypoxia.<sup>10,11,42,48</sup> After global cerebral ischemia in dogs, hyperventilation did not increase neuronal death;<sup>55</sup> however, the brains were assessed at 8 hours after reperfusion, and neuronal death may be delayed. Although ischemia may be considered a contributing mechanism in the observed augmented neuronal death, ischemia alone is an inadequate explanation for our findings in light of the preservation of CA1 neurons. Although CA1 neurons are known to be selectively vulnerable to ischemic injury,<sup>23</sup> they were not affected by hyperventilation in this paradigm. Furthermore, in our model, CA1 neurons are more proximal to the point of impact in the cortex compared with CA3 neurons. The lack of CA1 neuronal death in light of ischemic and (presumed) anatomical vulnerability weighs against ischemia and primary injury as putative mechanisms of neuronal death in the hippocampus in this model. One limitation in this study is that neuronal counting using traditional histological methods may underestimate cell loss because of a loss of hippocampal volume.<sup>52</sup> We did not use stereological methods in this study. However, CA1 neuronal counts did not differ between groups and were equivalent to those observed in sham-injured animals studied in our laboratory in prior published<sup>8</sup> and unpublished work. In addition, comparisons were only made between injured groups within this study.

Hyperventilation produces cerebral vasoconstriction

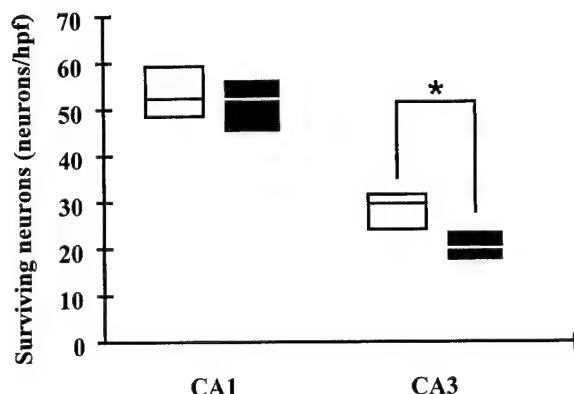


FIG. 5. Box plots representing the number of surviving CA1 and CA3 hippocampal neurons in coronal brain sections cut from the center of the lesion in the hemisphere ipsilateral to the contusion. Cells were counted 14 days postinjury. The median line is placed within the shaded 25th to 75th percentile range. There was a reduction in the number of surviving CA3 hippocampal neurons after injury in normal ventilation (open boxes) compared with hyperventilation (solid boxes) groups (29.7 cells/hpf, range 24.2–31.7 compared with 19.9 cells/hpf, range 17–23.7). \* $p < 0.05$ , Mann–Whitney rank-sum test.

and alkalosis.<sup>40</sup> Alkalosis exacerbates *N*-methyl-D-aspartate receptor-mediated neurotoxicity.<sup>17,18,21,43</sup> As a result of aggressive hyperventilation, the rats in our study were quite alkalotic as indicated by arterial pH measurements. Although we did not measure brain pH, a decrease in  $\text{PaCO}_2$  immediately reduces brain interstitial pH.<sup>40</sup> Although alkalosis appears to have deleterious effects on neurons, acidosis has been shown to have both beneficial and detrimental effects. Giffard, et al.,<sup>17</sup> and Takadera, et al.,<sup>54</sup> reported a neuroprotective effect of acidosis via an attenuation of the *N*-methyl-D-aspartate receptor activation in vitro. Rosner and Becker<sup>50</sup> reported a deleterious effect of tissue acidosis after experimental TBI in cats. The spatial distribution of brain pH around the contusion and in the hippocampus has not been determined for either normal ventilation or hyperventilation conditions in our model.

Finally, the potential effects of hyperventilation on other mechanisms such as posttraumatic seizures or axonal injury may contribute to the enhanced vulnerability of CA3 neurons. The lateralization of the deleterious effects also raises the possibility that spreading wave depression may be a component of the neurotoxic milieu after TBI in this model of focal contusion.<sup>20</sup> It could also be the case that the combined effect of alkalosis and further flow reduction by hyperventilation is deleterious in regions vulnerable to excitotoxicity such as CA3. Early, aggressive, or prophylactic hyperventilation, therefore, in the context of reduced CBF, may potentiate excitotoxic mechanisms and augment neuronal death.

Aggressive hyperventilation in the early low-flow period did not worsen functional outcome or expand the contusion, failing to support a significant portion of our initial hypothesis. Ultimate contusion size, in controlled cortical impact or other models of focal contusion, is relatively refractory to manipulation by a variety of interventions;<sup>4,8</sup>

however, application of hypothermia, particularly prior to injury, reduces contusion volume resulting from controlled cortical impact and lateral fluid-percussion injury.<sup>13,44</sup> Although we chose rather aggressive hyperventilation in an attempt to produce a maximum effect, we did not test the effect of hyperventilation on a milder contusion, which may be more manipulable to secondary insults. The contusion penumbra has not been clearly defined in either of the standard rodent TBI models (controlled cortical impact or fluid-percussion) for any level of injury. It is possible that selectively vulnerable CA3 hippocampal neurons are the only potential target for a deleterious effect of hyperventilation in our model. However, the effect of hyperventilation on the survival of neurons in the dentate gyrus or hilus (all vulnerable to TBI)<sup>9,29</sup> was not assessed.

Hippocampal damage and memory deficits are common after TBI in humans.<sup>26,28</sup> This study did not reveal any added effect of hyperventilation on functional outcome deficits as measured by beam balance and MWM latencies. A number of factors may have contributed to this. Our sample size may have limited statistical power; however, this sample size was adequate to detect the exacerbation of functional deficits by the addition of 30 minutes of moderate hypoxemia ( $\text{PaO}_2$  40 mm Hg) in our model.<sup>8</sup> Second, the cognitive deficits in this model are modest compared with those detailed in previous reports.<sup>15</sup> Bilateral hippocampal damage may be necessary to create more marked functional deficits.<sup>36,37</sup> In addition, CA3 damage may not mediate post-TBI memory deficits, as manifested in MWM test results. Finally, the specific functional outcome paradigm may not have the necessary sensitivity to detect subtle functional deficits. For example, more demanding MWM paradigms have been used by other investigators.<sup>27,52</sup> However, in support of the testing strategy used, our hypothesis was that hyperventilation would worsen functional deficits.

This study does not completely address the uncommon situation in which, soon after severe head injury, marked intracranial hypertension is observed. Hyperventilation may in fact be life saving in its ability to impede herniation. Similarly, we did not measure ICP or titrate ventilation to control cerebral perfusion pressure, and we evaluated only one level of hyperventilation and injury severity. We did not attempt to model the clinical scenario of optimum titration of ventilation when ICP is increased. In the clinical setting, some investigators have demonstrated a wide variety of beneficial effects of hyperventilation under those conditions, such as homogenization of CBF, normalization of cerebral glucose uptake, and improvement in autoregulation.<sup>12,41,42</sup> Rather, we chose the worst-case scenario, aggressive hyperventilation during the early posttrauma period when flow is already low and excitotoxicity is peaking.<sup>45</sup> However, our study does show that hyperventilation is associated with a tangible risk to vulnerable neurons in the controlled cortical impact model. To our knowledge, this is the first in vivo study demonstrating that hyperventilation can augment neuronal injury after TBI, suggesting that there is indeed a tradeoff associated with this intervention.

## Conclusions

We have demonstrated that aggressive, early hyperven-



tilation after TBI augments neuronal death in CA3 hippocampus. The further reduction of CBF with hyperventilation during the low CBF state immediately after severe TBI, coupled with alkalosis, may increase the vulnerability of selected neurons to traumatic injury. Further studies are needed to delineate the relative contributions of these mechanisms to the observed effects. The results of this study reinforce that meticulous attention is necessary to prevent secondary injury after TBI, and a risk in the use of hyperventilation is demonstrated.

## Acknowledgments

The authors thank Drs. Peter Safar and Walter Obrist for helpful comments. We also thank Raymond Griffith, Erica Baum, and Xiecheng Ma for technical assistance; Linda Amick and Marci Provins for preparing the manuscript; and Francie Siegfried for editorial assistance.

## References

- Artru AA: Reduction of cerebrospinal fluid pressure by hypocapnia: changes in cerebral blood volume, cerebrospinal fluid volume, and brain tissue water and electrolytes. *J Cereb Blood Flow Metab* 7:471-479, 1987
- Baldwin SA, Gibson T, Callihan TC, et al: Neuronal cell loss in the CA3 subfield of the hippocampus following cortical contusion utilizing the optical disector [sic] method for cell counting. *J Neurotrauma* 14:385-398, 1997
- Bouma GJ, Muizelaar JP, Stringer WA, et al: Ultra-early evaluation of regional cerebral blood flow in severely head-injured patients using xenon-enhanced computerized tomography. *J Neurosurg* 77:360-368, 1992
- Bramlett HM, Dietrich WD, Green EJ, et al: Chronic histopathological consequences of fluid-percussion brain injury in rats: effects of post-traumatic hypothermia. *Acta Neuropathol* 93:190-199, 1997
- Bryan RM Jr, Cherian L, Robertson C: Regional cerebral blood flow after controlled cortical impact injury in rats. *Anesth Analg* 80:687-695, 1995
- Bullock R, Chesnut RM, Clifton G, et al: Guidelines for the management of severe head injury. *J Neurotrauma* 13:699-703, 1996
- Chesnut RM, Marshall LF, Klauber MR, et al: The role of secondary brain injury in determining outcome from severe head injury. *J Trauma* 34:216-222, 1993
- Clark RS, Kochanek PM, Dixon CE, et al: Early neuropathologic effects of mild or moderate hypoxemia after controlled cortical impact in rats. *J Neurotrauma* 14:179-189, 1997
- Clark RSB, Kochanek PM, Marion DW, et al: Mild posttraumatic hypothermia reduces mortality after severe controlled cortical impact in rats. *J Cereb Blood Flow Metab* 16:253-261, 1996
- Cold GE: Does acute hyperventilation provoke cerebral oligemia in comatose patients after acute head injury? *Acta Neurochir* 96:100-106, 1989
- Crockard HA, Coppel DL, Morrow WFK: Evaluation of hyperventilation in treatment of head injuries. *Br Med J* 4:634-640, 1973
- Cruz J: An additional therapeutic effect of adequate hyperventilation in severe acute brain trauma: normalization of cerebral glucose uptake. *J Neurosurg* 82:379-385, 1995
- Dietrich WD, Alonso O, Busto R, et al: Post-traumatic brain hypothermia reduces histopathological damage following concussive brain injury in the rat. *Acta Neuropathol* 87:250-258, 1994
- Dixon CE, Lyeth BG, Povlishock JT, et al: A fluid percussion model of experimental brain injury in the rat. *J Neurosurg* 67:110-119, 1987
- Dixon CE, Ma X, Marion DW: Effects of CDP-choline treatment on neurobehavioral deficits after TBI and on hippocampal and neocortical acetylcholine release. *J Neurotrauma* 14:161-169, 1997
- Forbes ML, Hendrich KS, Kochanek PM, et al: Assessment of cerebral blood flow and CO<sub>2</sub> reactivity after controlled cortical impact by perfusion magnetic resonance imaging using arterial spin-labeling in rats. *J Cereb Blood Flow Metab* 17:865-874, 1997
- Giffard RG, Monyer H, Christine CW, et al: Acidosis reduces NMDA receptor activation, glutamate neurotoxicity and oxygen-glucose deprivation neuronal injury in cortical cultures. *Brain Res* 506:339-342, 1990
- Giffard RG, Weiss JH, Choi DW: Extracellular alkalinity exacerbates injury of cultured cortical neurons. *Stroke* 23:1817-1821, 1992
- Hicks RR, Smith DH, Lowenstein DH, et al: Mild experimental brain injury in the rat induces cognitive deficits associated with regional neuronal loss in the hippocampus. *J Neurotrauma* 10:405-414, 1993
- Hovda DA, Lee SM, Smith ML, et al: The neurochemical and metabolic cascade following brain injury: moving from animals to man. *J Neurotrauma* 12:903-906, 1995
- Hurn PD, Kochler RC, Traystman RJ: Extracellular alkalosis reduces recovery from global cerebral ischemia. *J Cereb Blood Flow Metab* 15 (Suppl 1):S201, 1995 (Abstract)
- Jacewicz M, Brint S, Tanabe J, et al: Nimodipine pretreatment improves cerebral blood flow and reduces brain edema in conscious rats subjected to focal cerebral ischemia. *J Cereb Blood Flow Metab* 10:903-913, 1990
- Jenkins LW, Moszynski K, Lyeth BG, et al: Increased vulnerability of the mildly traumatized rat brain to cerebral ischemia: the use of controlled secondary ischemia as a research tool to identify common or different mechanisms contributing to mechanical and ischemic brain injury. *Brain Res* 477:211-224, 1989
- Kennealy JA, McLennan JE, Loudon RG, et al: Hyperventilation-induced cerebral hypoxia. *Am Rev Resp Dis* 122:407-412, 1980
- Kochanek PM, Marion DW, Zhang W, et al: Severe controlled cortical impact in rats: assessment of cerebral edema, blood flow, and contusion volume. *J Neurotrauma* 12:1015-1025, 1995
- Kotapka MJ, Graham DI, Adams JH, et al: Hippocampal pathology in fatal non-missile human head injury. *Acta Neuropathol* 83:530-534, 1992
- Kraemer PJ, Brown RW, Baldwin SA, et al: Validation of a single-day Morris Water Maze procedure used to assess cognitive deficits associated with brain damage. *Brain Res Bull* 39:17-22, 1996
- Levin HS, Gary HE Jr, Eisenberg HM, et al: Neurobehavioral outcome 1 year after severe head injury. Experience of the Traumatic Coma Data Bank. *J Neurosurg* 73:699-709, 1990
- Lowenstein DH, Thomas MJ, Smith DH, et al: Selective vulnerability of dentate hilar neurons following traumatic brain injury: a potential mechanistic link between head trauma and disorders of the hippocampus. *J Neurosci* 12:4846-4853, 1992
- Madsen FF: Changes in regional cerebral blood flow after hyperventilation in the pig with an induced focal cerebral contusion. *Acta Neurochir* 106:164-169, 1990
- Marion DW, Darby J, Yonas H: Acute regional cerebral blood flow changes caused by severe head injuries. *J Neurosurg* 74:407-414, 1991
- Marshall LF, Smith RW, Shapiro HM: The outcome with aggressive treatment in severe head injuries. Part I. The significance of intracranial pressure monitoring. *J Neurosurg* 50:20-25, 1979
- Meis G, Ishimaru S, Xie Y, et al: Ischemic thresholds of cerebral protein synthesis and energy state following middle cere-

- bral artery occlusion in rat. **J Cereb Blood Flow Metab** 11: 753-761, 1991
34. Miller JD, Butterworth JF, Gudeman SK, et al: Further experience in the management of severe head injury. **J Neurosurg** 54:289-299, 1981
  35. Morris RGM, Garraud P, Rawlins JNP, et al: Place navigation impaired in rats with hippocampal lesions. **Nature** 297: 681-683, 1982
  36. Morris RGM, Hagan JJ, Rawlins JNP: Allocentric spatial learning by hippocampectomized rats: a further test of the "spatial mapping" and "working memory" theories of hippocampal function. **Q J Exp Psychol** 38B:365-395, 1986
  37. Moser E, Moser MB, Andersen P: Spatial learning impairment parallels the magnitude of dorsal hippocampal lesions, but is hardly present following ventral lesions. **J Neurosci** 13: 3916-3925, 1993
  38. Muizelaar JP, Marmarou A, DeSalles AAF, et al: Cerebral blood flow and metabolism in severely head-injured children. Part I: Relationship with GCS score, outcome, ICP and PVI. **J Neurosurg** 71:63-71, 1989
  39. Muizelaar JP, Marmarou A, Ward JD, et al: Adverse effects of prolonged hyperventilation in patients with severe head injury: a randomized clinical trial. **J Neurosurg** 75:731-739, 1991
  40. Muizelaar JP, van der Poel HG, Li Z: Pial arteriolar vessel diameter and CO<sub>2</sub> reactivity during prolonged hyperventilation in the rabbit. **J Neurosurg** 69:923-927, 1988
  41. Newell DW, Weber JP, Watson R, et al: Effect of transient moderate hyperventilation on dynamic cerebral autoregulation after severe head injury. **Neurosurgery** 39:35-44, 1996
  42. Obrist WD, Langfitt TW, Jaggi JL, et al: Cerebral blood flow and metabolism in comatose patients with acute head injury. Relationship to intracranial hypertension. **J Neurosurg** 61: 241-253, 1984
  43. Ou-Yang Y, Kristián T, Mellergård P, et al: The influence of pH on glutamate- and depolarization-induced increases of intracellular calcium concentration in cortical neurons in primary culture. **Brain Res** 646:65-72, 1994
  44. Palmer AM, Marion DW, Botscheller ML, et al: Therapeutic hypothermia is cytoprotective without attenuating the traumatic brain injury-induced elevations in interstitial concentrations of aspartate and glutamate. **J Neurotrauma** 10:363-372, 1993
  45. Palmer AM, Marion DW, Botscheller ML, et al: Traumatic brain injury-induced excitotoxicity assessed in controlled cortical impact model. **J Neurochem** 61:2015-2024, 1993
  46. Pfenninger EG, Reith A, Breitig D, et al: Early changes of intracranial pressure, perfusion pressure, and blood flow after acute head injury. Part I: An experimental study of the underlying pathophysiology. **J Neurosurg** 70:774-779, 1989
  47. Raichle ME, Plum F: Hyperventilation and cerebral blood flow. **Stroke** 3:566-575, 1972
  48. Raichle ME, Posner JB, Posner F: Cerebral blood flow during and after hyperventilation. **Arch Neurol** 23:394-403, 1970
  49. Rink A, Fung KM, Trojanowski JQ, et al: Evidence of apoptotic cell death after experimental traumatic brain injury in the rat. **Am J Pathol** 147:1575-1583, 1995
  50. Rosner MJ, Becker DP: Experimental brain injury: successful therapy with the weak base, tromethamine. With an overview of CNS acidosis. **J Neurosurg** 60:961-971, 1984
  51. Rosner MJ, Rosner SD, Johnson AH: Cerebral perfusion pressure: management protocol and clinical results. **J Neurosurg** 83:949-962, 1995
  52. Smith DH, Soares HD, Pierce JS, et al: A model of parasagittal controlled cortical impact in the mouse: cognitive and histopathologic effects. **J Neurotrauma** 12:169-178, 1995
  53. Soares HD, Sinson GP, McIntosh TK: Fetal hippocampal transplants attenuate CA3 pyramidal cell death resulting from fluid percussion brain injury in the rat. **J Neurotrauma** 12: 1059-1067, 1995
  54. Takadera T, Shimada Y, Mohri T: Extracellular pH modulates N-methyl-D-aspartate receptor-mediated neurotoxicity and calcium accumulation in rat cortical cultures. **Brain Res** 572: 126-131, 1990
  55. Vanicky I, Maršala M, Murár J, et al: Prolonged postischemic hyperventilation reduces acute neuronal damage after 15 min of cardiac arrest in the dog. **Neurosci Lett** 135:167-170, 1992
  56. Yamakami I, McIntosh TK: Alterations in regional cerebral blood flow following brain injury in the rat. **J Cereb Blood Flow Metab** 11:655-660, 1991
  57. Yamakami I, McIntosh TK: Effects of traumatic brain injury on regional cerebral blood flow in rats as measured with radiolabeled microspheres. **J Cereb Blood Flow Metab** 9:117-124, 1989
  58. Yoshida K, Marmarou A: Effects of tromethamine and hyperventilation on brain injury in the cat. **J Neurosurg** 74:87-96, 1991

Manuscript received June 9, 1997.

Accepted in final form October 23, 1997.

This study was funded by the following organizations: US Army Medical Research Acquisition Activity No. DAMD17-97-1-7009 (P.K.); the Laerdal Foundation (P.K.); National Institutes of Health Grant Nos. NS30318 (D.M.) and NS3198 (C.E.D.); Children's Hospital of Pittsburgh Seed Grant (M.F.); and Department of Veterans Medical Affairs Merit Review Program (S.G.).

Address reprint requests to: Patrick M. Kochanek, M.D., Safar Center for Resuscitation Research, 3434 Fifth Avenue, Pittsburgh, Pennsylvania 15260.

**Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock:  
From Mild Cooling to Suspended Animation**

<sup>1,2,3</sup>Patrick M. Kochanek, M.D., <sup>1,3</sup>Peter Safar, M.D., <sup>1,2,4</sup>Donald W. Marion, M.D.,  
<sup>1,3,5</sup>Samuel A. Tisherman, M.D., and <sup>1,2,6</sup>Steven T. DeKosky, M.D.

<sup>1</sup>*Safar Center for Resuscitation Research, <sup>2</sup>University of Pittsburgh Brain Trauma  
Research Center, and Departments of <sup>3</sup>Anesthesiology and Critical Care Medicine,  
<sup>4</sup>Neurosurgery, <sup>5</sup>Surgery, and <sup>6</sup>Psychiatry, University of Pittsburgh School of Medicine,  
Pittsburgh, PA*

**Correspondence to:**

Patrick M. Kochanek, M.D.

Director, Safar Center for Resuscitation Research

3434 Fifth Avenue

Pittsburgh, PA

15260

Telephone: (412) 383-1900

Fax: (412) 624-0943

email: [kochanek@smtp.anes.upmc.edu](mailto:kochanek@smtp.anes.upmc.edu)

## Objectives:

1. To familiarize the reader with contemporary studies on the application of resuscitative hypothermia in the treatment of traumatic brain injury and hemorrhagic shock.
2. To describe the potential mechanisms for the beneficial effects of hypothermia in these settings.
3. To present some recent findings from both laboratory and clinical studies of resuscitative hypothermia conducted at the University of Pittsburgh.
4. To discuss possible side effects and limitations of the application of therapeutic hypothermia.
5. To discuss future directions for novel applications of hypothermia in combination with pharmacologic interventions.

## Historical Perspective

One of the earliest reports of the potential beneficial effects of hypothermia in the treatment of traumatic brain injury was described by Charles Phelps in 1897 in his classic textbook *Traumatic injuries of the brain and its membranes* (1). It is fitting that this monograph was assembled on the 100th anniversary of this remarkable description.

*"The shaving of the head, which had been advised as a means of facilitating diagnosis, is at the same time a measure of treatment...The essential advantage ... to be derived from this procedure is that it permits the effective application of the ice-cap, which next to trephination, ... is most nearly a directly curative*

*resource... It is contraindicated in hemorrhages and cerebral lacerations when uncomplicated by serious contusion; but, as those lesions are constantly thus complicated, it may be held a proper resort when such symptoms are manifest, without regard to exact diagnosis."*

In the early 1940s, Fay (2,3) examined the deliberate application of hypothermia in traumatic brain injury, and this was followed by several additional series of case reports and uncontrolled trials between 1943 and 1979 by other pioneers in this field including Woringer et al (4), Sedzimir (5), Lazorhes and Campan (6), and Rosomoff (7) in traumatic brain injury, Albin et al (8) in spinal cord injury, Bigelow et al (9) and Swan et al (10) in cardiothoracic surgery, Rosomoff et al (11) in focal cerebral ischemia, Siebke et al (12) and Conn et al (13) in near drowning, Wolfe (14), Benson et al (15), Ravitch and Safar (16) in cardiopulmonary arrest, and Rush et al (17) in the application of deep hypothermia for total circulatory arrest. Although remarkable effects were suggested in many of these reports, they failed to demonstrate convincingly that hypothermia was beneficial and did not result in the widespread application of resuscitative hypothermia. These reports were complicated by a number of difficulties including variation in depth and duration of hypothermia, and failure to include concurrent normothermic controls. In addition, reports of potential infectious complications in patients treated with the sustained application of moderate hypothermia (18) tempered enthusiasm for further studying resuscitative hypothermia in a controlled fashion.

### **Laboratory studies supporting the application of therapeutic hypothermia in traumatic brain injury and hemorrhagic shock**

In the mid 1980s there was renewed interest in the laboratory investigation of the deliberate application of therapeutic hypothermia for protection (induced before the insult) or resuscitation (induced after the insult). This work was focused predominantly in models of global cerebral ischemia in rats and monkeys (19-23), cardiopulmonary arrest (24-28) and near drowning in dogs (29). Central to this resurgence in interest in hypothermia was the development of three novel concepts: 1) that remarkably mild hypothermia (a temperature reduction of between 3° and 5°C) was effective in reducing secondary brain damage (19-30), 2) that the duration of mild hypothermia necessary for a beneficial effect might be transient --as short as 1 or 2 hours (19,28) and 3) that brain temperature, not body temperature, was the critical therapeutic target (19). The chance discovery of the efficacy of mild, transient hypothermia in these studies revived the importance of hypothermia research because mild and transient hypothermia are safer and easier to induce than the previously tried moderate, sustained hypothermia. It is important to define the approximate temperature ranges commonly used to describe specific depths of therapeutic hypothermia. Generally accepted definitions of these ranges are mild (34° to 36°C), moderate (28° to 32°C), deep (15° to 25°C), and profound (< 15°C) hypothermia (31).

Specific investigation of the application of therapeutic hypothermia in the treatment of traumatic brain injury was renewed by the report of Clifton et al (32) who observed an

inverse correlation between functional outcome and brain temperature (between 30° and 40°C). This was followed by a series of reports from several laboratories further defining the beneficial effect of hypothermia in a wide variety of models (both rodent and canine) of traumatic brain injury (33-37).

Recent controlled laboratory studies of the utility of resuscitative hypothermia in models of hemorrhagic shock developed from the initial work of Crippen et al in our center (38) and of Meyer and Horton (39). This resuscitative effect was demonstrated in models of both controlled (38,40) and uncontrolled (41) hemorrhagic shock (Figure 1), and with both mild and moderate hypothermia (42,43).

In controlled laboratory studies addressing an additional hemorrhagic shock-related application of deliberate hypothermia, Tisherman et al (44,45) investigated the application of deep and profound hypothermic circulatory arrest to enable resuscitative surgery that would otherwise be impossible. Our series of studies into "suspended animation" has culminated so far in the study by Capone et al (46) who reported complete recovery of the brain in dogs after normothermic hemorrhagic shock of 1 hour followed by profound hypothermic circulatory arrest of 1 hour. This application of resuscitative hypothermia is being further developed as a possible novel therapeutic approach to the management of pulseless battlefield casualties, specifically, "suspended animation" for transport and repair of otherwise lethal extracranial wounds. "Suspended animation" could be induced and reversed by portable cardiopulmonary bypass (47) and followed by subsequent delayed resuscitation (48).

### **Why hypothermia: Proposed mechanisms for the beneficial effects of deliberate hypothermia in traumatic brain injury and hemorrhagic shock**

Laboratory and clinical trials in cerebral resuscitation from ischemic or traumatic brain injury have repeatedly highlighted the tremendous challenge involved in demonstrating reproducible efficacy, in a wide variety of injury models or injury types, when a single therapeutic agent is used (49-51). The complex, multifactorial nature of the cascades of secondary damage purported to occur in both ischemic and traumatic brain injury strongly suggests the need for multimodal therapies (23,48-52). A similar multifactorial pathogenesis is proposed in the evolution of visceral damage after hemorrhagic shock (31). A great deal of evidence suggests that hypothermia favorably and simultaneously influences a large number of secondary injury mechanisms including; energy failure (53), oxidant injury (54,55), delayed neuronal death (19,36), excitotoxicity (56), intracranial hypertension (37) edema formation (35,57), cytoskeletal protein degradation (58), blood-brain barrier permeability (59), IL-1 $\beta$  production (60) (Figure 2), and neutrophil accumulation (61). It is very likely that some critical combination of beneficial effects on these mechanisms is responsible for the success of therapeutic hypothermia in experimental and clinical trials.

### **Clinical investigation of therapeutic hypothermia in traumatic brain injury**

Although there is a much larger body of laboratory data supporting the use of mild, transient, resuscitative hypothermia in ischemic rather than traumatic brain injury,



clinical application of deliberate hypothermia has been spearheaded in controlled trials after traumatic brain injury. Uncontrolled trials of moderate hypothermia in patients after traumatic brain injury looked promising (57,62) but were abandoned because of management problems. Marion et al (63) reported a beneficial effect of moderate (32°C), transient (24 hours) hypothermia on intracranial hypertension in adults with severe closed head injury. A reduction in the need for other therapies for control of intracranial hypertension was observed. Clifton et al (64) reported a reduction in the incidence of posttraumatic seizures in adults treated with moderate hypothermia for 48 hours after severe head injury. A trend toward improved outcome was also observed. Similarly, Shiozaki et al (65) reported efficacy of mild hypothermia in controlling refractory intracranial hypertension in patients with severe traumatic brain injury. Most recently, Marion et al (66) demonstrated that moderate (32°C), transient (24 hours) hypothermia improved functional outcome as measured with the Glasgow outcome scale at 6 months after severe traumatic brain injury in 82 patients randomized to either hypothermia or normothermia. This beneficial effect extended to 12 months in the subgroup of patients with admission Glasgow coma score of 5 to 7 (Table 1). In addition, reductions in IL-1 $\beta$  and glutamate concentrations were demonstrated in cerebrospinal fluid samples from hypothermic vs normothermic patients, suggesting the possibility of beneficial effects of hypothermia on posttraumatic inflammation and excitotoxicity, respectively. Remarkably, a significant reduction in cerebral metabolic rate for oxygen was not observed (63,66), suggesting that this beneficial effect was not

due to a simple reduction in cerebral oxidative metabolic demands. A multicenter randomized controlled clinical trial of 48 hours of hypothermia vs normothermia in the treatment of human head injury is currently underway.

### **Potential limitations and complications of the application of deliberate hypothermia**

Hypothermia is associated with potentially limiting side effects. Suppression of acute inflammation (67) and an increased infection risk (18) are concerns. These complications appear to be importantly related to the duration of hypothermia and the underlying condition that is being treated. In traumatic brain injury, Marion et al (66) and Clifton et al (64) did not observe increases in the incidence of infection with 24 hour and 48 hour applications of hypothermia, respectively. However, longer applications of hypothermia may have considerable risk (18). In addition, application of mild hypothermia in settings not associated with ischemia but associated with considerable infection risk (such as elective abdominal surgery in patients with malignancies) increases infection rates (68).

Coagulopathy is suggested as another potential complication of hypothermia. However, in the studies of severely head injured patients by Marion et al (56), platelet counts and prothrombin times did not differ significantly between groups, and no difference in posttrauma intracranial hematomas or other hemorrhagic complications were noted despite the fact that some of the patients had multiple trauma. Cardiac arrhythmias were also not observed. The threshold for these complications appears to be

temperatures below 30°C (69,70). On the other hand, a recent report (71) suggested that morbid cardiac events after non-cardiac surgery were more common in mildly hypothermic patients compared to those who remained normothermic.

Finally, although the systemic complications appear relatively minimal for the transient (24 hour) application of mild or moderate hypothermia, one area of investigation that deserves further study is that of the effect of hypothermia on regenerative and endogenous defense mechanisms in brain. Goss et al (60) reported that 4 hours of moderate hypothermia resulted in a sustained inhibition of nerve growth factor production in brain after experimental contusion in rats. Nerve growth factor is an important homeostatic molecule in the central nervous system that upregulates antioxidant defenses and prevents apoptosis. The ramifications of this effect of hypothermia on brain parenchyma is currently under investigation.

Finally, another potential limitation of resuscitative hypothermia may be that it produces a temporary rather than sustained effect --i.e., delays rather than ameliorates damage. This possibility was first suggested in classic studies of the effect of hypothermia on acute inflammation (67,72), and was re-introduced in work by Dietrich et al (73) in models of global cerebral ischemia, where brief episodes (1-3 hours) of hypothermia only delayed death of neurons in selectively vulnerable brain regions. Recent work by Colbourne et al (74), however, suggests that longer durations of hypothermia may produce permanent benefit.

### Future directions

Some of the most intriguing recent work in the therapeutic application of hypothermia in laboratory studies involves the combination of hypothermia with other therapies. Dietrich et al (75) reported that combination of 3 hours of moderate hypothermia with sustained administration of the glutamate antagonist MK-801 produced a synergistic beneficial effect on neuronal survival in a model of global cerebral ischemia (Figure 3). Similar reports have been suggested for the combination of hypothermia and other therapies (76). Additional promising strategies that will require further study include the combination of hypothermia with either growth factors (60), anti-inflammatory agents or flow promoting treatments (60,77).

### References

1. Phelps C: Principles of Treatment. In Traumatic Injuries of the Brain and Its Membranes Critchley M, Flamm ES, Goodrich JT, et al. (eds), D. Appleton and Company, New York, p. 223, 1897.
2. Fay T: Observations on prolonged human refrigeration. NY State J Med 40: 1351, 1940.
3. Fay T: Observations on generalized refrigeration in cases of severe cerebral trauma. Assoc Res Nerv Ment Dis Proc 24: 611-619, 1943.
4. Woringer E, Schneider J, Baumgartner J, et al.: Essai critique sur l'effet de l'hibernation artificielle sur 19 cas de souffrance du tronc cerebral apres traumattisme selectionnes pour leur gravite parmi 270 comas postcommotionels. Anesth Analg (Paris) 11: 34-45, 1954.
5. Sedzimir CB: Therapeutic hypothermia in cases of head injury. J Neurosurg 16: 407-414, 1959.

6. Lazorhes G, Campan L: Hypothermia in the treatment of craniocerebral traumatism. *J Neurosurg* **15**: 162-167, 1958.
7. Rosomoff HL: Protective effects of hypothermia against pathological processes of the nervous system. *Ann NY Acad Sci* **80**: 475-486, 1959.
8. Albin MS, White RJ, Locke GE, et al.: Spinal cord hypothermia by localized perfusion cooling. *Nature* **210**: 1059, 1966.
9. Bigelow WG, Mustard WT, Evans JG: Some physiologic concepts of hypothermia and their applications to cardiac surgery. *J Thorac Surg* **28**: 463, 1954.
10. Swan H, Virtue RW, Blount SG, et al.: Hypothermias in surgery: Analysis of 100 clinical cases. *Ann Surg* **142**: 382, 1955.
11. Rosomoff HL: Hypothermia and cerebral vascular lesions. II. Experimental middle cerebral artery interruption followed by induction of hypothermia. *Arch Neurol & Psychiat* **78**: 454, 1957.
12. Siebke H, Rod T, Breivik H: Survival after 40 minutes submersion without cerebral sequelae. *Lancet* **1**: 1275-1277, 1975.
13. Conn AW, Edmonds JE, Barker GA: Cerebral resuscitation in near drowning. *Pediatr Clin North Am* **26**: 691, 1979.
14. Wolfe KB: Effect of hypothermia on cerebral damage resulting from cardiac arrest. *Am J Cardiol* **6**: 809, 1960.
15. Benson DW, Williams GR, Spencer FC, et al.: The use of hypothermia after cardiac arrest. *Anesth Analg* **38**: 423-428, 1958.
16. Ravitch M, Lane R, Safar P, et al.: Lightning Stroke. Recovery following cardiac massage and prolonged artificial respiration. *N Engl J Med* **264**: 36-38, 1961.
17. Rush BF, Wilder RJ, Fishbein R, et al.: Effects of total circulatory standstill in profound hypothermia. *Surgery* **50**: 40, 1962.
18. Bohn D, Biggar W, Smith C, et al.: Influence of hypothermia, barbiturate therapy, and intracranial pressure monitoring on morbidity and mortality after near-drowning. *Crit Care Med* **14**: 529, 1986.

19. Busto R, Dietrich WD, Globus MY-T, et al.: Small differences in intraischemic brain temperature critically determine the extent of ischemic neuronal injury. *J Cereb Blood Flow Metab* 7: 729-738, 1987.
20. Minamisawa H, Nordstrom C-H, Smith M-L, et al.: The influence of mild body and brain hypothermia on ischemic brain damage. *J Cereb Blood Flow Metab* 10: 365-374, 1990.
21. Chopp M, Chen H, Dereski MO, et al.: Mild hypothermic intervention after graded ischemic stress in rats. *Stroke* 22: 37-43, 1991.
22. Busto R, Dietrich WD, Globus MY, et al.: Postischemic moderate hypothermia inhibits CA1 hippocampal ischemic neuronal injury. *Neurosci Lett* 101: 299, 1989.
23. Gisvold SE, Safar P, Rao G, et al.: Multifaceted therapy after global brain ischemia in monkeys. *Stroke* 15: 803, 1984.
24. Sterz F, Safar P, Tisherman S, et al.: Mild hypothermic cardiopulmonary resuscitation improves outcome after prolonged cardiac arrest in dogs. *Crit Care Med* 19:379-389, 1991.
25. Weinrauch V, Safar P, Tisherman S, et al.: Beneficial effect of mild hypothermia and detrimental effect of deep hypothermia after cardiac arrest in dogs. *Stroke* 23:1454-1462, 1992.
26. Kuboyama K, Safar P, Radovsky A, et al.: Delay in cooling negates the beneficial effect of mild resuscitative cerebral hypothermia after cardiac arrest in dogs: A prospective, randomized, controlled study. *Crit Care Med* 21:1348-1358, 1993.
27. Leonov Y, Sterz F, Safar P, et al.: Moderate hypothermia after cardiac arrest of 17 minutes in dogs. Effect on cerebral and cardiac outcome. A preliminary study. *Stroke* 21: 1600, 1990.
28. Leonov Y, Sterz F, Safar P, et al.: Mild cerebral hypothermia during and after cardiac arrest improves neurologic outcomes in dogs. *J Cereb Blood Flow Metab* 10: 57, 1990.
29. Tisherman S, Chabal C, Safar P, et al.: Resuscitation of dogs from cold-water submersion using cardiopulmonary bypass. *Ann Emerg Med* 14: 389, 1985.

30. Safar P: Resuscitation from clinical death: Pathophysiologic limits and therapeutic potentials. *Crit Care Med* **16**:923-941, 1988.
31. Moore EE, Moore FA, Franciose RJ, et al.: Postischemic gut serves as a priming bed for circulating neutrophils that provoke multiple organ failure. *J Trauma* **37**:881-887, 1994.
32. Clifton GL, Jiang J, Lyeth B, et al.: Marked protection by moderate hypothermia after experimental traumatic brain injury. *J Cereb Blood Flow Metab* **11**: 114-121, 1991.
33. Palmer AM, Marion DW, Botscheller ML, et al.: Therapeutic hypothermia is cytoprotective without attenuating the traumatic brain injury-induced elevations in interstitial concentrations of aspartate and glutamate. *J Neurotrauma* **10**: 363-372, 1993.
34. Clark R, Kochanek PM, Marion D, et al.: Mild posttraumatic hypothermia reduces mortality after severe controlled cortical impact in rats. *J Cereb Blood Flow Metab* **16**: 253-261, 1996.
35. Mansfield RT, Schiding JK, Hamilton RL, et al.: Effects of hypothermia on traumatic brain injury in immature rats. *J Cereb Blood Flow Metab* **16**: 244-252, 1996.
36. Dietrich WD, Alonso O, Busto R, et al.: Post-traumatic brain hypothermia reduces histopathological damage following concussive brain injury in the rat. *Acta Neuropathol* **87**: 250-258, 1994.
37. Pomeranz S, Safar P, Radovsky A, et al.: The effect of resuscitative moderate hypothermia following epidural brain compression on cerebral damage in a canine outcome model. *J Neurosurg* **79**: 241-251, 1993.
38. Crippen D, Safar P, Porter L, et al.: Improved survival of hemorrhagic shock with oxygen and hypothermia in rats. *Resuscitation* **21**: 271-281, 1991.
39. Meyer DM, Horton JW: Effect of moderate hypothermia in the treatment of canine hemorrhagic shock. *Ann Surg* **207**:462, 1988.
40. Leonov Y, Safar P, Sterz F, et al.: Extending the golden hour of volume controlled hemorrhagic shock (VHS) in awake rats with oxygen (O<sub>2</sub>) plus moderate hypothermia (Hth). *Acad Emerg Med* **2**: 401 (abstract), 1995.

41. Kim S-H, Stezoski SW, Capone A, et al.: Hypothermia and minimal fluid resuscitation increase survival after uncontrolled hemorrhagic shock in rats. *J Trauma* **42**: 213-222, 1997.
42. Kim S-H, Stezoski SW, Safar P, et al.: Hypothermia, but not increased PaO<sub>2</sub> protects viscera during uncontrolled hemorrhagic shock in rats. *Crit Care Med* **25/1**(Suppl):A131, 1997 (abstract).
43. Safar P, Tisherman S, Carrillo P, et al.: Protecting the brain during severe hemorrhagic shock (HS). *Prehosp Disaster Med* **12**(Suppl): S12/80, 1997 (abstract).
44. Tisherman SA, Safar P, Radovsky A, et al.: Therapeutic deep hypothermic circulatory arrest in dogs: a resuscitation modality for hemorrhagic shock with 'irreparable' injury. *J Trauma* **30**: 836-847, 1990.
45. Tisherman SA, Safar P, Radovsky A, et al.: Profound hypothermia (< 10°C) compared with deep hypothermia (15°C) improves neurologic outcome in dogs after two hours' circulatory arrest induced to enable resuscitative surgery. *J Trauma* **31**: 1051-1062, 1991.
46. Capone A, Safar P, Radovsky A, et al.: Complete recovery after normothermic hemorrhagic shock and profound hypothermic circulatory arrest of 60 minutes in dogs. *J Trauma* **40**: 388-394, 1996.
47. Safar P, Abramson NS, Angelos M, et al.: Emergency cardiopulmonary bypass for resuscitation from prolonged cardiac arrest. *Am J Emerg Med* **8**:55-67, 1990.
48. Bellamy R, Safar P, Tisherman SA, et al.: Suspended animation for delayed resuscitation. *Crit Care Med* **24**: S24-S47, 1996.
49. Safar P: Cerebral resuscitation after cardiac arrest: Research initiatives and future directions. *Ann Emerg Med* **22**: 58-83, 1993.
50. Doppenberg EMR, Bullock R: Clinical neuro-protection trials in severe traumatic brain injury: Lessons from previous studies. *J Neurotrauma* **14**: 71-80, 1997.
51. McIntosh TK: Novel pharmacologic therapies in the treatment of experimental traumatic brain injury: A review. *J Neurotrauma* **10**: 215-261, 1993.
52. Natatori T, et al.: Delayed neuronal death in the CA1 pyramidal cell layer of the gerbil hippocampus following transient ischemia is apoptosis. *J Neurosci* **15**: 1001, 1995.



53. Kramer RS, Sanders AP, Lesage AM, et al.: The effect of profound hypothermia on preservation of cerebral ATP content during circulatory arrest. *J Thorac Cardiovasc Surg* **56**: 699, 1968.
54. Globus M, et al: Detection of free radical activity during transient global ischemia and recirculation: Effects of intra-ischemic brain temperature modulation. *J Neurochem* **65**: 1250, 1995.
55. Baiping L, Xiujuan T, Hongwei C, et al.: Effect of moderate hypothermia on lipid peroxidation in canine brain tissue after cardiac arrest and resuscitation. *Stroke* **25**:147, 1994.
56. Globus MY-T, Alonso O, Dietrich WD, et al.: Glutamate release and free radical production following brain injury: Effects of posttraumatic hypothermia. *J Neurochem* **65**: 1704-1711, 1995.
57. Rosomoff HL, Safar P: Management of the comatose patient. In: Respiratory Therapy Safar P (ed), FA Davis Co, Philadelphia, pp. 243-258, 1965.
58. Taft WC, Yang K, Dixon CE, et al.: Hypothermia attenuates the loss of hippocampal microtubule-associated protein 2 (MAP2) following traumatic brain injury. *J Cereb Blood Flow Metab* **13**: 796-802, 1993.
59. Smith SL, Hall ED: Mild pre- and posttraumatic hypothermia attenuates blood-brain barrier damage following controlled cortical impact injury in the rat. *J Neurotrauma* **13**: 1-9, 1996.
60. Goss J, Styren S, Miller P, et al.: Hypothermia attenuates the normal increase in interleukin 1b RNA and nerve growth factor following traumatic brain injury in the rat. *J Neurotrauma* **12**: 159-167, 1995.
61. Whalen MJ, Carlos TM, Clark RSB, et al.: The effect of brain temperature on acute inflammation after traumatic brain injury in rats. *J Neurotrauma*(in press).
62. Rosomoff HL, Kochanek PM, Clark R, et al.: Resuscitation from severe brain trauma. *Crit Care Med* **24**/S:S48-56, 1996.
63. Marion D, Obrist W, Carlier P, et al.: The use of moderate therapeutic hypothermia for patients with severe head injuries: A preliminary report. *J Neurosurg* **79**: 354-362, 1993.

64. Clifton GL, Allen S, Barrodale P, et al.: A phase II study of moderate hypothermia in severe brain injury. *J Neurotrauma* **10**: 263-271, 1993.
65. Shiozaki T, Sugimoto H, Taneda M, et al.: Effect of mild hypothermia on uncontrollable intracranial hypertension after severe head injury. *J Neurosurg* **79**: 363-368, 1993.
66. Marion DW, Penrod LE, Kelsey SF, et al.: Treatment of traumatic brain injury with moderate hypothermia. *N Engl J Med* **336**: 540-546, 1997.
67. Muschenheim C, Duerschner D, Hardy J, et al.: Hypothermia in experimental infections. *J Exp Med* **72**: 187-196, 1943.
68. Kurz A, Sessler DI, Lenhardt R: Perioperative normothermia to reduce the incidence of surgical-wound infection and shorten hospitalization. *N Engl J Med* **334**:1209-1215, 1996.
69. Mouritzen CV, Anderson MN: Mechanisms of ventricular fibrillation during hypothermia: relative changes in myocardial refractory period and conduction velocity. *J Thorac Cardiovasc Surg* **51**: 579-684, 1966.
70. Rohrer MJ, Natale AM: Effect of hypothermia on the coagulation cascade. *Crit Care Med* **20**: 1402-1405, 1992.
71. Frank SM, Fleisher LA, Breslow MJ, et al.: Perioperative maintenance of normothermia reduces the incidence of morbid cardiac events. A randomized clinical trial. *JAMA* **277**:1127-1134, 1997.
72. Svanes K: The influence of deep hypothermia on the formation of cellular exudate in acute inflammation in mice. *Acta Anesth Scand* **8**:143-156, 1964.
73. Dietrich WD, Busto R, Alonso O, et al.: Intraischemic but not postischemic brain hypothermia protects chronically following global forebrain ischemia in rats. *J Cereb Blood Flow Metab* **13**: 541-549, 1993.
74. Colbourne F, Corbett D: Delayed postischemic hypothermia: A six-month survival study using behavioral and histological assessments of neuroprotection. *J Neurosci* **15**: 7250, 1995.
75. Dietrich WD, Lin B, Globus MY-T, et al.: Effect of delayed MK-801 (Dizocilpine) treatment with or without immediate postischemic hypothermia on chronic neuronal survival after global forebrain ischemia in rats. *J Cereb Blood Flow Metab* **15**: 960-968, 1995.

76. Coimbra C, Drake M, Boris-Möller F, Wieloch T: Long-lasting neuroprotective effect of postischemic hypothermia and treatment with an anti-inflammatory/antipyretic drug. Evidence for chronic encephalopathic processes following ischemia. *Stroke* **27**:1578-1585, 1996.
77. Safar P, Xiao F, Radovsky A, et al.: Improved cerebral resuscitation from cardiac arrest in dogs with mild hypothermia plus blood flow promotion. *Stroke* **27**: 105-113, 1996.

Table 1. Glasgow Outcome Scores at 3, 6, and 12 months after severe head injury in patients treated with 24 hours of hypothermia (32 °C) or normothermia, from the study by Marion et al (66). A beneficial effect of hypothermia versus normothermia on outcome was demonstrated in all patients at 6 months after injury. In patients with initial Glasgow Coma Scores of 5-7, a favorable effect of hypothermia vs normothermia was observed at 3, 6 and 12 months after injury. Reprinted from the *New England Journal of Medicine* with permission.

Table 1

GLASGOW OUTCOME SCORE	At 3 MONTHS		At 6 MONTHS		At 12 MONTHS	
	HYPOTHERMIA	NORMOTHERMIA	HYPOTHERMIA	NORMOTHERMIA	HYPOTHERMIA†	NORMOTHERMIA
	number of patients (percent)					
All patients						
1 (Death)	8 (20)	9 (21)	8 (20)	10 (24)	9 (23)	10 (24)
2 (Vegetative state)	6 (15)	11 (26)	3 (8)	7 (17)	3 (8)	8 (19)
3 (Severe disability)	11 (28)	15 (36)	7 (18)	11 (26)	3 (8)	8 (19)
4 (Moderate disability)	8 (20)	4 (10)	7 (18)	8 (19)	9 (23)	5 (12)
5 (Mild or no disability)	7 (18)	3 (7)	15 (38)	6 (14)	15 (38)	11 (26)
Total	40	42	40	42	39	42
P value‡	0.12		0.05		0.18	
Patients with coma score of 5 to 7						
1 (Death)	2 (9)	5 (19)	2 (9)	6 (23)	2 (9)	6 (23)
2 (Vegetative state)	2 (9)	7 (27)	1 (5)	3 (12)	1 (5)	4 (15)
3 (Severe disability)	6 (27)	9 (35)	3 (14)	8 (31)	3 (14)	6 (23)
4 (Moderate disability)	6 (27)	3 (12)	4 (18)	6 (23)	5 (23)	2 (8)
5 (Mild or no disability)	6 (27)	2 (8)	12 (55)	3 (12)	11 (50)	8 (31)
Total	22	26	22	26	22	26
P value‡	0.01		0.01		0.04	

\*Percentages may not add to 100 because of rounding.

†One patient was lost to follow-up.

‡P values are for comparisons of all five outcomes in the hypothermia and normothermia groups.

### Figure Legends

Figure 1. Survival after uncontrolled hemorrhagic shock (UHS) in rats from the study of Kim et al (78). The insult in all groups is comprised of a volume controlled initial hemorrhage followed by tail amputation. Treatments include normothermia (Nth, Group 1), hypothermia (Hth, Group 2, 30°C applied between 15 min and 120 min), normothermia plus lactated Ringers (LR) fluid resuscitation (Nth + LR, Group 3), or hypothermia plus fluid resuscitation (Hth + LR, Group 4). Survival to 72 hours was maximal in rats treated with hypothermia plus LR. Reprinted from the *Journal of Trauma* with permission.

Figure 2. Desitometric analysis of RNA gel blot hybridizations for IL-1 $\beta$  message before, and at serial times after experimental cerebral contusion in rats, from the work of Goss et al (60). Filled circles represent data from rats maintained at brain temperature 37 °C, while solid squares represent data from rats maintained at a brain temperature of 32 °C for 4 hours after injury. A marked increase in IL-1 $\beta$  message was observed at 5.5 hours after injury which was partially attenuated by hypothermia. Reprinted from *the Journal of Neurotrauma* with permission.

Figure 3. Bar graph from the work of Dietrich et al (75) showing the number of normal appearing neurons in striatum at 2 months after sham operation or cerebral ischemia in rats treated with normothermia (37 °C), the glutamate receptor antagonist MK-801,

hypothermia (30 °C), or the combination of hypothermia plus MK-801. Neuronal survival was maximal after treatment with the combination of moderate hypothermia and MK-801. Reprinted from the *Journal of Cerebral Blood Flow and Metabolism* with permission.

### CME Questions

1. Randomized, controlled clinical trials of moderate hypothermia in cerebral resuscitation have progressed most rapidly for which of the following insults?
  - a. Stroke
  - b. Cardiopulmonary arrest
  - c. Traumatic brain injury
  - d. Near drowning
  - e. Hemorrhagic shock
2. This factor re-vitalized interest in the potential application of deliberate hypothermia to cerebral resuscitation.
  - a. Remarkably mild hypothermia (a temperature reduction of between 3° and 5°C) was effective in reducing secondary brain damage in laboratory models.
  - b. Brain temperature, not body temperature, was a critical therapeutic target.
  - c. The duration of hypothermia necessary for a beneficial effect in some

models was transient --as short as 1 hour,

d. b and c

e. All of the above

3. Controlled clinical trials of moderate hypothermia, applied for 24 hours, suggest that the incidence of side effects such as coagulopathy, arrhythmias, or infectious complications are not significantly different from that seen during normothermia.

a. True

b. False

4. Based on the results of clinical trials, beneficial effects of mild or moderate hypothermia in injured brain may result from a reduction of which of the following?

a. Excitotoxicity

b. Regeneration

c. Inflammation

d. a and c

e. All of the above

5. Recent studies in laboratory models of hemorrhagic shock have demonstrated that application of mild or moderate hypothermia during the "golden hour" reduces mortality rate.

a. True

b. False



Answers: 1 (c), 2 (e), 3 (a), 4 (d), 5 (a)

Figure 1

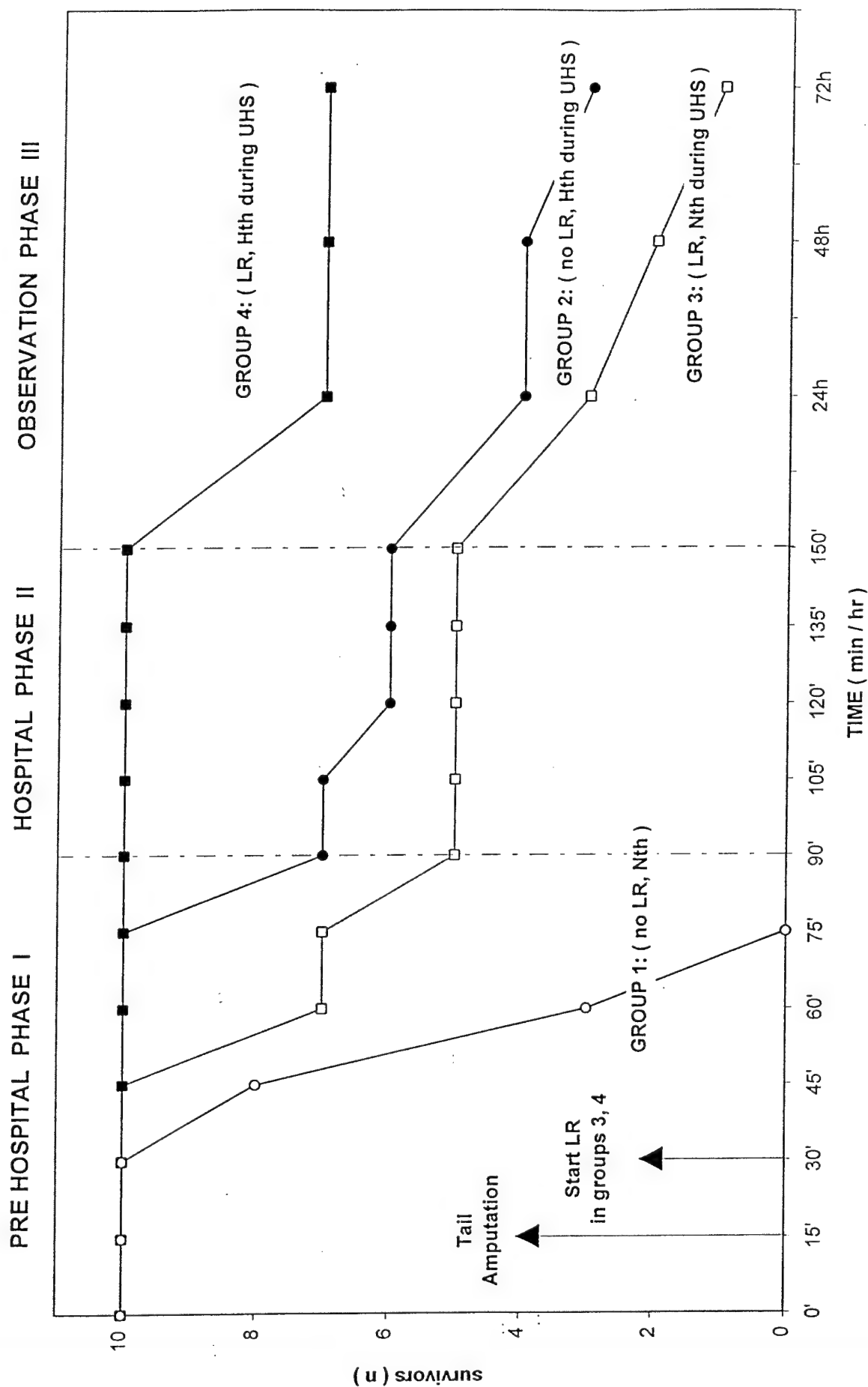


Figure 2

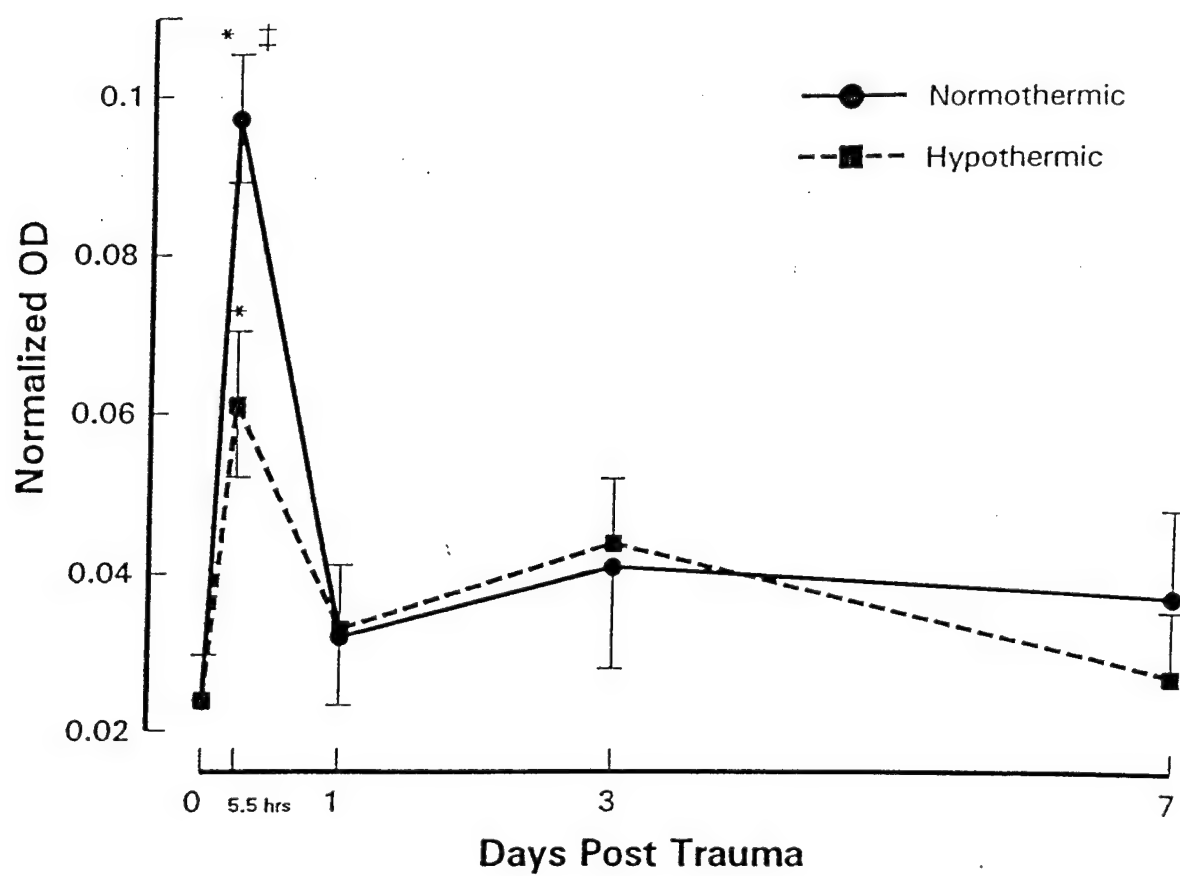
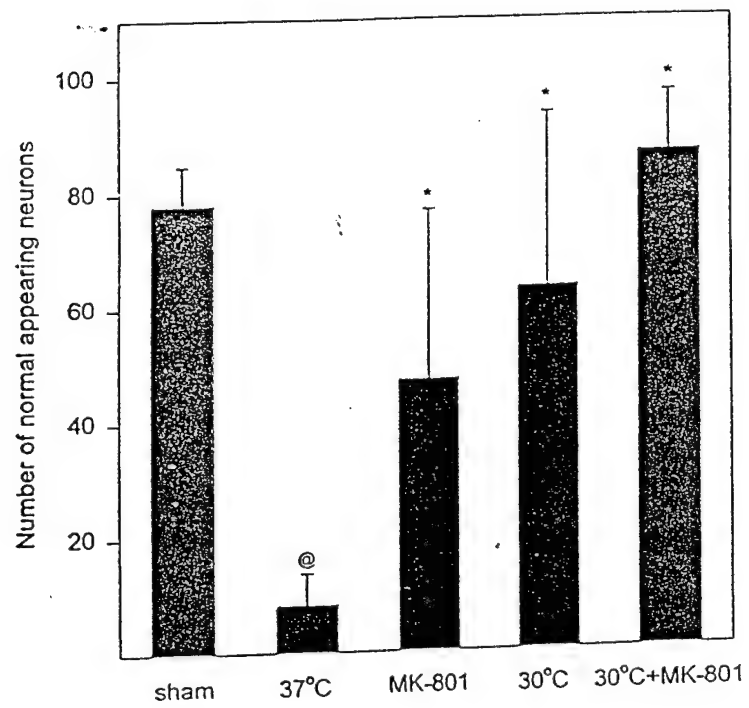


Figure 3



# No long-term benefit from hypothermia after severe traumatic brain injury with secondary insult in rats

Courtney L. Robertson, MD; Robert S. B. Clark, MD; C. Edward Dixon, PhD; Henry L. Alexander, BS; Steven H. Graham, MD, PhD; Stephen R. Wisniewski, PhD; Donald W. Marion, MD; Peter J. Safar, MD; Patrick M. Kochanek, MD

**Objectives:** To evaluate the effect of application of transient, moderate hypothermia on outcome after experimental traumatic brain injury (TBI) with a secondary hypoxemic insult.

**Design:** Prospective, randomized study.

**Setting:** University-based animal research facility.

**Subjects:** Male Sprague-Dawley rats.

**Interventions:** All rats were subjected to severe TBI followed by 30 mins of moderate hypoxemia, associated with mild hypotension. Rats were randomized to three groups: a) normothermia ( $37^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$ ); b) immediate hypothermia ( $32^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$  initiated after trauma, before hypoxemia); and c) delayed hypothermia ( $32^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$  after hypoxemia). The brain temperature was controlled for 4 hrs after TBI and hypoxemia.

**Measurements and Main Results:** Animals were evaluated after TBI for motor and cognitive performance using beam balance (days 1–5 after TBI), beam walking (days 1–5 after TBI), and

Morris Water Maze (days 14–18 after TBI) assessments. On day 21 after TBI, rats were perfused with paraformaldehyde and brains were histologically evaluated for lesion volume and hippocampal neuron counts. All three groups showed marked deficits in beam balance, beam walking, and Morris Water Maze performance. However, these deficits did not differ between groups. There was no difference in lesion volume between groups. All animals had significant hippocampal neuronal loss on the side ipsilateral to injury, but this loss was similar between groups.

**Conclusions:** In this rat model of severe TBI with secondary insult, moderate hypothermia for 4 hrs posttrauma failed to improve motor function, cognitive function, lesion volume or hippocampal neuronal survival. Combination therapies may be necessary in this difficult setting. (Crit Care Med 2000; 28:000–000)

**KEY WORDS:** traumatic brain injury; hypothermia; hypoxemia

Secondary insults after experimental traumatic brain injury (TBI) have been shown to exacerbate disturbances in key physiologic variables, including hypoperfusion, energy failure, cerebral edema, and electroencephalographic suppression (1–3). In addition, animals subjected to a secondary hypoxemic insult after TBI have worse motor and histologic outcomes than those subjected to TBI alone (1, 3, 4). After severe TBI, patients often

experience a variety of secondary systemic insults related to extracerebral traumatic injury. As many as 20% to 50% of patients presenting with severe TBI have experienced a period of hypoxemia (5–7). Postmortem findings of head-injured patients (8) demonstrate evidence of ischemic neuronal death throughout the brain. Similarly, clinical studies have demonstrated higher morbidity and mortality among head-injured patients who had experienced a secondary insult, specifically hypoxemia or hypotension (7). Often, the most severely devastated patients are those who experience the combination of TBI with hypoxemia and hypotension.

Hypothermia has been used as a successful treatment modality after brain injury in many experimental models and clinical settings. Neuroprotective effects of hypothermia in animal models include attenuation of release of excitatory amino acids (9–11), reduction in hydroxyl radicals (9) and inflammatory mediators (12, 13), and reduction in disruption of the blood-brain barrier (14). In the setting of experimental TBI, hypothermia improves

outcome (15, 16). In models of fluid percussion injury (15) and controlled cortical impact (CCI) (16), reductions in both functional and motor deficits are observed in animals treated with moderate hypothermia after TBI when compared with normothermic animals. Transient, moderate hypothermia applied after global or focal ischemic insult in animal models has improved histologic outcomes (17–19). Clinical studies have similarly demonstrated improvements in functional outcomes (20) and intracranial pressure (21, 22) in patients treated with moderate hypothermia.

Despite the importance of secondary insults to clinical outcome after TBI, the variety of experimental models of TBI and secondary insult that have been developed, and the success of hypothermia in both clinical and experimental TBI, the effect of the application of hypothermia in the setting of TBI with secondary insult has not been studied. We hypothesized that moderate hypothermia would improve outcome after CCI with secondary insult in rats.

From the Departments of Anesthesiology and Critical Care Medicine (Drs. Robertson, Clark, Alexander, Safar, Kochanek), Pediatrics (Drs. Clark, Kochanek), Neurological Surgery (Drs. Dixon, Marion), and Neurology (Dr. Graham), the School of Public Health (Dr. Wisniewski), the Safar Center for Resuscitation Research, and the Brain Trauma Research Center, University of Pittsburgh; and the Geriatric Research Educational and Clinical Center, Pittsburgh VAHS (Dr. Graham), Pittsburgh, PA.

Supported, in part, by U.S. Army #DAMD 17-97-1-7009.

Address requests for reprints to: Patrick M. Kochanek, MD, Safar Center for Resuscitation Research, 3434 Fifth Avenue, Pittsburgh, PA 15260. E-mail: kochanek@smtp.anes.upmc.edu

Copyright © 2000 by Lippincott Williams & Wilkins

## MATERIALS AND METHODS

This study was approved by the University of Pittsburgh Animal Care and Use Committee. The care and handling of animals were in accord with National Institute of Health guidelines.

**Experimental Protocol.** Virus-free male Sprague-Dawley rats (329–460 g) were studied. The animals were allowed free access to food and water before and after surgery. All surgical procedures were performed using aseptic technique.

Anesthesia was induced in a plastic jar with 4% isoflurane (Anaquest, Memphis, TN) in oxygen. The trachea was intubated with a 14-gauge angiocatheter and the lungs were mechanically ventilated with 2% isoflurane/66% N<sub>2</sub>O/balance oxygen. A femoral arterial catheter (PE-50) was inserted for continuous monitoring of blood pressure and arterial blood sampling. Pancuronium bromide (0.1 mg/kg/hr) (Elkins-Sinn, Cherry Hill, NJ) was given for immobilization. A rectal probe was inserted to monitor core temperature.

**Traumatic Brain Injury Model.** The head was fixed in a stereotactic device (David Kopf, Tujunga, CA) and a midline scalp incision was made to expose the parietal bone. A craniotomy was made over the left parietal cortex with a dental drill, using the coronal and interparietal sutures as margins. The intact dura and bone flap were left in place until immediately before trauma. A temperature probe (outside diameter, 0.009 inch) (Physiotemp, Clifton, NJ) was inserted through a burr hole into the left parietal cortex 5 mm anterior to the bregma and 2 mm lateral to the sagittal suture. Rats were equilibrated under anesthesia (1.1% isoflurane/66% N<sub>2</sub>O/balance oxygen) at a brain temperature of  $37^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$  for 30 mins before TBI. An arterial blood sample (0.5 mL) was obtained 15 mins before trauma to verify normal arterial blood gas tensions, serum glucose, and hematocrit.

TBI was performed using the CCI device (23, 24), with minor modifications to the procedure previously described (25). Briefly, after removal of the bone flap, injury was produced using a device with a 6-mm metal impactor tip that is pneumatically driven in the vertical plane at a predetermined depth, velocity, and duration of brain deformation. For all studies, a depth of penetration of 2.5 mm, a velocity of  $4.0 \pm 0.2$  m/sec, and a duration of deformation of 50 msec were used. After trauma, the bone flap was replaced and sealed with dental cement (Koldmount, Vernon Banisoff, Albany, NY) and the scalp incision was closed.

**Secondary Insult.** Beginning 1 min after CCI, all rats underwent a 30-min period of hypoxemia as previously described (4). Air and oxygen were blended to achieve an FIO<sub>2</sub> of 11% (1.1% isoflurane/74% N<sub>2</sub>O/19% air/6% oxygen). This produced a PaO<sub>2</sub> range of 46–51 and MAP range of 50–74. Arterial blood gas samples were obtained in all rats at 10 mins and 25 mins during the hypoxic period.

**Hypothermia.** Rats were randomized into three groups: normothermia, immediate hypothermia, and delayed hypothermia. Hypothermia ( $32^{\circ}\text{C}$ ) was achieved by the external application of ice packs to the head to lower brain temperature over a 15-min period. This temperature was maintained for 4 hrs and the brain was rewarmed over 1 hr. Arterial blood gas samples were obtained every hour during the hypothermia period. Physiologic variables (MAP, brain temperature, rectal temperature) were recorded every 30 mins during hypothermia and during rewarming.

The immediate hypothermia group ( $n = 10$ ) had brain cooling initiated immediately after trauma, coincident with the onset of hypoxemia. The delayed hypothermia group ( $n = 10$ ) had cooling initiated on the completion of hypoxemia, 30 mins after TBI. The normothermia group ( $n = 19$ ) had brain temperature maintained at  $37^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$  throughout the experimental period. At the end of the experiment, after completion of rewarming, anesthesia was discontinued. Rats were extubated, placed in 100% oxygen for an additional 30 mins, then returned to their cages, where they were allowed free access to food and water.

**Motor Function Assessments.** Gross vestibulomotor function was assessed using a beam-balance task (24, 26). The rats were trained by three trials before TBI to obtain a baseline measurement. Beam-balance latency (up to 60 secs) was measured on days 1–5 after TBI.

Fine vestibulomotor function and coordination were assessed using a beam-walking task (27). Performance was assessed by measuring the rat's latency to traverse the beam and enter a goal box. Beam-walking latency was measured on days 1–5 after TBI.

**Cognitive Assessment (Morris Water Maze).** Water-maze testing started on day 14 postinjury. The hidden platform task assesses the rat's ability to learn spatial relations between distal cues and the escape platform. Performance is impaired by cortical and hippocampal lesions. We used a variant of the Morris water maze (28). Rats were given 120 secs to find the hidden platform. If the rat failed to find the platform within 120 secs, it was placed on the platform by the experimenter. Rats were given four swimming trials per day for 5 consecutive days. Water maze tests were given on days 14–18 after TBI. The last 2 days of testing consisted of a visible platform task in which the platform was raised 2 cm above the water surface. This visible task controls for potential nonspecific visual or motor deficits.

**Lesion Volume Analysis.** At 21 days after TBI, rats were anesthetized and perfused with 500 mL of 4% buffered formaldehyde. Brains were removed and postfixed for a minimum of 1 wk at  $4^{\circ}\text{C}$  and cryoprotected in sucrose. Coronal sections (10- $\mu\text{m}$ ) were prepared through the entire brain at 1-mm intervals from the occiput. Sections were stained with

cresyl violet. In the serial sections taken at 1-mm intervals, the margins of both the contusion and the total left hemisphere were outlined by a blinded observer using image analysis (Image Research). Contusion and hemispheric areas were measured. Contusion volume was calculated and expressed as mm<sup>3</sup>.

**Hippocampal Cell Counts.** Neuronal loss in hippocampal CA1 and CA3 pyramidal layers was quantified using a method previously described by Clark et al. (4). A coronal section through the dorsal hippocampus underlying the area of contusion was used for analysis. This location was  $\sim 2.6$  mm posterior to bregma. The regions were visualized at magnification  $\times 400$  by a blinded observer. CA1 and CA3 hippocampal neurons were counted in six separate fields for each region in both injured and uninjured hemispheres. Only complete cells with a defined cell body and intact nucleus were counted. Hippocampal neuron survival was reported as the average number of surviving neurons per high power field ipsilateral to injury.

**Statistical Analysis.** All data are presented as mean  $\pm$  SEM. Because of the large number of physiologic variables recorded, comparisons of physiologic data were made using a multiple regression analysis, evaluating the effect of treatment and time for each variable. Beam balance, beam walking, and Morris water maze data were analyzed using repeated-measures analysis of variance using GB-STAT statistical software. Lesion volumes and hippocampal neuron counts were compared using the Kruskal-Wallis test and Dunn's test. A significance level of  $p < .05$  was used for all tests.

## RESULTS

**Physiologic Variables.** Physiologic data (including brain and rectal temperature, MAP, arterial pH and blood gasses, blood glucose, and hematocrit) from all animals that survived the experimental protocol are presented in Table 1. There were no differences between groups in pH, PaCO<sub>2</sub>, and hematocrit. As expected, the groups differed in brain and rectal temperatures ( $p < .05$ ), and this difference was seen between groups when controlling for time. Brain temperature remained at  $37^{\circ}\text{C} \pm 0.2^{\circ}\text{C}$  in the normothermia group. Brain temperature decreased from  $37.2^{\circ}\text{C} \pm 0.1^{\circ}\text{C}$  before trauma to  $32.0^{\circ}\text{C} \pm 0.1^{\circ}\text{C}$  by 25 mins of cooling in the immediate hypothermia group. Brain temperature decreased from  $37.1^{\circ}\text{C} \pm 0.1^{\circ}\text{C}$  before trauma to  $32.1^{\circ}\text{C} \pm 0.1^{\circ}\text{C}$  by 25 mins of cooling in the delayed hypothermia group.

Groups differed in MAP when controlling for time ( $p < .05$ ). Initial MAP in the normothermia group was 91–92 mm Hg and decreased to 62–63 mm Hg during

AQ: B

AQ:

AQ: I

TI

Table 1. Physiologic data

Group	Brain Temperature (°C)	Rectal Temperature (°C)	MAP (mm Hg)	pH	Paco <sub>2</sub> (mm Hg)	Pao <sub>2</sub> (mm Hg)	Glucose (mg/dL)	HCT (%)
37°C (n = 19)								
Baseline	37.1 ± 0.1	37.3 ± 0.1	91 ± 3	7.44 ± 0.01	39 ± 1	169 ± 4	182 ± 10	40 ± 0
Insult	37.0 ± 0.1	37.3 ± 0.0	92 ± 3	—	—	—	—	—
10 mins	37.1 ± 0.1	37.1 ± 0.1	63 ± 4	7.43 ± 0.01	40 ± 1	46 ± 1	—	—
25 mins	37.0 ± 0.0	37.1 ± 0.1	62 ± 3	7.42 ± 0.01	40 ± 1	47 ± 1	173 ± 13	39 ± 0
1 hr	37.1 ± 0.0	37.2 ± 0.1	84 ± 2	7.43 ± 0.01	38 ± 1	160 ± 3	145 ± 4	39 ± 0
3 hrs	37.1 ± 0.1	37.2 ± 0.0	85 ± 3	7.42 ± 0.01	37 ± 1	172 ± 3	148 ± 4	38 ± 0
32°C—Immediate (n = 10)								
Baseline	37.2 ± 0.1	37.3 ± 0.1	103 ± 2	7.45 ± 0.01	40 ± 1	157 ± 3	171 ± 8	41 ± 1
Insult	37.2 ± 0.0	37.3 ± 0.1	104 ± 3	—	—	—	—	—
10 mins	32.5 ± 0.2	32.5 ± 0.2	74 ± 5	7.44 ± 0.01	37 ± 1	51 ± 1	—	—
25 mins	32.0 ± 0.1	32.1 ± 0.1	68 ± 3	7.45 ± 0.01	38 ± 1	50 ± 1	208 ± 22	39 ± 0
1 hr	31.9 ± 0.1	32.0 ± 0.1	91 ± 4	7.45 ± 0.01	38 ± 1	201 ± 5	166 ± 11	39 ± 0
3 hrs	32.1 ± 0.1	32.0 ± 0.1	94 ± 3	7.42 ± 0.01	38 ± 0	193 ± 6	182 ± 15	39 ± 0
32°C—Delayed (n = 10)								
Baseline	37.1 ± 0.1	37.3 ± 0.1	85 ± 2	7.43 ± 0.01	39 ± 1	176 ± 7	196 ± 14	41 ± 1
Insult	37.1 ± 0.1	37.3 ± 0.1	84 ± 2	—	—	—	—	—
10 mins	36.9 ± 0.1	37.1 ± 0.1	54 ± 3	7.42 ± 0.01	40 ± 1	47 ± 2	—	—
25 mins	36.9 ± 0.1	37.1 ± 0.1	50 ± 2	7.41 ± 0.01	39 ± 1	47 ± 1	173 ± 10	39 ± 0
1 hr	31.9 ± 0.1	31.9 ± 0.1	76 ± 2	7.43 ± 0.00	38 ± 1	205 ± 3	188 ± 11	39 ± 0
3 hrs	31.9 ± 0.1	32.0 ± 0.1	71 ± 5	7.41 ± 0.01	36 ± 0	215 ± 3	182 ± 9	37 ± 1

MAP, mean arterial pressure; HCT, hematocrit.  
Values are mean ± SEM.

hypoxemia, returning to a postinsult level of 84–85 mm Hg. The immediate hypothermia group started with a baseline MAP of 103–104 mm Hg, decreased to 68–74 mm Hg during hypoxemia, and returned to 91–94 mm Hg postinsult. The delayed hypothermia group had the lowest overall MAP, starting with a baseline of 84–85 mm Hg, decreasing to 50–54 mm Hg during hypoxemia and returning to 71–76 mm Hg postinsult.

Groups also differed in Pao<sub>2</sub> and glucose when controlling for time (both  $p < .05$ ). The differences between groups appeared modest and unlikely to be clinically significant. Glucose levels were also different between groups over time. The normothermia group had initial glucose levels of 182 mg/dL and decreased to levels of 148 mg/dL at 3 hrs postinsult. The immediate and delayed hypothermia groups had more stable glucose levels.

**Survival.** Survival rate to 21 days for completion of motor and cognitive testing was 79% for the normothermia group, 80% for the immediate hypothermia group, and 62% for the delayed hypothermia group. Motor performance, cognitive performance, lesion volume analysis and hippocampal neuron counts are reported on all animals that survived to cognitive testing at 20 days and were able to swim for water maze testing.

**Motor Performance.** All three groups showed a decrease in beam balance per-

formance and an increase in beam walking latency after trauma. However, there was no difference in beam balance duration (Fig. 1) or beam walking latency (Fig. 2) between normothermic and hypothermic groups.

**Cognitive Performance (Morris Water Maze).** All three groups showed a marked latency in finding the submerged platform on days 14–18 after trauma. However, there was no difference between groups in performance in the water maze (Fig. 3). The groups were similar in discovery of the visible platform on days 19–20 after trauma (Fig. 3).

**Lesion Volume Analysis.** Lesion volumes (mm<sup>3</sup>) measured at 21 days after TBI are shown in Table 2. There appeared to be a reduction in lesion volume in the hypothermia vs. normothermic groups. However, this reduction in lesion volume was not statistically different from normothermia. Lesion area at various distances from the occiput is shown in Figure 4.

**Hippocampal Neuron Counts.** Surviving hippocampal neuron counts are shown in Table 3. Both normothermic and hypothermic animals had significant hippocampal neuronal loss on the side ipsilateral to injury. For comparison, average neuron counts in CA1 and CA3 hippocampus on the side contralateral to injury were 48–56 cells/high-power field. There were no significant differences in

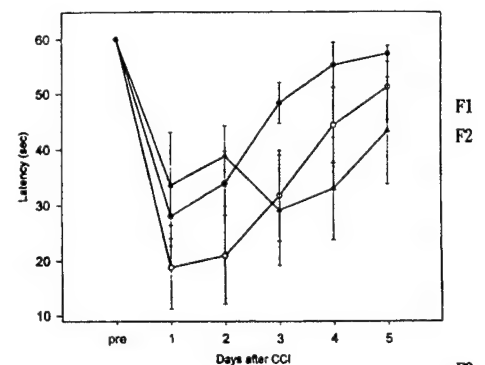


Figure 1. Beam balance latency in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) on days 1–5 after controlled cortical impact (CCI). Values are mean ± SEM.

the number of surviving CA1 or CA3 hippocampal neurons in the hypothermic vs. normothermic groups.

## DISCUSSION

To our knowledge, this is the first study to evaluate the effect of hypothermia on severe experimental TBI with secondary insult. Surprisingly, no difference in motor performance, cognitive performance, lesion volume, or hippocampal neuronal survival was observed with the application of moderate hypothermia after severe TBI with secondary insult in a rat model. Also unexpectedly, both the



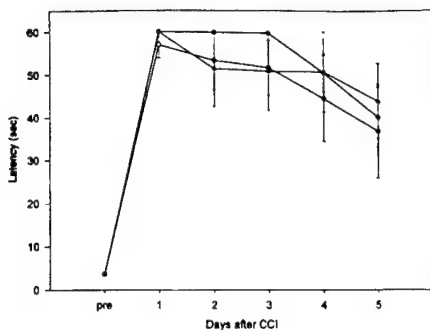


Figure 2. Beam walking latency in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) on days 1–5 after controlled cortical impact (CCI). Values are mean  $\pm$  SEM.

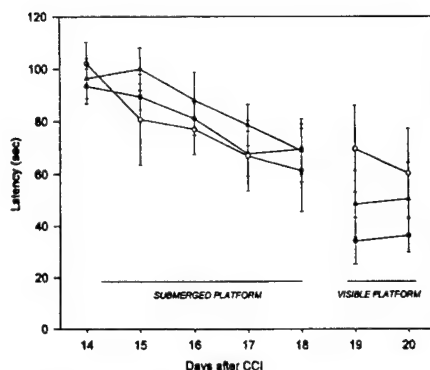


Figure 3. Morris Water Maze performance in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) on days 14–18 after controlled cortical impact (CCI); and visible platform latency in all groups on days 19–20 after CCI. Values are mean  $\pm$  SEM.

Table 2. Lesion Volume

Group	Lesion Volume (mm <sup>3</sup> ) <sup>a</sup>
Normothermia	65.3 $\pm$ 6.9
Immediate HT	50.2 $\pm$ 8.2
Delayed HT	53.7 $\pm$ 7.9

HT, hypothermia.

<sup>a</sup> $p = .32$  by analysis of variance. Values are expressed as mean  $\pm$  SEM.

immediate hypothermia group, with hypothermia initiated after trauma and before secondary hypoxemia, and the delayed hypothermia group, with hypothermia applied after both brain trauma and hypoxemia, demonstrated similar functional and histologic deficits when compared with each other and to the normothermia group.

Beneficial effects of hypothermia on histopathologic outcome after TBI have

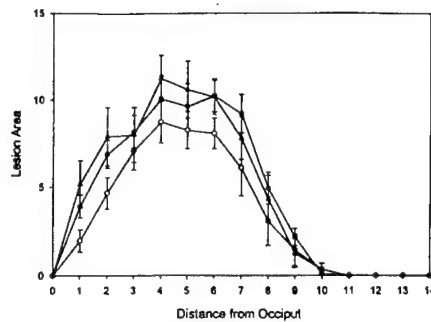


Figure 4. Lesion area at various distances from the occiput (mm) in normothermia (solid circles), immediate hypothermia (open circles), and delayed hypothermia (triangles) groups. Values are mean  $\pm$  SEM.

Table 3. Hippocampal neuronal survival

Group	CA1 Neurons (cells/hpf) <sup>a</sup>	CA3 Neurons (cells/hpf) <sup>a</sup>
Normothermia	19.4 $\pm$ 4.2	19.8 $\pm$ 4.6
Immediate HT	13.2 $\pm$ 8.7	15.6 $\pm$ 7.3
Delayed HT	13.7 $\pm$ 5.8	18.5 $\pm$ 7.3

hpf, high-powered field; HT, hypothermia.

<sup>a</sup>Ipsilateral to injury. Values are expressed as mean  $\pm$  SEM.

been demonstrated by several investigators. The timing of this histologic evaluation appears to be important. Dietrich et al. (29) showed a reduction in cortical contusion volume and frequency of necrotic cortical neurons in rats that received 3 hrs of immediate posttrauma hypothermia (30°C) after parasagittal fluid percussion injury. These animals were evaluated at 3 days posttrauma. However, in the same model, investigators found no difference in hippocampal CA1, CA3, CA4, or dentate neuronal survival in rats receiving posttrauma hypothermia compared with normothermic animals when brains were analyzed at 8 wks after TBI (30). In models of ischemic brain injury, transient application of therapeutic hypothermia has also shown a temporary beneficial effect on hippocampal neuronal survival. Early evaluation revealed decreased hippocampal CA1 cell loss, but this protection by posttrauma hypothermia (30°C) was not seen in the animals evaluated at 2 months after ischemia insult (31). In our model, severe TBI was followed by 30 mins of hypoxemia. All animals showed a reduction in systemic blood pressure during the hypoxemic period, likely highlighting an ischemic component to the secondary insult.

**I**n this rat model of severe controlled cortical impact with hypoxemia, moderate hypothermia for 4 hrs post-trauma failed to improve hippocampal neuron survival, lesion volume, motor function, or cognitive function.

The amount of tissue loss after experimental TBI varies greatly dependent on the model. This model of severe CCI followed by 30 mins of hypoxemia produced lesion volumes of 50–65 mm<sup>3</sup>. This is much larger than contusion volumes seen in other traumatic injury models, such as lateral fluid percussion (2.14 mm<sup>3</sup>) (27), or in similar CCI models without hypoxemia (~30 mm<sup>3</sup>) (5). The severe insult produced in this model might explain the failure of posttrauma hypothermia to show a significant reduction in lesion volume. However, other experimental TBI models applying hypothermia after injury have also failed to reduce necrotic volumes (30, 32). Cherian et al. (33) showed increasing sizes of contusion volume as the degree of secondary insult (bilateral carotid occlusion) increased after CCI in rats. In addition, it is likely that the lesion volume observed at 21 days postinjury is the result of damage by many different mechanisms operating in the early and late posttrauma phases. Clark et al. (4) has demonstrated cells with either necrotic or apoptotic phenotypes in various brain regions after TBI in a similar model with hypoxemia. It is unclear if earlier assessment would have revealed more hippocampal protection with posttraumatic hypothermia in this model. However, our goal is to favorably influence long-term outcome. Hypothermia as a single treatment modality, and applied for only 4 hrs after TBI, might be unable to reduce overall lesion volumes in such a model.

Previous experimental studies applying moderate hypothermia after TBI have demonstrated protection against motor

and spatial memory deficits after both CCI (16) and fluid percussion injury (30, 34). However these models did not include a period of hypoxemia or any other secondary insults after trauma. This secondary hypoxic insult worsens histologic outcome and could therefore worsen behavioral outcome after TBI. In a similar model of CCI with secondary hypoxemia, rats demonstrated progressively worse motor function (beam-balance latency) with increasing amounts of posttrauma hypoxemia (4). This trend was seen even in rats who received mild hypoxemia ( $\text{PaO}_2$ , 58–63 mm Hg) after CCI. In a fluid percussion injury model, Ishige et al. (1) showed significantly worse neurologic status scores in rats that underwent impact injury followed by a 30-min period of hypoxemia ( $\text{PaO}_2$ , 35–40 mm Hg) vs. those injured without secondary hypoxemia. These neurologic deficits were also not observed in rats that received hypoxemia alone.

Clinical studies of patients with head injury have also reported marked worsening of outcome variables in the setting of TBI with secondary insult such as hypoxemia or hypotension. In an analysis of 717 patients from the Traumatic Coma Data Bank, Chesnut et al. (7) found that hypoxia and hypotension were independently associated with increases in morbidity and mortality from severe head injury. This study showed a marked shift toward vegetative/dead outcomes in patients who endured hypoxia ( $\text{PaO}_2 \leq 60$  mm Hg) or hypotension (systolic blood pressure  $\leq 90$ ) during the prehospital or resuscitation period. This is especially relevant to our model of TBI because rats underwent a planned 30-min period of hypoxemia, which was also associated with a decrease in systemic blood pressure. The difference in MAP between groups may have caused additional experimental differences. The delayed hypothermia group had an overall lower MAP trend and may have experienced more significant secondary ischemic flows.

Clinical studies applying hypothermia after TBI have yielded a variety of positive results. In a phase II study of moderate hypothermia, Clifton et al. (35) found a reduction in incidence of posttraumatic seizures. Shiozaki et al. (22) documented improved control of intracranial pressure with the application of mild (34°C) hypothermia after conventional therapies had failed to control intracranial pressure (22). Most recently, Marion et al. (20) demonstrated faster recovery of func-

tional outcome with the application of moderate hypothermia (32°C) after severe TBI in 82 patients randomized to either normothermia or hypothermia for 24 hrs after injury. Relevant to our findings, these clinical trials showed important distinctions in the subset of very severely injured patients. Marion's study included all patients with initial Glasgow Coma Scale (GCS) scores of  $\leq 8$ , but the beneficial effect of hypothermia only extended to 12 months in the subset of patients with an initial GCS score of 5–7 (20). In the study by Shiozaki et al. (22), the subset of patients admitted with GCS scores of 3–4 had a much lower incidence of favorable outcome at 6 months after injury (only 1 of 22 patients, 4.5%) vs. the group with GCS scores of 5–7 (11 of 40 patients, 27.5%), despite the application of mild hypothermia. Importantly, Marion et al. (20) excluded patients with hypoxia or hypotension.

Our model of CCI with hypoxemia results in a very severe injury relative to other models. In addition, during the 30-min period of hypoxemia, the rats experience a significant drop in their mean arterial pressure. In a recent experimental TBI model (36), posttraumatic application of moderate hypothermia (30°C for 3hrs) resulted, after rewarming, in a lower cerebral perfusion without a corresponding decrease in cerebral glucose utilization, creating a state of metabolism to blood flow mismatch. This may be especially important in the clinical setting in which severely head-injured patients undergo a secondary insult of hypoxia and/or hypotension before initiation of treatment for their head injury. Analysis of trauma patients has consistently revealed worse outcomes in patients who experienced sustained hypotension in the prehospital setting vs. those who remained normotensive. Specifically, Chesnut et al. (7) showed hypotension was associated with a 150% increase in mortality rate. Wald et al. (37) also found that prehospital hypotension doubled the incidence of adverse outcome (37). In a review of pediatric trauma patients, Pigula et al. (38) found that hypotension significantly increased the mortality rate. As a result, patients with significant secondary insult have been excluded from evaluation in clinical trials (20). This subset clearly represents a population in which currently available interventions, even those proven to be effective in the sets of TBI alone, may have limited efficacy.

There are many mechanisms of cell injury and death after TBI. Investigators have shown additional pathophysiologic mechanisms operating when secondary insults were added to the already vulnerable traumatically injured brain (1, 3, 33, 39). Recent experimental investigations have focused on the period after trauma, during which secondary insults may potentiate neuronal damage, utilizing a wide variety of therapies. However, many of these therapies have been tested in models of TBI where oxygenation and ventilation of animals after TBI are controlled. Given that many head-injured patients present with preceding hypoxemia, it may be important to reassess these therapies in models imitating this clinical setting. It is possible that proven treatments may be less effective in a model with applied hypoxemia and accompanying hypotension. Novel therapies targeting this complex clinical scenario have yet to be developed.

In conclusion, in this rat model of severe CCI with hypoxemia, moderate hypothermia for 4 hrs posttrauma failed to improve hippocampal neuron survival, lesion volume, motor function, or cognitive function. Combination therapies or development of novel therapies may be necessary to see significant improvement in outcome in this difficult setting.

## ACKNOWLEDGMENT

We thank Marci Provins for assistance with preparation of the manuscript.

## REFERENCES

1. Ishige N, Pitts LH, Hashimoto T, et al: Effect of hypoxia on traumatic brain injury in rats: Part 1. *Neurosurgery* 1987; 20:848–853
2. Ishige N, Pitts LH, Pogliani L, et al: Effect of hypoxia on traumatic brain injury in rats: Part 2. *Neurosurgery* 1987; 20:854–858
3. Ishige N, Pitts LH, Berry I, et al: The effect of hypoxia on traumatic head injury in rats: alterations in neurologic function, brain edema, and cerebral blood flow. *J Cereb Blood Flow Metab* 1987; 7:759–767
4. Clark RSB, Kochanek PM, Dixon CE, et al: Early neuropathologic effects of mild or moderate hypoxemia after controlled cortical impact injury in rats. *J Neurotrauma* 1997; 14:179–189
5. Katsurada K, Yamada R, Sugimoto T, et al: Respiratory insufficiency in patients with severe head injury. *Surgery* 1973; 73:191–199
6. Sinha RP, Sicker TB, Perot PL Jr.: Arterial oxygenation findings and its significance in central nervous system trauma patients. *JAMA* 1973; 224:1258–1260

7. Chesnut RM, Marshall LF, Klauber MR, et al: The role of secondary brain injury in determining outcome from severe head injury. *J Trauma* 1993; 34:216-222
8. Kotapka MJ, Graham DI, Adams JH, et al: Hippocampal pathology in fatal non-missile human head injury. *Acta Neuropathol (Berl)* 1992; 83:530-534
9. Globus MY, Alonso O, Dietrich WD, et al: Glutamate release and free radical production following brain injury: Effects of post-traumatic hypothermia. *J Neurochem* 1995; 65:1704-1711
10. Busto R, Globus MY, Dietrich WD, et al: Effect of mild hypothermia on ischemia-induced release of neurotransmitters and free fatty acids in rat brain. *Stroke* 1989; 20:904-910
11. Koizumi H, Fujisawa H, Ito H, et al: Effects of mild hypothermia on cerebral blood flow-independent changes in cortical extracellular levels of amino acids following contusion trauma in the rat. *Brain Res* 1997; 747:304-312
12. Goss JR, Styren SD, Miller PD, et al: Hypothermia attenuates the normal increase in interleukin 1 $\beta$  RNA and nerve growth factor following traumatic brain injury in the rat. *J Neurotrauma* 1995; 12:159-167
13. Whalen MJ, Carlos TM, Clark RSB, et al: Hypothermia reduces acute inflammation after traumatic brain injury in rats.
14. Jiang JY, Lyeth BG, Kapasi MZ, et al: Moderate hypothermia reduces blood-brain barrier disruption following traumatic brain injury in the rat. *Acta Neuropathol (Berl)* 1992; 84:495-500
15. Clifton GL, Jiang JY, Lyeth BG, et al: Marked protection by moderate hypothermia after experimental traumatic brain injury. *J Cereb Blood Flow Metab* 1991; 11:114-121
16. Dixon CE, Markgraf CG, Angileri F, et al: Protective effects of moderate hypothermia on behavioral deficits but not necrotic cavitation following cortical impact injury in the rat. *J Neurotrauma* 1998; 15:95-103
17. Busto R, Dietrich WD, Globus MY, et al: Small differences in intraschemic brain temperature critically determine the extent of ischemic neuronal injury. *J Cereb Blood Flow Metab* 1987; 7:729-738
18. Busto R, Dietrich WD, Globus MY, et al: Postischemic moderate hypothermia inhibits CA1 hippocampal ischemic neuronal injury. *Neurosci Lett* 1989; 101:299-304
19. Morikawa E, Ginsberg MD, Busto R, et al: Effect of moderate intraschemic hypothermia on brain focal injury following reversible middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 1991; 11(Suppl):S116
20. Marion DW, Penrod LE, Kelsey SF, et al: Treatment of traumatic brain injury with moderate hypothermia. *N Engl J Med* 1997; 336:540-546
21. Schwab S, Schwarz S, Spranger M, et al: Moderate hypothermia in the treatment of patients with severe middle cerebral artery infarction. *Stroke* 1998; 29:2461-2466
22. Shiozaki T, Sugimoto H, Taneda M, et al: Selection of severely head injured patients for mild hypothermia therapy. *J Neurosurg* 1998; 89:206-211
23. Lighthall JW, Goshgarian HG, and Pinderski CR: Characterization of axonal injury produced by controlled cortical impact. *J Neurotrauma* 1990; 7:65-76
24. Dixon CE, Clifton GL, Lighthall JW, et al: A controlled cortical impact model of traumatic brain injury in the rat. *J Neurosci Methods* 1991; 39:253-262
25. Kochanek PM, Marion DW, Zhang W, et al: Severe controlled cortical impact in rats: Assessment of cerebral edema, blood flow, and contusion volume. *J Neurotrauma* 1995; 12:1015-1025
26. Dixon CE, Lyeth BG, Povlishock JT, et al: A fluid percussion model of experimental brain injury in the rat. *J Neurosurg* 1987; 67:110-119
27. Feeney DM, Gonzalez A, Law WA: Amphetamine, haloperidol and experience interact to affect rate of recovery after motor cortex injury. *Science* 1982; 217:855-857
28. Dixon CE, Ma X, Marion DW: Effects of CDP-choline treatment on neurobehavioral deficits after TBI and on hippocampal and neocortical acetylcholine release. *J Neurotrauma* 1997; 14:161-169
29. Dietrich WD, Alonso O, Busto R, et al: Post-traumatic brain hypothermia reduces histopathological damage following concussive brain injury in the rat. *Acta Neuropathol* 1994; 87:250-258
30. Bramlett HM, Dietrich WD, Green EJ, et al: Chronic histopathological consequences of fluid-percussion brain injury in rats: Effects of post-traumatic hypothermia. *Acta Neuropathol (Berl)* 1997; 93:190-199
31. Dietrich WD, Busto R, Alonso O, et al: Intraschemic but not postischemic brain hypothermia protects chronically following global forebrain ischemia in rats. *J Cereb Blood Flow Metab* 1993; 13:541-549
32. Mansfield RT, Schiding JS, Hamilton RL, et al: Effects of hypothermia on traumatic brain injury in immature rats. *J Cereb Blood Flow Metab* 1996; 16:244-252
33. Cherian L, Robertson CS, Goodman JC: Secondary insults increase injury after controlled cortical impact in rats. *J Neurotrauma* 1996; 13:371-383
34. Clifton GL, Jiang JY, Lyeth BG, et al: Marked protection by moderate hypothermia after experimental traumatic brain injury. *J Cereb Blood Flow Metab* 1991; 11:114-121
35. Clifton GL, Allen S, Barrodale P, et al: A phase II study of moderate hypothermia in severe brain injury. *J Neurotrauma* 1993; 10:263-271
36. Zhao W, Alonso OF, Loo JY, et al: Influence of early posttraumatic hypothermia on local cerebral blood flow and glucose metabolism after fluid-percussion brain injury. *J Neurosurg* 1999; 90:510-519
37. Wald SL, Shackford SR, Fenwick J, et al: The effect of secondary insults on mortality and long-term disability after severe head injury in a rural region without a trauma system. *J Trauma* 1993; 34:377
38. Pigula FA, Wald SL, Shackford SR, et al: The effect of hypotension and hypoxia on children with severe head injuries. *J Pediatr Surg* 1993; 28:310-316
39. Jenkins LW, Moszynski K, Lyeth BG, et al: Increased vulnerability of the mildly traumatized rat brain to cerebral ischemia: the use of controlled secondary ischemia as a research tool to identify common of different mechanisms contributing to mechanical and ischemic brain injury. *Brain Res* 1989; 477:211-224

10: G

X 6

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.

## AWARDS

(1999) Women in Neurotrauma Award to Dr. Kimberly Statler for her presentation to the 1999 Meeting of the National Neurotrauma Society entitled, "Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats."

(2000) Educational Scholarship to Dr. Kimberly Statler from the Society of Critical Care Medicine for her abstract entitled, "Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats."

## (9) CONCLUSIONS

1. Based on the important finding in this study where the use of fentanyl in our CCI model produced deleterious effects on outcome after TBI, animal models should utilize clinically relevant sedative/analgesic treatments. The beneficial mechanisms of isoflurane (possibly promotion of cerebral blood flow or reduction of excitotoxicity) should be investigated for the development of novel treatments. Isoflurane anesthesia likely provides powerful beneficial effects at or near the time of injury. Sedative agents with anti-excitotoxic and/or cerebral blood flow promoting actions similar to isoflurane may thus represent better alternatives early after TBI. **Narcotics may not be the optimal sedative/analgesic early after TBI. Better field therapy than narcotics must be developed.**
2. Based on studies in rats using the CCI model, we have demonstrated tangible risk to aggressive, indiscriminate hyperventilation early after injury—specifically—augmentation of neuronal death in selectively vulnerable brain regions. This suggests that aggressive hyperventilation should not be indiscriminately used in the field

treatment of TBI, rather it should be applied if there are signs and/or symptoms of herniation. Mild hyperventilation (used in our control group) or normocapnia may be preferable.

3. Based on our studies in rats, hypothermia, although showing some beneficial effects, particularly early after TBI (such as a reduction in DNA damage, etc), may have some deleterious effects which result in only modest overall beneficial effects on long-term outcome. This is particularly true in the setting of severe injury (such as TBI plus secondary hypoxemic insults) where it is possible that there is little to gain except side effects. Also based on our narcotic (fentanyl) vs isoflurane study and our multi-sedation study, hypothermia should be re-examined in future studies with narcotic anesthesia, since beneficial effects of isoflurane may be masking any benefit from hypothermia.

## (10) REFERENCES

1. Forbes ML, Clark RSB, Dixon CE, Graham SH, Marion DW, DeKosky ST, Schiding JK, Kochanek PM: Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact. *Journal of Neurosurgery* 88:549-556, 1998.
2. Alexander HL, Robertson CL, Dixon CE, Clark RSB, Graham SH, Safar PJ, Kochanek PM: Vertical Versus Angled Controlled Cortical Impact in Rats. *Journal of Neurotrauma* 15:854, 1998.
3. Clark RSB, Robertson CL, Dixon CE, Alexander HL, Graham SH, Safar PJ, Kochanek PM: Effect of Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Journal of Neurotrauma* 15:864, 1998.
4. Robertson CL, Clark R, Dixon CE, Graham S, Alexander H, Wisniewski S, Marion D, Safar P, Kochanek P: No Long-Term Benefit from Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Critical Care Medicine* 27(1):A52, 1999.
5. Robertson CL, Clark RSB, Dixon CE, Graham ST, Alexander HL, Wisniewski SR, Marion DW, Safar PJ, Kochanek PM: No Long Term Benefit from Hypothermia after Severe Traumatic Brain Injury with Secondary Insult in Rats. *Critical Care Medicine* (in press).
6. Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.
7. Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.



8. Kochanek PM, Safar P, Marion DW, Tisherman SA, Clark RSB, DeKosky ST: Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock: From Mild Cooling To Suspended Animation In: Hypothermia in Trauma: Deliberate or Accidental. ITACCS Monograph, CE Smith and CM Grande, Editors, Baltimore, Maryland, 1997, pp 17-20.
9. Ruppel RA, Kochanek PM, Dixon CE, Alexander HL, Graham SH, Clark RSB, Wisniewski SR, Marion DW, Safar PJ: MK-801 improves functional outcome in rats after controlled cortical impact. *Journal of Neurotrauma* 16:986, 1999.
10. Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Society for Neuroscience Abstract* 25:537, 1999.
11. Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.
12. Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.
13. Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Graham SH, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome vs. fentanyl after traumatic brain injury in rats. *Journal of Neurotrauma* (in press).

## **(11) APPENDIX**

### **1. 1997 Report**

### **2. 1998 Report**

### **3. Curriculum Vitae**

### **4. Manuscripts:**

Forbes ML, Clark RSB, Dixon CE, Graham SH, Marion DW, DeKosky ST, Schiding JK, Kochanek PM: Augmented neuronal death in CA3 hippocampus following hyperventilation early after controlled cortical impact. *Journal of Neurosurgery* 88:549-556, 1998.

Kochanek PM, Safar P, Marion DW, Tisherman SA, Clark RSB, DeKosky ST: Therapeutic Hypothermia After Traumatic Brain Injury or Hemorrhagic Shock: From Mild Cooling To Suspended Animation In: Hypothermia in Trauma: Deliberate or Accidental. ITACCS Monograph, CE Smith and CM Grande, Editors, Baltimore, Maryland, 1997, pp 17-20.

Robertson CL, Clark RSB, Dixon CE, Graham ST, Alexander HL, Wisniewski SR, Marion DW, Safar PJ, Kochanek PM: No Long Term Benefit from Hypothermia after Severe Traumatic Brain Injury with Secondary Insult in Rats. *Critical Care Medicine* (in press).

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Graham SH, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome vs. fentanyl after traumatic brain injury in rats. *Journal of Neurotrauma* (in press).

#### **5. Abstracts:**

Alexander HL, Robertson CL, Dixon CE, Clark RSB, Graham SH, Safar PJ, Kochanek PM: Vertical Versus Angled Controlled Cortical Impact in Rats. *Journal of Neurotrauma* 15:854, 1998.

Clark RSB, Robertson CL, Dixon CE, Alexander HL, Graham SH, Safar PJ, Kochanek PM: Effect of Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Journal of Neurotrauma* 15:864, 1998.

Robertson CL, Clark R, Dixon CE, Graham S, Alexander H, Wisniewski S, Marion D, Safar P, Kochanek P: No Long-Term Benefit from Hypothermia After Severe Traumatic Brain Injury with Secondary Hypoxemia in Rats. *Critical Care Medicine* 27(1):A52, 1999.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Society for Neuroscience Abstracts* 24:252, 1998.

Whalen M, Chen M, Clark R, Jin K, Kochanek P, Marion D, Graham S: DNA Damage is Temperature Dependent Early After Traumatic Brain Injury in Rats. *Critical Care Medicine* 27:A51, 1999.

Ruppel RA, Kochanek PM, Dixon CE, Alexander HL, Graham SH, Clark RSB, Wisniewski SR, Marion DW, Safar PJ: MK-801 improves functional outcome in rats after controlled cortical impact. *Journal of Neurotrauma* 16:986, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Society for Neuroscience Abstract* 25:537, 1999



Statler KD, Kochanek PM, Dixon CE, Alexander H, Warner DS, Clark RSB, Wisniewski S, Graham SH, Jenkins LW, Ma X, Marion DW, Safar P: Fentanyl versus isoflurane anesthesia: Effect on outcome after traumatic brain injury in rats. *Journal of Neurotrauma* 16:965, 1999.

Statler KD, Kochanek PM, Dixon CE, Alexander HL, Warner DS, Clark RSB, Wisniewski SR, Jenkins LW, Marion DW, Safar PJ: Isoflurane improves long-term neurologic outcome compared to fentanyl after traumatic brain injury in rats. *Critical Care Medicine* 27:A38, 1999.

Alexander H, Statler KD, Kochanek PM, Dixon CE, Clark RSB, Vagni V, Graham SH, Marion DW, Safar P: Post-traumatic application of seven different anesthetic agents in the controlled cortical impact model of traumatic brain injury: effect on outcome. *Journal of Neurotrauma* (in press).

**(12) BINDING (N/A)**

**(13) FINAL REPORTS**

**a) Bibliography of all publications and abstracts (see appendix)**

**b) List of personnel receiving pay from the research effort**

Patrick M. Kochanek, M.D.

Peter Safar, M.D.

Henry Alexander

Scott Heineman

Marci Provins

Linda Amick

ISOFLURANE IMPROVES LONG-TERM NEUROLOGIC OUTCOME VS  
FENTANYL AFTER TRAUMATIC BRAIN INJURY IN RATS

Kimberly D. Statler, M.D.,<sup>1,6</sup> Patrick M. Kochanek, M.D.,<sup>1,2,6</sup> C. Edward Dixon,  
Ph.D.,<sup>3,6</sup> Henry L. Alexander,<sup>1,6</sup> David S. Warner, M.D.,<sup>8</sup> Robert S.B. Clark,  
M.D.,<sup>1,2,6</sup> Stephen R. Wisniewski, Ph.D.,<sup>4</sup> Steven H. Graham, M.D., Ph.D.,<sup>7</sup>  
Larry W. Jenkins, Ph.D.,<sup>1,6</sup> Donald W. Marion, M.D.,<sup>3</sup> Peter J. Safar, M.D.<sup>1,6</sup>

Departments of Anesthesiology and Critical Care Medicine,<sup>1</sup> Pediatrics,<sup>2</sup>  
Neurosurgery,<sup>3</sup> Epidemiology and Public Health,<sup>4</sup> and Neurology,<sup>5</sup> University of  
Pittsburgh School of Medicine, Pittsburgh, Pennsylvania  
Safar Center for Resuscitation Research,<sup>6</sup> Pittsburgh, Pennsylvania  
Geriatric Research Educational and Clinical Center,  
Veterans Administration Pittsburgh Health Center,<sup>7</sup>  
Department of Anesthesia, Duke University,<sup>8</sup> Durham, North Carolina

**Running Title:** Isoflurane vs fentanyl after TBI in rats

Please direct correspondence to:

Patrick M. Kochanek, M.D.

Safar Center for Resuscitation Research

3434 Fifth Avenue, Suite 201

Pittsburgh, PA 15260

Phone: (412) 383-1900

Fax: (412) 624-0946

Email: [kochanekpm@anes.upmc.edu](mailto:kochanekpm@anes.upmc.edu)

## **Abstract**

Despite routine use of fentanyl in patients after traumatic brain injury (TBI), it is unclear if it is the optimal sedative/analgesic agent. Isoflurane is commonly used in experimental TBI. We hypothesized that isoflurane would be neuroprotective vs fentanyl after TBI. Rats underwent controlled cortical impact (CCI) and received 4 h of N<sub>2</sub>O:O<sub>2</sub> (2:1) and either fentanyl (10 µg/kg iv bolus, 50 µg/kg/h infusion) or isoflurane (1% by inhalation) with controlled ventilation. Shams underwent identical preparation, without CCI. Functional outcome (beam balance, beam walking, Morris water maze [MWM] tasks) was assessed over 20 days. Lesion volume and hippocampal neuron survival were quantified on d 21. Additional rats underwent identical CCI and anesthesia with intracranial pressure (ICP) monitoring, and brain water content was assessed.

Motor and MWM performances were better in injured rats treated with isoflurane vs fentanyl ( $p < 0.05$ ). CA1 hippocampal damage was attenuated in isoflurane-treated rats ( $p < 0.05$ ). Fentanyl-treated rats had higher mean arterial blood pressure after injury ( $p < 0.05$ ); however, ICP and brain water were similar between groups.

Isoflurane improved functional outcome and attenuated damage to CA1 vs fentanyl in rats subjected to CCI. Isoflurane may be neuroprotective by augmenting cerebral blood flow and/or reducing excitotoxicity, not by reducing

ICP or brain water content. Alternatively, fentanyl may be detrimental.

Isoflurane may mask beneficial effects of novel agents tested in TBI models.

Additionally, fentanyl may not be optimal early after TBI in humans.

**Key words:** sedation, analgesia, anesthesia, head injury, narcotics, opioids

## **Introduction**

In current clinical practice, opioids are routinely administered after traumatic brain injury (TBI). Fentanyl is one of the first-line agents because of its short half-life and low incidence of hypotension. Despite standard clinical use, it remains unclear if fentanyl represents the optimal sedative/analgesic agent in the acute period following TBI. Unlike the clinical arena, opioids are rarely used in experimental TBI. In fact, most models of TBI use isoflurane or pentobarbital anesthesia.

Much of the study of opioids in TBI has focused on the actions of endogenous opiates, such as dynorphin, and specific opiate receptor effects (Hayes et al., 1990; Lyeth et al., 1993, 1995; McIntosh et al., 1987; Vink et al., 1990). Although mu receptor agonists, such as morphine and fentanyl, have been shown to have some beneficial effects after central nervous system injury (Lyeth et al., 1993, 1995), recent studies in cerebral ischemia and focal cryogenic lesion suggest that isoflurane may be neuroprotective compared to fentanyl (Miura et al., 1998; Murr et al., 1993, 1995; Soonthon-Brant et al., 1999). In rats subjected to global cerebral ischemia, isoflurane reduced neuronal damage and improved motor function compared to fentanyl (Miura et al., 1998). Similarly, after focal ischemia, rats anesthetized with isoflurane had smaller infarct volumes than those receiving fentanyl. Lesion volumes in rats treated with fentanyl were similar to

those in unanesthetized control rats (Soonthon-Brant et al., 1999). Isoflurane has been reported to enhance post-insult cerebral blood flow (CBF), produce widespread increases in brain surface  $PO_2$  and reduce edema in a rabbit model of focal cryogenic lesion. Conversely, both CBF and regional  $PO_2$  were decreased, and edema was increased, in rabbits anesthetized with fentanyl (Murr et al., 1993, 1995). Using a fluid-percussion model in cats, DeWitt et al, addressed the question of whether fentanyl was detrimental in TBI and found that fentanyl produced no adverse effects compared to vehicle. However, in that study, fentanyl was administered to cats already anesthetized with isoflurane (Bedell et al., 1998).

To our knowledge, isoflurane has not been directly compared to fentanyl in a contemporary model of TBI with long-term functional outcome and histologic assessment. We hypothesized that isoflurane would be neuroprotective compared to fentanyl when administered before and early after TBI. To test our hypothesis, we directly compared fentanyl and isoflurane anesthesia in a controlled cortical impact (CCI) model of TBI.

## **Materials and Methods**

Virus-free, mature male (280 - 400g, total n=51) Sprague-Dawley rats were used in this study. The rats had free access to food and water before and

after surgery. All studies were approved by the University of Pittsburgh Animal Care and Use Committee. All surgical procedures were performed using aseptic technique.

### ***Outcome Protocol***

Rats were initially anesthetized with N<sub>2</sub>O:O<sub>2</sub> (2:1) and 4% isoflurane (IsoFlo, Abbott Laboratories, North Chicago, IL) via a nose cone and then endotracheally intubated with a 14-gauge angiocatheter and mechanically ventilated. Anesthesia was maintained for the duration of surgical preparation with 2 - 2.5% isoflurane and N<sub>2</sub>O:O<sub>2</sub> (2:1). Femoral venous and arterial vessels were cannulated for continuous blood pressure measurement, blood sampling, and administration of medications. Pancuronium bromide (0.1 mg/kg/h, Elkins-Sinn, Cherry Hill, NJ) was given intravenously for muscle relaxation. A rectal probe was inserted to monitor core temperature. The rat was then placed in a stereotaxic frame (David Kopf, Tujunga, CA) and a left parietal craniotomy (7mm x 8mm) was performed using a high-speed dental drill. The dura and bone flap were left in place until immediately before CCI. A burr hole was drilled into the left frontal bone for temperature probe (2.28-mm outside diameter, Physiotemp Corp., Clifton, NJ) placement into the frontal lobe. Continuously monitored physiologic parameters included mean arterial blood pressure (MAP) and rectal and brain

temperatures. Parameters monitored intermittently included blood glucose, hematocrit, and arterial blood gas samples, which were assessed every 15 minutes for the initial hour and every 30 minutes thereafter. Throughout the experiment, PaCO<sub>2</sub> was controlled at 35 - 45 mm Hg. This protocol produced a PaO<sub>2</sub> of greater than 70 mm Hg in all preparations. Both brain and rectal temperatures were maintained at 37.0 ± 0.5 °C.

Rats were allowed to stabilize for 5 min after completion of surgical preparation and then randomized to receive either fentanyl or isoflurane anesthesia. In the fentanyl group ( $n = 9$ ), isoflurane was discontinued and 10 µg/kg of fentanyl (50 µg/ml, Elkins-Sinn, Cherry Hill, NJ) was administered intravenously, followed by a continuous intravenous infusion at 50 µg/kg/h. In the isoflurane group ( $n = 9$ ), inspired isoflurane concentration was reduced to 1%, and normal saline, the vehicle for fentanyl-treated rats, was administered to match the volume received by fentanyl infusion. Both anesthetic groups continued to receive N<sub>2</sub>O:O<sub>2</sub> (2:1). After 30 min of equilibration, TBI was induced by CCI using a pneumatic-driven piston device that has been shown (with isoflurane anesthesia) to deliver a reliable and reproducible degree of injury with a mortality rate of less than 5% (Dixon et al., 1991; Kochanek et al., 1995). In pilot studies comparing isoflurane and fentanyl using our standard CCI model (6-mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.5-mm deformation depth),



all of the fentanyl-treated rats developed pulmonary edema and died early after injury. Thus, to compare the effect of isoflurane vs fentanyl on long-term outcome in our model, our standard injury was reduced (6 mm tip, 4 m/s velocity, 50 msec duration of deformation and 2.0-mm deformation depth). After CCI, the bone flap was replaced and sealed with dental cement, and the scalp incision was closed. Anesthesia was continued for 4 h in the isoflurane group and 3.5 h in the fentanyl group to facilitate similar extubation times (Miura et al., 1998). At the end of the anesthetic period, rats received 100% oxygen, were allowed to awaken and resume spontaneous breathing, and were then extubated and returned to their cages. Sham rats underwent identical preparation and anesthesia, but no CCI ( $n = 6$  per anesthetic group).

Motor function, including beam balance and beam walking tasks, was tested by an observer blinded to group assignment on days 1 - 5 after injury (Dixon et al., 1987, 1991). Briefly, in the beam balance task, the rat was placed on a suspended, narrow wooden beam (1.5-cm wide) and the time that the rat remained on the beam was recorded (up to 60 sec). For beam walking, the rat was placed at one end of the beam and a dark, quiet chamber was located at the other end. An adverse stimulus of loud white noise was applied and the time for the rat to escape across the beam into the chamber was recorded (up to 60 sec). Rats were

trained with three trials before CCI or sham injury, which also served as baseline values.

Morris Water Maze (MWM) testing was performed using an acquisition paradigm on days 14 - 20 after injury (Hamm et al., 1992). Briefly, on post-injury days 14 - 18, the rat was placed into a pool (2-m diameter) and required to locate a hidden platform in order to escape the water. On post-injury days 19 and 20, the platform was raised so that the surface was visible 5-cm above the water level. Latency to find the platform was used to compare performances. Swim speed was measured on post-injury day 20 to insure that rats in all experimental groups exhibited equal motivation and motor function.

Lesion volume and hippocampal neuronal survival were assessed on day 21 after injury (Clark et al., 2000; Kochanek et al., 1995). Rats were re-anesthetized and then perfused with heparinized saline followed by 4% paraformaldehyde. Brains were removed, post-fixed and cryoprotected. Serial coronal sections (10- $\mu$ m) were made at 1-mm intervals through the entire brain. Sections were mounted on slides and stained with cresyl violet. The areas of both tissue loss and the entire uninjured hemisphere were determined by an observer blinded to experimental group using an image analysis system (MCID, Imaging Research, St. Catharines, Ontario, Canada). Lesion volume was reported in cubic mm and as a percentage of uninjured hemisphere. Surviving hippocampal

neurons were counted under 400X magnification in the entire anatomic CA1 and CA3 hippocampal regions in a coronal section taken 5-mm from the occiput by an observer blinded to experimental group (KS). Neuronal counts were reported as the mean number of surviving neurons per 400X field.

### ***ICP Protocol***

Based both on results of the outcome protocol showing that fentanyl-treated rats had higher MAP throughout the experiment and on a recent report that increased MAP may exacerbate injury after TBI (Kroppenstedt et al., 1999), ICP and brain water were monitored in a separate cohort of rats ( $n = 9$  per anesthetic group) subjected to either fentanyl or isoflurane anesthesia and CCI in an identical paradigm to that used to assess functional outcome.

Surgical preparation, randomization, anesthetic administration, and CCI were identical to the outcome protocol, with minor exceptions. Specifically, an intraparenchymal ICP monitor (Codman microsensor transducer, outer diameter 1.0-mm, Johnson and Johnson, Raynham, MA) was inserted through a burr hole in the frontal bone into the contralateral (right) frontal cortex at the time of craniotomy. After CCI, anesthesia was continued and ICP was monitored for 4 h in both anesthetic groups. Cerebral perfusion pressure (CPP) was calculated as the difference between MAP and ICP. Rats were killed by decapitation at the end

of the anesthetic period. Brains were immediately removed and a 3-mm coronal slice was made through the center of the contusion using a rat brain slicer. Percent brain water was determined in the coronal slice using the wet-dry weight method (Kochanek et al., 1995). The section was weighed immediately, dried in an oven at 110° C for 48 hours and then reweighed. Brain water content was determined in both the injured and the homologous region of the uninjured hemispheres.

As an additional control, a separate cohort of rats ( $n = 3$ ) was subjected to CCI and allowed to recover without anesthesia. These rats were prepared for CCI under isoflurane anesthesia as described, allowed to recover a tail-pinch response and then subjected to CCI. MAP was monitored via an indwelling femoral arterial catheter for 4 h during recovery without anesthesia.

### ***Statistical Analysis***

Physiological parameters (PaCO<sub>2</sub>, PaO<sub>2</sub>, glucose, hematocrit, MAP, ICP, CPP) and beam balance, beam walking, and MWM latencies were assessed by two-way analysis of variance for repeated measures. Tukey's test was used in all post-hoc comparisons involving repeated measures. Swim speed, brain water content, and hippocampal neuronal survival were compared by one-way analysis of variance. The Student-Newman-Keuls test was used for post-hoc comparisons

after one-way analysis of variance. Time to extubation and lesion volume were compared between treatment groups using unpaired student's t-test. A  $p$  value  $< 0.05$  was considered statistically significant. Values are expressed as mean  $\pm$  SEM.

## Results

### *Outcome Protocol*

Average time to extubation did not differ after injury between isoflurane and fentanyl treatment groups ( $269 \pm 21$  min vs  $275 \pm 45$  min,  $p = 0.29$ ). Physiologic values, including  $\text{PaCO}_2$ ,  $\text{PaO}_2$ , blood glucose and hematocrit did not differ between anesthetic groups. In contrast, MAP was higher in injured rats treated with fentanyl compared to their isoflurane counterparts ( $p < 0.05$ ) during the entire post-trauma period (Figure 1). Similarly, MAP was higher in shams treated with fentanyl vs isoflurane ( $p < 0.05$ ) during the entire duration of anesthesia (Figure 1). Fentanyl-treated rats had a MAP of approximately 150 mm Hg compared to approximately 105 mm Hg in the isoflurane groups.

Rats anesthetized with isoflurane performed better on beam balance and beam walking tasks after TBI compared to their fentanyl counterparts ( $p < 0.05$ , Figure 2). Following injury, isoflurane-anesthetized rats also performed better than their fentanyl-treated counterparts during MWM testing with a hidden

platform ( $p < 0.05$ , Figure 3). Motor and MWM performances did not differ between sham groups. All experimental groups showed improved MWM performance during visible (vs hidden) platform testing ( $p < 0.05$  for all groups) and had similar swim speeds (Figure 3), indicating that all rats had similar motivation and motor ability during MWM testing. However, performance on the visible platform paradigm of the MWM was better in isoflurane vs fentanyl treated rats after TBI ( $p < 0.05$ , Figure 3). This suggests that the difference in MWM performance may not be solely attributable to cognitive deficits.

Lesion volume, expressed as  $\text{mm}^3$  or as percent of uninjured hemisphere, at 21 days after TBI did not differ significantly between isoflurane and fentanyl treatment groups (Figure 4A). In contrast, neuronal counts in the injured CA1 hippocampus were markedly greater in isoflurane-treated rats ( $p < 0.05$ , Figure 4B). Neuronal counts in the injured CA3 hippocampus did not differ significantly between treatment groups (Figure 4C). In the uninjured hemisphere, neuronal counts in both CA1 and CA3 hippocampus did not differ between isoflurane- and fentanyl-treated rats, and were similar to shams (data not shown).

### ***ICP Protocol***

Physiologic values, including  $\text{PaCO}_2$ ,  $\text{PaO}_2$ , glucose and hematocrit, did not differ between anesthetic groups. As in the outcome protocol, MAP was

higher in rats treated with fentanyl compared to their isoflurane counterparts ( $p < 0.05$ , Figure 5A). ICP was similar in both anesthetic groups; however, there was a trend toward higher ICP in rats anesthetized with isoflurane by 3 - 4 h after TBI (Figure 5B). This strongly suggests that the higher MAP in fentanyl vs isoflurane treated rats did not exacerbate intracranial hypertension. As expected from the difference in MAP, CPP was significantly higher in the fentanyl treatment group (Figure 5C). Brain water content, assessed at 4 h after injury, was higher in the injured vs uninjured hemisphere ( $p < 0.05$ ) for both anesthetic groups (Figure 6). However, brain water in either the injured or uninjured hemisphere did not differ between isoflurane- and fentanyl-treated rats, indicating that edema was not exacerbated in the fentanyl group.

Average MAP during the 4 h post-injury observation period did not differ significantly between fentanyl-treated rats and those recovering from CCI without anesthesia ( $157 \pm 18.6$  mm Hg vs  $147 \pm 12.3$  mm Hg). In contrast, isoflurane-anesthetized rats had lower MAP ( $105 \pm 16.5$  mm Hg) compared to both fentanyl-treated rats and rats recovering without anesthesia ( $p < 0.05$  vs both groups, one-way ANOVA).

## Discussion

Isoflurane anesthesia administered to rats subjected to CCI improved performance on both motor and MWM tasks and attenuated damage to CA1 hippocampus after TBI. Although fentanyl-treated rats had higher MAP and CPP than their isoflurane counterparts, this was not accompanied by increased ICP or brain water during the first 4 h after TBI. This suggests that the improved functional outcome in rats anesthetized with isoflurane may be a direct result of either beneficial actions of isoflurane, and/or detrimental effects of fentanyl.

The pathophysiology of TBI includes a primary injury caused by the mechanical disruption of tissue and various secondary injuries mediated, at least in part, by post-insult hypoperfusion, ischemia, and excitotoxicity (Bryan et al., 1995; Hendrich et al., 1999; Hovda et al., 1995; Kochanek et al., 1995; McIntosh, 1993). Isoflurane anesthesia may be neuroprotective vs fentanyl by decreasing excitotoxicity and/or augmenting cerebral blood flow.

Following TBI, interstitial levels of excitatory amino acids (EAAs), such as glutamate, are increased due to direct tissue injury and secondary ischemia. EAAs stimulate NMDA, AMPA/kainate, and metabotropic receptors, leading to neuronal membrane depolarization, cellular swelling, calcium influx, and ultimately, neuronal death (McIntosh et al., 1999). Although models of cerebral ischemia have shown conflicting effects of isoflurane on glutamate levels in blood



and brain interstitial fluid (Patel et al., 1995; Stover et al., 1999), isoflurane has been shown to inhibit glutamate receptors, reduce NMDA-mediated calcium influx and delay neuronal injury induced by cerebral ischemia (Bickler et al., 1994; Patel et al., 1998).

To attribute the potential neuromodulatory actions of isoflurane on neuronal injury to only glutamate toxicity (Bickler et al., 1994) and glutamate signaling transduction (Dildy-Mayfield et al., 1996), however is an oversimplification since isoflurane has many neural actions that could contribute to neuroprotection. Some include the inhibition of some voltage sensitive potassium channels (Kamatchi et al., 1999), the activation of specific receptor-coupled and voltage sensitive potassium channels (Magyar and Szabo, 1996; Winegar et al., 1996; Zorn et al., 1993), the uncoupling of muscarinic receptors (Anthony et al., 1989; Minami et al., 1997; Nietgen et al., 1998), the enhancement of GABA A channels (Lin et al., 1992), and the reduction of intracellular calcium stores and inhibition of IP3 sensitive intracellular calcium release (Hossain et al., 1991). Potential detrimental actions such as the enhancement of NMDA linked nNOS activation have also been documented (Rengasamy et al., 1997); however, the potential beneficial actions of isoflurane on excitotoxic cascades far outweigh the potential detrimental actions.

Additionally, isoflurane is a potent cerebral vasodilator. Studies of CBF after experimental TBI have shown significant reductions in both local and global CBF early (0.5 – 4 h) after injury (Bryan et al., 1995; Hendrich et al., 1999). Effects are greatest near the impact site, but global reductions in CBF are seen as well (Hendrich et al., 1999). In clinical studies, early post-traumatic hypoperfusion has been strongly correlated with poor outcome (Bouma and Muizelaar, 1992; Marion et al., 1991). Although the effects of fentanyl on CBF have been subjected to limited study, Safo et al (1985), have reported that CBF was markedly reduced in rats treated with fentanyl (100 µg/kg iv) vs control rats anesthetized with N<sub>2</sub>O. In addition, using perfusion MRI in normal rats anesthetized with doses identical to those used in this study, we have shown that CBF is 2 - 3 times higher in rats treated with isoflurane vs fentanyl (unpublished data). By promoting CBF, isoflurane may help attenuate post-traumatic hypoperfusion, reducing secondary injury and improving recovery. The selective neuroprotection of CA1, but not CA3, hippocampus in isoflurane- vs fentanyl-treated rats is consistent with this concept. Additionally, another CBF promoting strategy, L-arginine, has recently been shown to improve outcome after TBI (Cherian et al., 1998; DeWitt et al., 1997). Augmentation of CBF with an associated increase in cerebral blood volume therefore offers a potential explanation for both improved functional and histological outcome and the

tendency toward higher ICP and brain water seen in isoflurane- vs fentanyl-treated rats in this study. Indeed, the combination of CBF promotion and reduced excitotoxicity may be particularly beneficial.

Alternatively, fentanyl may be detrimental after TBI. Opioids generally suppress neuronal excitability, however, mu receptor agonists, such as fentanyl, may contribute to hippocampal neuron excitation (Bradley and Brookes, 1984; Neumaier and Chavkin, 1986; Neumaier et al., 1988). In fact, high-dose fentanyl (25 - 100  $\mu\text{g/kg}$  in humans and 400  $\mu\text{g/kg}$  in rats) has been associated with subcortical seizures (Faden et al., 1985; Kearse et al., 1993; McIntosh et al., 1986; Ohta et al., 1995; Safo et al., 1985; Tempelhoff et al., 1992). Although fentanyl exhibits low affinity for kappa receptors (Ohta et al., 1995), kappa receptor antagonists have been shown to improve both neurological outcome after spinal cord injury and CBF after fluid-percussion injury in cats (Faden et al., 1985; McIntosh et al., 1986).

Other factors may contribute to improved outcome in isoflurane- vs fentanyl-treated rats. These include the disparity in both MAP and CPP and possible differences in the depth of anesthesia between treatment groups. Although increased MAP can have detrimental effects after TBI, in our study, ICP and brain water were not significantly different in isoflurane vs fentanyl treatment groups at the end of the 4 h treatment period. This suggests that the higher MAP

associated with fentanyl vs isoflurane anesthesia had no acute detrimental effects on intracranial hypertension or brain edema. Additionally, MAP did not differ significantly between rats treated with fentanyl and those allowed to recover without anesthesia. The values for MAP observed in both isoflurane and fentanyl groups were within the reported range of cerebral autoregulation (50 - 170 mm Hg) for normotensive rats (Hernandez et al., 1978; Hoffman et al., 1991; Zaharchuk et al., 1999). The disparity in MAP is therefore unlikely to have contributed importantly to the observed difference in functional outcome. Although a recent study using the CCI model in rats suggests that increased MAP and CPP may exacerbate injury after TBI, in that study, systemic hypertension was induced by large doses of dopamine (Kroppenstedt et al., 1999). Higher doses of dopamine may have produced detrimental effects after injury which were independent of blood pressure (Beaumont et al., 1999). Additionally, our study does not address the important detrimental effect of hypotension following clinical TBI, particularly with multiple trauma. The blood pressure supporting effects of fentanyl or its derivatives (vs other sedative agents) may be beneficial in this section (Tipps et al., 2000).

Although comparison of anesthetic depth between inhalation and intravenous agents is difficult, differences in anesthetic depth are unlikely to explain the observed difference in outcome seen between isoflurane- and fentanyl-

treated rats. The dose of isoflurane used (1% by inhalation), in combination with N<sub>2</sub>O:O<sub>2</sub> (2:1) represents approximately 1.2 minimal alveolar concentration (Hansen et al., 1989). The dose of fentanyl falls in the range between the ED<sub>50</sub> for purposeful movement and complete blockade of this response (Kissin et al., 1983) and is similar to standard doses used in rat and cat models of central nervous system injury (Bedell et al., 1998; Miura et al., 1998; Murr et al., 1993, 1995; Soonthon-Brant et al., 1999). Additionally, fentanyl-treated rats did not exhibit signs of increased stress vs isoflurane-treated rats, such as higher blood glucose, suggesting that anesthesia was adequate. However, there is a massive catecholamine surge immediately after TBI (Rosner et al., 1984), and it is likely that this response is more effectively blunted by isoflurane than fentanyl (Mackensen et al., 1999). Finally, rats in both treatment groups emerged from anesthesia similarly in our paradigm. After discontinuation of fentanyl, the rats were fully alert and exhibited similar activity to isoflurane-treated rats following extubation.

The theoretical advantages of isoflurane vs fentanyl are compelling; however, explanations for the observed improvement in neurologic outcome after TBI in isoflurane-anesthetized rats remain to be determined. What has become clearer is that anesthetic agents may have considerable impact upon outcome following TBI.

The results of this study suggest two important potential ramifications. First, isoflurane may not represent the optimal anesthetic in experimental TBI since it may mask potential benefits of novel therapies. Second, despite common use, fentanyl may not be the optimal sedative/analgesic agent to administer to patients in the acute phase after severe head injury. Although we do not suggest that isoflurane represents a clinically applicable therapy for the initial stabilization and treatment of patients after TBI, further study of the mechanistic differences between isoflurane and fentanyl anesthesia is warranted. Defining the factors responsible for improved outcome with isoflurane may help to direct the clinical application of more optimal sedative/analgesic agents and possibly to identify novel therapies. Finally, more comprehensive comparisons of clinically relevant sedative/analgesic agents are needed in experimental TBI.

**Acknowledgement:** We thank the US Army DAMD 17-97-1-7009 for generous support of this project. We thank Marci Provins for assistance with preparation of the manuscript. This study was presented, in part, at the 1999 meeting of the National Neurotrauma Society. Dr. Statler received the *1999 Women in Neurotrauma Research Award* for this work.

## References

- ANTHONY, B.L., DENNISON, R.L., and ARONSTAM, R.S. (1989). Disruption of muscarinic receptor-G protein coupling is a general property of liquid volatile anesthetics. *Neurosci. Lett.* **99**, 191-196.
- BEAUMONT, A., MARMAROU, A., FATOUROS, P., et al. (1999). The effects of dopamine on cerebral edema formation in two models of traumatic brain injury. *Proceedings of the Eleventh International Symposium on Brain Oedema and Mechanisms of Cellular Injury Scientific Programme and Abstracts*, p 42.
- BEDELL, E.A., DEWITT, D.S., and PROUGH, D.S. (1998). Fentanyl infusion preserves cerebral blood flow during decreased arterial blood pressure after traumatic brain injury in cats. *J. Neurotrauma* **15**, 985-992.
- BICKLER, P.E., BUCK, L.T., and HANSEN, B.M. (1994). Effects of isoflurane and hypothermia on glutamate receptor-mediated calcium influx in brain slices. *Anesthesiology* **81**, 1461-1469.
- BOUMA, G.J. and MUIZELAAR, J.P. (1992). Cerebral blood flow, cerebral blood volume, and cerebrovascular reactivity after severe head injury. *J. Neurotrauma* **9**, S333-S348.

- BRADLEY, P.B. and BROOKES, A. (1984). A microiontophoretic study of the actions of mu-, delta-, and kappa-opiate receptor agonists in the rat brain. *Br. J. Pharmacol.* **83**, 763-772.
- BRYAN, R.M., CHERIAN, L., and ROBERTSON, C. (1995). Regional cerebral blood flow after controlled cortical impact injury in rats. *Anesth. Analg.* **80**, 687-695.
- CHERIAN, L., CHACKO, G., GOODMAN, J.C., and ROBERTSON, C.S. (1998). L-arginine attenuates histological damage after controlled cortical impact injury in rats. *J. Neurotrauma* **10**, 863.
- CLARK, R.S.B., KOCHANIEK, P.M., WATKINS, S.C., et al. (2000). Caspase-3 mediated neuronal death after traumatic brain injury in rats. *J. Neurochem.* **74**, 1-14.
- DEWITT, D.S., SMITH, T.G., DEYO, D.J., et al. (1997). L-arginine and superoxide dismutase prevent or reverse cerebral hypoperfusion after fluid-percussion traumatic brain injury. *J. Neurotrauma* **14**, 223-233.
- DILDY-MAYFIELD, J.E., EGER, E.I., and HARRIS, R.A. (1996). Anesthetics produce subunit-selective actions on glutamate receptors. *J. Pharmacol. Exp. Ther.* **276**, 1058-1065.



- DIXON, C.E., CLIFTON, G.L., LIDTHALL, L.W., YAGHAMAI, A.A., and HAYES, R.L. (1991). A controlled cortical impact model of traumatic brain injury in the rat. *J. Neurosci. Meth.* **39**, 253-262.
- DIXON, C.E., LYETH, B.G., POVLISHOCK, J.T., et al. (1987). A fluid percussion model of experimental brain injury in the rat. *J. Neurosurg.* **67**, 110-119.
- FADEN, A.I., KNOBLACH, S., MAYS, C., and JACOBS, T.P. (1985). Motor dysfunction after spinal cord injury is mediated by opiate receptors. *Peptides* **6**, 15-17.
- HAMM, R.J., DIXON, C.E., GBADEBO, D.M., SINGHA, A.K., JENKINS, L.W., LYETH, B.G., and HAYES, R.L. (1992). Cognitive deficits after traumatic brain injury produced by controlled cortical impact. *J. Neurotrauma* **9**, 11-20.
- HANSEN, T.D., WARNER, D.S., TODD, M.M., and VUST, L.J. (1989). Effects of nitrous oxide and volatile anaesthetics on cerebral blood flow. *Br. J. Anaesth.* **63**, 290-295.
- HAYES, R.L., LYETH, B.G., JENKINS, L.W., ZIMMERMAN, R., MCINTOSH, T.K., CLIFTON, G.L., and YOUNG, H.F. (1990). Possible protective effect of endogenous opioids in traumatic brain injury. *J. Neurosurg.* **72**, 252-261.

- HENDRICH, K.S., KOCHANNEK, P.M., WILLIAMS, D.S., SCHIDING, J.K., MARION, D.W., and HO, C. (1999). Early perfusion after controlled cortical impact in rats: quantification by arterial spin-labeled MRI and the influence of spin-lattice relaxation time heterogeneity. *Magn. Reson. Med.* **42**, 673-681.
- HERNANDEZ, M.J., BRENNAN, R.W., and BOWMAN, G.S. (1978). Cerebral blood flow autoregulation in the rat. *Stroke* **9**, 150-155.
- HOFFMAN, W.E., EDELMAN, G., KOCHS, E., WERNER, C., SEGIL, L., and ALBRECHT, R.F. (1991). Cerebral autoregulation in awake versus isoflurane-anesthetized rats. *Anesth. Analg.* **73**, 753-757.
- HOSSAIN, M.D., and EVERS, A.S. (1994). Volatile anesthetic-induced efflux of calcium from IP<sub>3</sub>-gated stores in clonal (GH3) pituitary cells. *Anesthesiology* **80**, 1379-1389.
- HOVDA, D.A., LEE, S.M., SMITH, M.L., et al. (1995). The neurochemical and metabolic cascade following brain injury: moving from animal models to man. *J. Neurotrauma* **12**, 903-906.
- KAMATCHI, G.L., CHAN, C.K., SNUTCH, T., DURIEUX, M.E., and LYNCH, C. (1999). Volatile anesthetic inhibition of neuronal Ca channel currents expressed in *Xenopus* oocytes. *Brain Res.* **83**, 85-96.

- KEARSE, L.A., KOSKI, G., HUSAIN, M.V., PHILBIN, D.M., and MCPECK, K. (1993). Epileptiform activity during opioid anesthesia. *Electroencephalogr. Clin. Neurophysiol.* **87**, 374-379.
- KISSIN, I., KERR, R., and SMITH, L.R. (1983). Assessment of anaesthetic action of morphine and fentanyl in rats. *Can. Anaesth. Soc. J.* **30**, 623-628.
- KOCHANIEK, P.M., MARION, D.W., ZHANG, W., et al. (1995). Severe controlled cortical impact in rats: assessment of cerebral edema, blood flow, and contusion volume. *J. Neurotrauma* **12**, 1015-1025.
- KROPFENSTEDT, S.-N., KERN, M., THOMALE, U.-W., SCHEIDER, G., LANKSCH, W.R., and UNTERBERG, A.W. (1999). Effect of cerebral perfusion pressure on contusion volume following impact injury. *J. Neurosurg.* **90**, 520-526.
- LIN, L.H., CHEN, L.L., ZIRROLI, J.A., and HARRIS, R.A. (1992). General anesthetics potentiate gamma-aminobutyric acid actions on gamma-aminobutyric acid A receptors expressed by *Xenopus* oocytes: lack of involvement of intracellular calcium. *J. Pharmacol. Exp. Ther.* **263**, 569-578.
- LYETH BG, LIU S, and HAMM RJ (1993) Combined scopolamine and morphine treatment of traumatic brain injury in the rat. *Brain Res.* **617**, 69-75.

LYETH, B.G., JIANG, J.Y., GONG, Q.-Z., HAMM, R.J., and YOUNG, H.F.

(1995). Effects of mu opioid agonist and antagonist on neurologic outcome following traumatic brain injury in the rat. *Neuropeptides* **29**, 11-19.

MACKENSEN, G.B., NELLGARD, B., MIURA, Y., CHU, C.T., DEXTER, F.,

PEARLSTEIN, R.D., and WARNER, D.S. (1999). Sympathetic ganglionic blockade masks beneficial effect of isoflurane on histologic outcome from near-complete forebrain ischemia in the rat. *Anesthesiology* **90**, 873-871.

MAGYAR, J., SZABO, G. (1996). Effects of volatile anesthetics on the G

protein-regulated muscarinic potassium channel. *Mol. Pharmacol.* **50**, 1520-1528.

MARION, D.W., DARBY, J., and YONAS, H.R. (1991). Acute regional cerebral

blood flow changes caused by severe head injuries. *J. Neurosurg.* **74**, 407-414.

MCINTOSH, T.K., FERNYAK, S., and FADEN, A.I. (1986). Endogenous

opioids, opiate receptors and traumatic brain injury. *NIDA Res. Mongr.* **75**, 527-530.

MCINTOSH, T.K., HAYES, R.L., DEWITT, D.S., AGURA, V., and FADEN,

A.I. (1987). Endogenous opioids may mediate secondary damage after experimental brain injury. *Am. J. Phys.* **253**, E565-574.

- MCINTOSH, T.K. (1993). Novel pharmacologic therapies in the treatment of experimental traumatic brain injury: a review. *J. Neurotrauma* **10**, 215-61.
- MINAMI, K., VANDERAH, T.W., MINAMI, M., and HARRIS, R.A. (1997). Inhibitory effects of anesthetics and ethanol on muscarinic receptors expressed in *Xenopus* oocytes. *Eur. J. Pharmacol.* **339**, 237-244.
- MIURA, Y., GROCOTT, H.P., BART, R.D., PEARLSTEIN, R.D., DEXTER, F., and WARNER, D.S. (1998). Differential effects of anesthetic agents on outcome from near-complete but not incomplete global ischemia in the rat. *Anesthesiology* **89**, 391-400.
- MURR, R., BERGER, S., SCHURER, L., PETER, K., and BAETHMANN, A. (1995). Influence of isoflurane, fentanyl, thiopental, and  $\alpha$ -chloralose on formation of brain edema resulting from a focal cryogenic lesion. *Anesth. Analg.* **80**, 1108-1115.
- MURR, R., SCHURER, L., BERGER, S., ENZENBACH, R., PETER, K., and BAETHMANN, A. (1993). Effects of isoflurane, fentanyl, or thiopental anesthesia on regional cerebral blood flow and brain surface PO<sub>2</sub> in the presence of a focal lesion in rabbits. *Anesth. Analg.* **77**, 898-907.
- NEUMAIER, J.F. and CHAVKIN, C. (1986). NIDA Research Monograph **75**, 101-104.

- NEUMAIER, J.F., MAILHEAU, S., and CHAVKIN, C. (1988). Opioid receptor-mediated responses in the dentate gyrus and CA1 region of the rat hippocampus. *J. Pharmacol. Exp. Ther.* **244**, 564-570.
- NIETGEN, G.W., HONEMANN, C.W., CHAN, C.K., KAMATCHI, G.L., and DURIEUX, M.E. (1998). Volatile anaesthetics have differential effects on recombinant m1 and m3 muscarinic acetylcholine receptor function. *Br. J. Anaesth.* **81**, 569-577.
- OHTA, S., NIWA, M., NOZAKI, M., HATTORI, M., SHIMONAKA, H., and DOHI, S. (1995). The mu, delta and kappa properties of various opioids. *Masui* **44**, 1228-1232.
- PATEL, P.M., DRUMMOND, J.C., COLE, D.J., and GOSKOWICZ, R.L. (1995). Isoflurane reduces ischemia-induced glutamate release in rats subjected to forebrain ischemia. *Anesthesiology* **82**, 996-1003.
- PATEL, P.M., DRUMMOND, J.C., COLE, D.J., KELLY, P.J., and WATSON, M. (1998). Isoflurane and pentobarbital reduce the frequency of transient ischemic depolarizations during focal ischemia in rats. *Anesth. Analg.* **86**, 773-783.
- RENGASAMY, A., PAJEWSKI, T.N., and JOHNS, R.A. (1997). Inhalational anesthetic effects on rat cerebellar nitric oxide and cyclic guanosine monophosphate production. *Anesthesiology* **86**, 689-698.

- ROSNER, M.J., NEWSOME, H.H., and BECKER, D.P. (1984). Mechanical brain injury: the sympathoadrenal response. *J Neurosurg* **61**, 76-86.
- SAFO, Y., YOUNG, M.L., SMITH, D.S., et al. (1985). Effects of fentanyl on local cerebral blood flow in the rat. *Acta Anaesthesiol. Scand.* **29**, 594-598.
- SOONTHON-BRANT, V., PATEL, P.M., DRUMMOND, J.C., COLE, D.J., KELLY, P.J., and WATSON, M. (1999). Fentanyl does not increase brain injury after focal cerebral ischemia in rats. *Anesth. Analg.* **88**, 49-55.
- STOVER, J., KROPPESTEDT, S., THOMALE, U., KEMPSKI, O., LANKSCH, W., and UNTERBERG, A. (1999). Isoflurane doubles plasma glutamate and increases brain edema. *Proceedings of the Eleventh International Symposium of Brain Oedema and Mechanisms of Cellular Injury Scientific Programme and Abstracts*, p 141.
- TEMPELHOFF, R., MODICA, P.A., BERNARDO, K.L., and EDWARDS, I. (1992). Fentanyl-induced electrocorticographic seizures in patients with complex partial epilepsy. *J. Neurosurg.* **77**, 201-208.
- TIPPS, L.B., COPLIN, W.M., MURRY, K.R., and RHONEY, D.H. (2000). Safety and feasibility of continuous infusion of remifentanyl in the neurosurgical intensive care unit. *Neurosurgery* **46**, 596-602.
- VINK, R., MCINTOSH, T.K., RHOMHANYI, R., and FADEN, A.I. (1990). Opiate antagonist nalmefene improves intracellular free  $Mg^{2+}$ , bioenergetic

- state, and neurologic outcome following traumatic brain injury in rats. *J. Neurosci.* **10**, 3524-3530.
- WINEGAR, B.D., OWEN, D.F., YOST, C.S., FORSAYETH, J.R., and MAYERI, E. (1996). Volatile general anesthetics produce hyperpolarization of Aplysia neurons by activation of a discrete population of baseline potassium channels. *Anesthesiology* **85**, 889-900.
- ZAHARCHUK, G., MANDEVILLE, J.B., BOGDANOV, A.A., WEISSLEDER, R., ROSEN, B.R., and MAROTA, J.J. (1999). Cerebrovascular dynamics of autoregulation and hypoperfusion. An MRI study of CBF and changes in total and microvascular cerebral blood volume during hemorrhagic hypotension. *Stroke* **30**, 2197-2205.
- ZORN, L., KULKARNI, R., ANANTHARAM, V., BAYLEY, H., and TREISTMAN, S.N. (1993). Halothane acts on many potassium channels, including a minimal potassium channel. *Neurosci. Lett.* **161**, 81-84.



**Figure 1:** MAP vs time after injury, outcome protocol. MAP in both fentanyl-treated injured (○) and sham (△) rats was approximately 50 mm Hg higher than in isoflurane-treated injured (●) and sham (▲) rats at all time points. \*  $p < 0.05$ , isoflurane vs fentanyl at each time point after injury, §  $p < 0.05$ , isoflurane vs fentanyl at all time points, including baseline, in shams.

**Figure 2:** **A.** Beam balance latency vs time in days after injury. Sham rats (△ = fentanyl, ▲ = isoflurane) had similar latencies throughout the 5-day testing period. After injury, beam balance latency was shorter for both isoflurane (●) and fentanyl (○) treatment groups vs sham; however, isoflurane-anesthetized rats recovered more quickly (vs fentanyl). \*  $p < 0.05$ , injured vs sham. **B.** Beam walking latency vs time in days after injury. Symbol designations are identical to those used in (A). Again, performance was impaired after injury in both anesthetic groups vs shams. Although isoflurane-treated rats recovered by post-injury day 3, fentanyl-treated rats failed to regain normal function by the end of the 5-day testing period. \*  $p < 0.05$ , injured vs sham.

**Figure 3:** **A.** Latency to find a platform vs time after injury in an acquisition paradigm of the MWM. Sham rats anesthetized with isoflurane (▲) or fentanyl (△) had similar performances throughout the testing period. During the first few

days of testing with a hidden platform, injured rats in both fentanyl (○) and isoflurane (●) treatment groups had impaired performance vs sham. By the third day of testing, latencies to find the hidden platform were similar in injured isoflurane-anesthetized rats and shams. In contrast, longer latencies persisted in injured fentanyl-treated rats throughout the 5-day hidden platform testing period. Latencies in all experimental groups improved during visible (vs hidden) platform testing; however, shams and injured isoflurane-anesthetized rats performed better than injured fentanyl-treated rats on both days of visible platform testing. \*  $p < 0.05$ , injured vs sham; §  $p < 0.05$ , isoflurane vs fentanyl. **B.** Swim speed, tested on day 20 after injury, did not differ between experimental groups.

**Figure 4:** **A.** Lesion volume, measured on post-injury day 21, did not differ significantly between isoflurane (■) and fentanyl (□) treatment groups. **B.** Neuronal counts in injured CA1 hippocampus were greater in isoflurane- (■) vs fentanyl-treated (□) rats. Neuronal counts in uninjured CA1 hippocampus were similar in both treatment groups. \*  $p < 0.05$ , injured vs uninjured. **C.** CA3 hippocampal neuronal counts in either injured or uninjured hemisphere did not differ significantly between isoflurane- (■) and fentanyl-treated (□) rats.

**Figure 5:** A. MAP vs time after injury, ICP protocol. MAP was approximately 40 mm Hg higher in rats treated with fentanyl (□) vs isoflurane (■) throughout the observation period. \*  $p < 0.05$ , fentanyl vs isoflurane vs fentanyl. B. ICP vs time after injury. Initial ICP was approximately 4 mm Hg in both isoflurane (■) and fentanyl (□) treatment groups. ICP progressively increased, reaching 10 - 18 mm Hg by 4 h after injury. Although ICP was similar between anesthetic groups, isoflurane-anesthetized rats exhibited a trend toward higher ICP after injury (vs fentanyl treated rats) that did not reach significance. C. CPP vs time after injury. CPP was increased in rats treated with fentanyl (□) vs isoflurane (■), \*  $p < 0.05$ , isoflurane vs fentanyl at all time points except 3.5 and 4h.

**Figure 6:** Brain Water 4 h after TBI. Brain water in the injured hemisphere was increased compared to the respective non-injured hemisphere in both isoflurane- and fentanyl-anesthetized rats; however, brain water did not differ between anesthetic groups. \*  $p < 0.05$ , isoflurane vs fentanyl.

5

CHANGES IN MITOCHONDRIAL MEMBRANE POTENTIAL IN STRETCH-INJURED ASTROCYTES AND NEURONS. S.M. Ahmed\*, B.A. Rzigalinski and E.F. Ellis, Dept. of Pharmacology and Toxicology, Medical College of Virginia, Virginia Commonwealth University, Richmond, VA.

The dynamics of energy failure in traumatically injured astrocytes and neurons are unclear. In order to better understand mitochondrial function and cell energetics following trauma we utilized the fluorescent dye Rhodamine 123, which is normally sequestered in mitochondria where its fluorescence is quenched. When mitochondrial membrane potential (MMP) decreases, such as with mitochondrial poisons, the dye moves to the cytoplasm and fluorescence is increased. Pure neuronal or astrocytic cultures were subjected to mild (5.7 mm), moderate (6.5 mm) or severe (7.5 mm) stretch-induced injury and the change in MMP measured. There were no significant changes in MMP in mildly to moderately injured neurons at 15 min, 24 or 48 hr post-injury. However severely injured neurons displayed an immediate 33% decrease in MMP that persisted to 48 hr. In contrast, mild and moderate astrocyte injury caused a dramatic, 39-52% drop in MMP at 15 min, with MMP returning to normal by 24 hr. Our results indicate that direct trauma-induced alterations in cell energetics vary greatly in neurons and astrocytes. We suggest that in vivo the deficit induced in astrocytes may alter astrocyte function, which in turn may produce dramatic effects on neuronal function. Supported by NS-27214 and NS-07288.

7

#### EVIDENCE FOR APOPTOTIC CELL DEATH FOLLOWING SUBDURAL HEMATOMA IN RATS.

B. Alessandri\*, X. Di, H. Chen, R. Bullock, Div. of Neurosurgery, Medical College of Virginia, Richmond, VA, 23298, USA

Subdural hematoma (SDH) is a common and dangerous secondary event following traumatic brain injury. The mechanisms leading to neuronal death, even after SDH removal, are not fully understood. A mechanism which might contribute to cell death is apoptotic cell death (ACD), which has been shown to be involved in the development of traumatic and ischemic brain damage.

The hematoma was produced by subdural injection of 250  $\mu$ L of autologous venous blood in Halothane anesthetized rats. Animals were allowed to survive 1 (n=3), 2 (n=3), 4 (n=3) or 7 days (n=4) after injection of SDH. Brain sections were stained by a commercially available apoptosis detection kit (FragEL™) for apoptotic cells (visualized by diaminobenzidine, DAB; counterstained by hematoxylin). Brain sections were examined light microscopically and DAB-positive cells were counted in both hemispheres in cortical, subcortical and hippocampal areas.

The DBA-pos. cell counts were  $2.3 \pm 2$ ,  $54.5 \pm 8$ ,  $13.7 \pm 8$ , and  $12.8 \pm 3$  at 1, 2, 4 and 7 days after SDH, respectively. All apoptotic cells were within the cortex, within and in the border zone of the SDH lesion. There were no DAB-pos. cells in the contralateral side. The number of DAB-pos. cells was highly correlated with the lesion area ( $r^2=0.689$ ,  $p<0.001$ ).

The results indicate that ACD occurs following SDH, and is maximally seen at 2 days. DAB-pos. cells were only found within or in the border zone of the lesion. The correlation of ACD and lesion area underlines the importance of this type of cell death in SDH. The contribution of ACD to SDH-induced brain damage and its relevance for therapy needs further study.

6

HYPERTHERMIA ADVERSELY AFFECTS OUTCOME AFTER MODERATE HEAD INJURY. Philipp R. Aldana<sup>1\*</sup>, J. Marquez<sup>1</sup>, D. S. Petrin<sup>1</sup>, D. Johns<sup>1</sup>, W. D. Dietrich<sup>2</sup>, P. A. Villanueva<sup>1</sup>, Department of Neurological Surgery<sup>1</sup>, Neurotrauma Research Center<sup>2</sup>, University of Miami School of Medicine, Miami, Florida, 33101, USA.

Hypothermia has been shown to have beneficial effects after traumatic brain injury (TBI) in both human and animal studies. Conversely, hyperthermia after TBI has been shown to have deleterious effects in animals. No studies have addressed the effects of hyperthermia after moderate head injury in humans.

104 patients admitted with a Glasgow Coma Score 9-12 due to blunt head trauma were studied. Demographics, comorbid factors and characteristics of the hyperthermic episodes ( $>38.6^\circ\text{C}$ ) were examined. The number of patients either dead, in a vegetative state or severely disabled during discharge was significantly larger for the hyperthermic group vs. the normothermic group (42.4% vs. 17.5%, respectively). A significantly larger percentage of the normothermic group had a good outcome compared to the hyperthermic group (50% vs. 20.3%, respectively). Among the hyperthermic patients, those with associated infections had significantly worse outcomes and a higher frequency of hyperthermic episodes than those without infections. We conclude that hyperthermia in the face of an associated infection may adversely affect the outcome of patients with moderate head injury. We advocate maintenance of at least normothermic conditions if moderate hypothermia cannot be achieved and treatment of any underlying infection after TBI.

8

#### VERTICAL VERSUS ANGLED CONTROLLED CORTICAL IMPACT IN RATS. H.L. Alexander\*, C.L.

Robertson, C.E. Dixon, R.S.B. Clark, S.H. Graham, P.J. Safar, P.M. Kochanek. Safar Center for Resuscitation Research, Univ. of Pittsburgh, PA 15213

Although a variety of modifications of the controlled cortical impact (CCI) model exist, a comparison between the two most common variants, vertical<sup>1</sup> and angled impact<sup>2</sup>, has not been performed. Rats were subjected to vertical (n = 8), angled (n = 8) or sham (n = 8) insults (4 m/s, 2.5 mm) to the left parietal cortex, using a CCI model with hypoxemia.<sup>3</sup> Motor (beam balance, d1-5), cognitive (Morris Water Maze, d14-21) and histologic (lesion volume, CA1 and CA3 neuron counts, d21) outcomes were studied. Motor and MWM performance were impaired, but did not differ between injury groups. Lesion volumes also did not differ (vertical =  $92.2 \pm 7.2$  mm<sup>3</sup>, angled =  $79.4 \pm 7.8$ ,  $p = 0.25$ ). CA1 neuron counts were decreased ipsilateral to injury in both groups vs sham (vertical =  $20.4 \pm 8.4$  cells/hpf, angled =  $32.7 \pm 15.8$ , sham =  $55.5 \pm 3.9$ ,  $p < .05$ ). However, CA3 neuron counts were decreased ipsilateral to injury in the vertical group vs sham ( $23.2 \pm 8.5$  vs  $52.1 \pm 6.6$ , respectively,  $p < .05$ ), but the angled group ( $32.7 \pm 15.8$ ) was not different from sham. We conclude that the vertical and angled variants of the CCI model produce similar functional deficits; however, the vertical impact appears to produce greater local damage, particularly in CA3 neurons. <sup>1</sup>J Neurotrauma 12:1015 <sup>2</sup>J Neurosci Methods 39:253; <sup>3</sup>J Neurotrauma 14:179; Support: US Army #DAMD17-97-1-7009

## ABSTRACT REPRODUCTION FORM

18th Annual National Neurotrauma Society Symposium, November 3 and 4, 2000  
New Orleans, Louisiana

Postmark Deadline: June 6, 2000

Please mark x appropriately  
below in all areas. These  
indicators will determine the  
time & placement of your  
poster for viewing:

Area: Please mark one:

- ☒ Brain Injury  
☐ Spinal Cord Injury  
☐ Neurodegenerative Diseases  
☐ Ischemia  
☐ Other \_\_\_\_\_

Model: Please mark one:

- ☒ In Vivo Models  
☐ Genetic Models  
☐ In Vitro Models  
☐ Clinical Studies  
☐ Basic Science Study  
☐ Other \_\_\_\_\_

Technique: Please mark your first, second  
and third choice with a number 1, 2 & 3.

- ☐ Behavior  
☐ Cell Biology  
☐ Imaging  
☐ Molecular Biology  
☐ Neuroanatomy/Neuropathology  
☐ Neurochemistry  
☐ Neuroimmunology  
☒ Neuropharmacology  
☐ Neurophysiology  
☐ Neuropsychology  
☐ Other \_\_\_\_\_

Subject: Please mark your first, second  
and third choice with a number 1, 2 & 3.

- ☐ Blood Flow/Metabolism  
☐ Gene Expression  
☐ Intracranial Pressure/Edema  
☒ Neurotoxicity/Neuroexcitation  
☐ Neuroprotection  
☐ Neurotransplantation  
☐ Receptor-Mediated Changes  
☐ Regeneration and Plasticity  
☐ Secondary Injury  
☐ Model Characterization  
☐ Blood Brain Barrier  
☐ Inflammation  
☐ Cell Death  
☐ Developmental / Aging

Participants may be first (presenting author) on only one abstract. \$35 Submission fee due.

Presenting Author: Henry Alexander

(Please print or type)

Address: Safar Center for Resuscitation Research

3434 Fifth Avenue, Pittsburgh, PA 15260

Telephone (office) (412) 383-1900

(outside U.S., include country code)

Facsimile (412) 624-0943

E-Mail alexanderhl@anes.upmc.edu

Type abstract here. You must stay within the border.

(See sample on back of instruction form.)

### EFFECTS OF SEVEN ANESTHETICS ON OUTCOME IN EXPERIMENTAL TRAUMATIC BRAIN INJURY

H. Alexander, K.D. Statler\*, P.M. Kochanek, C.E. Dixon,  
R.S.B. Clark, V. Vagni, S.H. Graham, D.W. Marion, P. Safar  
Safar Center, University of Pittsburgh, Pittsburgh, PA 15260

We have shown improved outcome with isoflurane vs fentanyl in the rat controlled cortical impact (CCI) model. We now present the first comprehensive study of the effect of anesthetics on outcome in experimental TBI. Rats were prepared for CCI under isoflurane and allowed to recover to tail pinch. After CCI, rats were randomized to 1 h of either isoflurane, fentanyl, morphine, pentobarbital, diazepam, ketamine, or propofol treatment or recovery without additional anesthesia (n=9/group). Brain temperature, mean arterial blood pressure (MAP), blood gases, and glucose were monitored. Motor function was assessed on d 1-5 after CCI. MAP was higher in fentanyl vs isoflurane treated rats ( $p < 0.05$ ), but did not differ between other groups. After CCI, beam balance and beam walking performances were worse for rats treated with morphine or propofol ( $p < 0.05$  vs sham). Rats administered isoflurane only before injury did as well as those given isoflurane for 1h post-trauma. Taken in context of our prior study showing improved outcome with isoflurane vs fentanyl before TBI, these data suggest benefits of isoflurane near the time of impact. Considering their common clinical use, the detrimental effects of morphine or propofol deserve further characterization. Cognitive and histological assessment in these animals is ongoing.

Support: U.S. Army#DAMD17-97-1-7009

\* ☒ Mark x if this abstract should be considered for oral presentation competition.

\* Is this abstract for Student Poster Competition? ☐ Yes ☒ No

\* Students, residents and fellows submitting an abstract must provide Dept. Head or Dean's signature to confirm student status. \_\_\_\_\_ Dept. Head / Dean signature.

\* By signing this abstract form, the author certifies that all experimental procedures have been done according to approved ethical guidelines. Henry Alexander First Author's Signature

45

**CHRONIC OVEREXPRESSION OF AMYLOID PRECURSOR PROTEIN (APP) AFTER TRAUMATIC BRAIN INJURY IN RATS.** J. R. Ciallella<sup>1</sup>\*, H. Q. Yan<sup>1</sup>, X. Ma<sup>1</sup>, D. W. Marion<sup>1</sup>, S. T. DeKosky<sup>2</sup>, and C. E. Dixon<sup>1</sup>. Departments of <sup>1</sup>Neurosurgery and <sup>2</sup>Psychiatry, University of Pittsburgh Medical Center, Pittsburgh, PA USA. Traumatic brain injury (TBI) and Alzheimer's disease (AD) produce cholinergic and metabolic deficits that may contribute to neurodegeneration. There is increasing evidence linking AD and TBI, including upregulation of APP in head injured patients (McKenzie et al. 1994 *NeuroRep* 6:161). To further investigate this linkage, we tested the hypothesis that controlled cortical impact (CCI) injury would produce chronic upregulation of APP protein levels at 4 weeks following injury. Our previous studies demonstrated significant changes in cholinergic proteins at this time point (Ciallella et al. 1998. *Exp. Neurol.* In Press). APP immunohistochemistry (n=3-5) and western blot (n=4) were performed on cortical and hippocampal regions from injured and sham animals. The same N-terminal antibody was used in all studies. A marked increase in cortical and hippocampal APP protein was demonstrated bilaterally by both immunohistochemistry and western blot in injured rats compared to sham controls. This demonstrates that a single TBI can lead to chronic upregulation of APP, concurrent with chronic alterations in cholinergic markers. Supported by AG05133, NINDS-T32NS07391, CDC-CCR312296, NIH-NS30313, and NIH-NS33150.

47

**THE SUPPRESSION OF HIPPOCAMPAL NGF mRNA AFTER CEREBRAL ISCHEMIA IN RAT TREATED WITH ANTISENSE DNA TO C-fos.** J.-K. Cui<sup>1</sup>\*, C. Y. Hsu<sup>2</sup>, and P. K. Liu<sup>1</sup>. <sup>1</sup>Department of Neurosurgery, Baylor College of Medicine, Houston, TX 77030; <sup>2</sup>Department of Neurology, Washington University, St Louis, MO 63110.

The biological effects of Fos expression in the brain were examined using phosphorothioated oligodeoxynucleotides (s-ODNs) to c-fos, mcfosr<sub>115</sub>. Biotinylated antisense mcfosr<sub>115</sub> (bio-mcfosr<sub>115</sub>) plus lipofectin were delivered into the brain of male Long-Evans rats (225-250 gm) via intracerebroventricular infusion. The distribution of bio-mcfosr<sub>115</sub> was detected using antibodies against biotin. Using dot blot analysis on the recovered bio-mcfosr<sub>115</sub>, the bio-mcfosr<sub>115</sub> uptake in hippocampus peaked at 29-48 hrs, and the internalized bio-mcfosr<sub>115</sub> was degraded within 72 hr of infusion. The s-ODN uptake in the brain was confirmed by 3'-end-labeling with digoxigenin-dUTP, using terminal transferase and anti-digoxigenin IgG-FITC. The presence of fluorescent aggregates in the brain cells near the vessel wall in animals treated with antisense mcfosr<sub>115</sub> + lipofectin suggests lipofectin mediated s-ODN transfer across the blood brain barrier. The uptake increased with time and with the dose delivered. The effectiveness of antisense mcfosr<sub>115</sub> was shown by an inhibition of ischemia-induced Fos expression, and was accomplished by an inhibition of ischemia-induced hippocampal NGF mRNA expression in the brain of animals pretreated with antisense mcfosr<sub>115</sub>. The specificity of Fos suppression was suggested by a lack of antisense mcfosr<sub>115</sub> effect on the expression of NT-3 and  $\alpha$ -actin mRNA.

46

**EFFECT OF HYPOTHERMIA AFTER SEVERE TRAUMATIC BRAIN INJURY WITH SECONDARY HYPOXEMIA IN RATS.** R.S.B. Clark\*, C.L. Robertson, C.E. Dixon, H.L. Alexander, S.H. Graham, P.J. Safar, P.M. Kochanek. Safar Center for Resus Res., U of Pgh, PA 15213.

Many reports have shown benefit from hypothermia (HT) in traumatic brain injury (TBI); but, its effect on TBI with secondary insult remains undefined. We hypothesized that HT would improve outcome after controlled-cortical impact (CCI) with secondary hypoxemia. Rats received severe CCI injury followed by 30 min of hypoxemia,<sup>1</sup> and randomized to normothermia (NT=37°C brain temp, n=19), immediate HT (IHT=32°C, after CCI, n=10), or delayed HT (DHT=32°C, after hypoxemia, n=14) for 4 h. Motor (beam balance/walking, d1-5), cognitive (Morris Water Maze [MWM], d14-21) and histologic outcomes (lesion volume, hippocampal neuron counts, d21) were evaluated. Motor and MWM performance were impaired but did not differ between groups. Lesion volumes (mm<sup>3</sup>) did not differ between groups (NT=65.3±6.9, IHT=50.2±8.2, DHT=53.7±7.9). Neuron counts (CA1, CA3) were decreased 60-70% ipsilateral to CCI, but did not differ between groups. Mortality doubled (43% vs 20-21%) in DHT vs NT or IHT ( $p = 0.3$ ). HT did not improve outcome after severe CCI with secondary insult. Clinical studies<sup>2</sup> exclude patients with secondary insults, and suggest HT is not effective after severe injury (GCS 3-4). Novel therapies may be needed in this setting. <sup>1</sup>J Neurotrauma 14:179; <sup>2</sup>NEJM 336:540-6; Support: US Army #DAMD17-97-1-7009

48

**LOSS OF GLIAL POTASSIUM CURRENTS AND IMPAIRMENT OF POTASSIUM HOMEOSTASIS FOLLOWING FLUID PERCUSSION INJURY.** R. D'Ambrosio\*, D.O. Maris, M.S. Grady, and D. Janigro. Dept. of Neurosurgery, Univ. of Washington, Seattle, WA 98104

We compared the early effects of moderate in vivo fluid percussion injury (FPI) on the functional expression of potassium currents expressed in oligodendroglia and astrocytes from acutely isolated rat hippocampal slices. Whole cell recordings were performed from post-FPI and naïve slices of 30 d.o. rats. Cs<sup>+</sup> (1 mM) was used to block inward potassium currents. K<sup>+</sup>-selective electrodes were employed to measure K<sup>+</sup> accumulation in radiatum CA3. GFAP immunostaining was enhanced in CA3 24-48 hrs following FPI, while immunostaining for oligodendroglia was reduced. A significant decrease in Cs<sup>+</sup>-sensitive potassium currents was observed following lesion in both oligodendroglia and astrocytes. Cells characterized by complex electrophysiological profiles as well as those characterized by inward rectification were equally affected (-60% and -55% at -140 mV). Morphologically, complex cells visualized by biocytin staining could be classified as oligodendrocytes. Stimulation (1 Hz) of Schaffer collaterals induced K<sup>+</sup> accumulation in radiatum CA3. Slices obtained from naïve rats always showed a recovery of extracellular K<sup>+</sup> to basal levels within 10 seconds following stimulation (n=5). Slices obtained from post-FPI rats displayed recovery times ranging from 10 to 40 seconds (n=8). Additionally, 75% of the post-FPI slices generated multiple afterdischarges during stimulation, while only 20% of the control slices did. These results indicate that 1) post-FPI CA3 astrocytes are reactive or injured, 2) loss of Cs<sup>+</sup>-sensitive potassium current occurs in oligodendrocytes and astrocytes post-FPI, 3) neuronal-activity-induced elevation of [K<sup>+</sup>]<sub>out</sub> is more persistent at early time point post-FPI, 4) hyperexcitability is observed after trauma without detectable neuronal loss. We conclude that FPI may affect extracellular K<sup>+</sup>-homeostasis by impairing glial potassium currents. Supported by NIH-51614 and RO-1 NS33107.



# EFFECT OF CALCIUM CHLORIDE ON REGIONAL CEREBRAL BLOOD FLOW DURING CARDIOPULMONARY RESUSCITATION IN PIGLETS

Melody Palmer Land, John Kuluz, Barry Gelman, Michael Nares, En Xu, and Charles Schleien. Pediatric Critical Care Medicine, University of Miami School of Medicine, Miami, FL 33101.

**Introduction:** The use of calcium chloride ( $\text{CaCl}_2$ ) during CPR remains controversial.  $\text{CaCl}_2$  may improve the effectiveness of CPR by increasing systemic vascular tone and vital organ perfusion. Alternatively,  $\text{CaCl}_2$  may cause regional vasoconstriction in the brain and heart, resulting in secondary ischemic injury. We hypothesized that administration of the standard dose of  $\text{CaCl}_2$  during CPR decreases rCBF.

**Methods:** Under pentobarbital anesthesia, 2-4 week old piglets underwent 6 min of cardiac arrest by ventricular fibrillation, and 30 min of standard CPR. rCBF was measured with microspheres at baseline and after 5, 15 and 30 min of CPR.  $\text{CaCl}_2$  20 mg/kg (n=5) or saline (n=5) was given after 1 and 19 min of CPR. Data (mean $\pm$ SE) were analyzed by ANOVA and Student's t-test (\* p<0.05).

**Results:** Ionized (io) Ca decreased from  $1.40\pm 0.03$  at baseline to  $1.16\pm 0.05^*$  at 15 min and  $1.18\pm 0.05^*$  at 30 min CPR. After  $\text{CaCl}_2$ , ioCa increased to  $2.58\pm 0.16^*$  at 5 min and  $2.04\pm 0.21^*$  at 30 min, and was not different from baseline at 15 min CPR. Calcium increased aortic pressure ( $44\pm 2$  vs  $38\pm 2^*$ ) and cerebral perfusion pressure at 5 min CPR. Total CBF was not different between groups at any time point; however, severe regional ischemia (CBF<15 ml/100g/min) was more common after 30 min CPR when  $\text{CaCl}_2$  was given, particularly in subcortical regions (p<0.03).

**Conclusion:** These data show that  $\text{CaCl}_2$  administration has adverse effects on rCBF during prolonged CPR and may worsen ischemic brain injury. Future studies will determine the effect of  $\text{CaCl}_2$  on functional and neuropathologic outcome.

# NO LONG-TERM BENEFIT FROM HYPOTHERMIA AFTER SEVERE TRAUMATIC BRAIN INJURY WITH SECONDARY HYPOXEMIA IN RATS

Courtney L. Robertson, Robert Clark, C. Edward Dixon, Steven Graham, Henry Alexander, Stephen Wisniewski, Donald Marion, Peter Safar, Patrick Kochanek. Depts of Anesthesiology/CCM, Pediatrics, Neurology, Epidemiology, and Neurosurgery. Safar Center for Resuscitation Research, University of Pittsburgh. PA 15213.

**Introduction:** Many reports have shown benefit from hypothermia in traumatic brain injury (TBI); but its effect in the setting of TBI with secondary insult remains undefined. Clinical studies show an increase in morbidity and mortality after severe TBI with secondary brain insult.<sup>1</sup> In experimental rat models, outcomes were worse in brain injury with secondary hypoxia.<sup>2</sup> Recently, we characterized a model of TBI with secondary hypoxemia and reported prominent neuronal apoptosis after injury.<sup>3,4</sup> We hypothesized that hypothermia would improve outcome after controlled-cortical impact (CCI) with secondary hypoxemic insult in rats.

**Methods:** Rats were subjected to severe CCI injury followed by 30 min of hypoxemia ( $\text{PaO}_2=35-45$  mm Hg).<sup>5</sup> Rats were then randomized to normothermia (NT=37°C, n=19), immediate hypothermia (IHT=32°C, after CCI, n=10), or delayed hypothermia (DHT=32°C, after hypoxemia, n=14) for 4 h. Motor (beam balance/beam walking, d 1-5), cognitive (Morris Water Maze [MWM], d 14-21) and histologic outcome (lesion volume, hippocampal neuron counts, d21) were evaluated.

**Results:** Motor and MWM performance were impaired, but did not differ between groups. Lesion volumes did not differ significantly between groups (NT=65.3 mm<sup>3</sup>  $\pm$  6.9, IHT=50.2 $\pm$ 8.2, DHT=53.7 $\pm$ 7.9). Hippocampal neuron counts (CA1, CA3) were decreased on the injured side, but did not differ between groups (NT-CA1=19.8 $\pm$ 4.2 cells/hpf, NT-CA3=19.8 $\pm$ 4.6, IHT-CA1=13.2 $\pm$ 8.7, IHT-CA3=15.6 $\pm$ 7.3, DHT-CA1=13.7 $\pm$ 5.8, DHT-CA3=18.5 $\pm$ 7.3). Mortality rate did not differ significantly between groups.

**Conclusions:** Immediate or delayed hypothermia did not improve long-term outcome after severe CCI with secondary hypoxemia in rats. The severity of the combined insult may be outside of the therapeutic window of opportunity. Clinical studies<sup>5</sup> have excluded patients with secondary insult, and have indicated that hypothermia is of limited efficacy in the subset of severely injured (GCS 3-4) patients. Novel therapeutic approaches or combination therapies may be necessary in this setting. <sup>1</sup>J Trauma 34:216, <sup>2</sup>J Cereb Blood Flow Metab 7:759, <sup>3</sup>J Neurotrauma 14:179, <sup>4</sup>J Neurosci 17:9172, <sup>5</sup>NEJM 336:540; Support: US Army DAMD17-97-1-7009

# SYSTEMIC AND GLOBAL CEREBRAL OXYGEN EXTRACTION ( $\text{O}_2\text{Ext}$ ) IN ACUTE CARBON MONOXIDE (CO) TOXICITY

Deborah M. Lopez, Jacqueline Weingarten-Arams, Lewis Singer, Edward Conway Jr. Department of Pediatrics, Division of Critical Care Medicine, Albert Einstein College of Medicine, Montefiore Medical Center, Bronx, New York 10467

**Introduction:** Carbon Monoxide is a colorless and odorless gas produced by incomplete combustion of carbon containing compounds. CO affects most notably those organs with high metabolic rates such as the central nervous system. CO causes a left shift of the oxyhemoglobin dissociation curve. Limited adult animal studies have suggested that systemic  $\text{O}_2\text{Ext}$  is decreased in the presence of CO toxicity. The purpose of this study is to evaluate systemic and cerebral  $\text{O}_2\text{Ext}$  in a pediatric model of CO toxicity.

**Methods:** 15 Yorkshire piglets were anesthetized with pentobarbital. Tracheostomy, femoral arterial, pulmonary arterial and retrograde jugular venous bulb pressure catheters were inserted. Following a one hour rest period, baseline data was collected. CO was administered via the endotracheal tube to achieve and maintain a level of 60% carboxyhemoglobin (COHb). Arterial, mixed venous and internal jugular blood samples were drawn within five minutes of each other. Blood samples were measured with the Radiometer OSM 3 Hemoximeter.  $\text{O}_2\text{Ext}$  was calculated via standard formula. Measurements were stratified by the corresponding COHb level: mild toxicity = 0-10%, moderate =  $\geq 10-40\%$  and severe  $\geq 40\%$ .

**Results:** 158 sets of blood samples were obtained. Mean values are summarized below. Systemic versus cerebral  $\text{O}_2\text{Ext}$  were analyzed via two tailed t-test and were significant at all levels of COHb with \*p< 0.01.

COHb%	$\text{O}_2$ delivery (ml $\text{O}_2$ /min)	CO (L/min)	Systemic $\text{O}_2\text{Ext}$	Cerebral $\text{O}_2\text{Ext}$
0-10	102.6	1.14	0.40	0.28 *
>10-40	95	1.17	0.43	0.36 *, #
>40	56.8	1.40	0.41	0.33 *, #

In addition, cerebral oxygen extraction significantly increased as the percentage of COHb escalated with no change in systemic oxygen extraction. (ANOVA, # p = 0.05)

**Conclusion:** Cerebral oxygen extraction increased with increasing COHb level. Contrary to adult animal studies, systemic oxygen extraction remained unchanged despite increasing COHb levels. Cardiac output remained the same and as expected oxygen delivery decreased with increasing level of COHb.

# BRAIN NITRIC OXIDE CHANGES AFTER CONTROLLED CORTICAL IMPACT INJURY IN RATS

Leela Cherian, J. Clay. Goodman, Claudia S. Robertson. Departments of Neurosurgery and Pathology, Baylor College of Medicine, Houston, TX-77030

**Introduction:** The marked reduction in CBF that occurs after severe controlled cortical impact (CCI) injury in rats can be ameliorated by postinjury infusion of L-arginine. Since L-arginine is a substrate for nitric oxide synthase, these studies suggest that a reduced production of nitric oxide (NO) may play a role in the CBF reduction that occurs after brain trauma. The purpose of this study was to measure brain tissue concentrations of NO after severe CCI.

**Methods:** 12 Long Evans rats were anesthetized with isoflurane and subjected to severe CCI (5 m/sec, 3 mm deformation) or sham CCI. NO was directly measured using a NO electrode which was inserted at the site of the impact after calibration using S-nitroso N-acetyl-D-L-penicillamine at 37°C. A microdialysis probe was inserted near the NO electrode and perfused with artificial CSF at 2  $\mu$ l/min. The concentration of nitrate/nitrite was measured using a chemiluminescent method in serial 20 minute collections of dialysate. These measurements were obtained prior to injury, and for 3 hours after injury. Values were expressed as % of the pre-injury baseline values.

**Results:** Impact injury caused a transient increase in brain tissue NO concentrations to 178 % of the baseline values in the CCI animals, compared to 98 % in the sham injured animals (p=0.002). After the initial transient increase in NO, the concentrations of NO declined and remained significantly lower than in the sham animals throughout the 3 hr study period. The results are summarized below as median (25<sup>th</sup> percentile, 75<sup>th</sup> percentile). A similar reduction in nitrate/nitrite was observed in the microdialysates.

Time after Injury	NO (%baseline) CCI injury (n=6)	NO (% baseline) Sham injury (n=6)	P value
2min	177.5 (158,197)	97.5 (96,102)	0.002
1hr	75.5(70,80)	98.5(96,110)	0.004
2hr	73 (69,79)	101 (95,114)	0.001
3hr	70 (67,78)	95.5 (92,100)	0.002

**Conclusions:** This study suggests that NO is released immediately after a severe brain injury and subsequently is found in decreased concentrations in the brain for at least 3 hours after injury. The reduction in CBF that occurs after severe CCI may be related to the reduced NO levels.

Supported by NIH grant #P01-NS27616

**E48**

**MK-801 improves functional outcome in rats after controlled cortical impact.** RA Ruppel, PM Kochanek, CE Dixon, HL Alexander, SH Graham, RSB Clark, SR Wisniewski, DW Marion, PJ Safar. Safar Center for Resuscitation Research, Univ of Pitt, Pgh, PA

Excitotoxicity is implicated as a key mechanism of secondary neuronal damage after traumatic brain injury (TBI). The NMDA receptor antagonist MK-801 has been shown to attenuate cerebral injury in focal ischemia and some models of TBI, but it has not been tested in controlled cortical impact (CCI). We hypothesized that MK-801 would improve functional and histopathologic outcomes in rats following CCI. Anesthetized Sprague-Dawley rats ( $n=8/\text{grp}$ ) were subjected to CCI (4 m/s, 2.5 mm depth), then randomized to immediate treatment with either MK-801 (1 mg/kg IP) or vehicle. Rats treated with MK-801 recovered motor function significantly earlier than vehicle controls, as shown by beam balance/walking performance (d 1-5). MK-801 treated rats also showed improvement in the probe trial of the Morris water maze (d 14-20) vs vehicle ( $p<0.05$ ), but no differences were seen in latencies to target. Contusion volume and hippocampal cell counts (d 21) did not differ between the groups. These data demonstrate an important role for excitotoxicity early after cerebral contusion and support continued evaluation of anti-excitotoxic therapies for use in TBI.

**Funding:** DAMD17-97-1-7009

**E50**

**TREATMENT OF SPINAL CORD INJURY WITH GK-11.** Gaetano Menna, Wencheng Huang and Wise Young. Neuroscience Center, Rutgers, The State University of New Jersey, 604 Allison Rd, Piscataway, New Jersey 08854.

GK-11 is a long-lasting blocker of NMDA receptors. Several studies have reported that NMDA receptor blockers reduce tissue damage. We assessed the effects of GK-11 on a well-standardized rat spinal cord contusion model, comparing 2.7 mg/kg, 0.9 mg/kg, and 0.3 mg/kg doses and vehicle started at 15 minutes or 60 minutes after 12.5 mm or 25.0 mm contusions with the NYU weight drop impactor. A total of 134 adult Long-Evan's hooded rats were studied in this study. The rats were anesthetized with intraperitoneal pentobarbital (male 60 mg/kg and female 45 mg/kg) and injured at T9-10 cord exposed with laminectomy. At 24 hours after injury, the spinal cords were rapidly removed and frozen. Six cord samples were removed from each rat and each sample was approximately 5 mm in length. To quantify tissue damage, we measured spinal cord lesion volumes, cell volume fractions (CVF), tissue Na, K concentrations and edema at and around the impact site. The results show that GK11 did not have any beneficial effects in tissue Na, K, water concentrations, cell volume fractions and lesion volumes compared to vehicle groups ( $p>0.05$ ). The analyses also rule out a possible effect of GK-11 only on the surrounding cord because repeated measures analyses did not reveal any consistent treatment-related Na, K, CVF or lesion volume differences at specific sample sites. We therefore conclude that GK-11 in the dose ranges of 0.3-2.7 mg/kg given at 15 and 60 minutes after injury does not alter the cell loss in spinal cords contusion injury in the presence of pentobarbital anesthesia.

**E49**

**PROTECTION WITH MK-801 AGAINST SUSCEPTIBILITY OF MICE EXPRESSING HUMAN APOLIPOPROTEIN E4 TO CA1 NEURONAL INJURY FROM TRAUMA AND OXIDATIVE STRESS.** R.A. Wallis<sup>1,2</sup>, K.L. Panizzon<sup>1,3</sup>, B. Teter<sup>2,3</sup>, G.M. Cole<sup>1,2,3</sup>, S.A. Frautsch<sup>1,2,3</sup>, J.R. Gilbert<sup>4</sup>. <sup>1</sup>Dept. of Neuro., UCLA; <sup>2</sup>Dept. of Geri., UCLA, LA, CA 90024; <sup>3</sup>Sepulveda VAMC, Sepulveda, CA 91343; <sup>4</sup>Dept. of Neuro., Duke University, NC 27710.

Susceptibility of CA1 neurons to trauma and oxidative stress was assessed in transgenic mice expressing the human apolipoprotein E4 gene and promoter in the ApoE knockout background. Mild trauma to hippocampal slices from ApoE knockout mice produced virtually full recovery of mean orthodromic (ortho.) and antidromic (anti.) population spike (PS) amplitude recovery of  $94\% \pm 2$  and  $95\% \pm 1$  one hr after trauma, while slices from ApoE4 mice showed ortho. and anti. PS recoveries of CA1 of  $16\% \pm 5$  and  $14\% \pm 3$  ( $p<0.05$ ). MK-801 32  $\mu\text{M}$  treatment reversed ApoE4 susceptibility with CA1 ortho. and anti. PS recoveries of  $66\% \pm 5$  and  $64\% \pm 3$  ( $p<0.05$ ). Oxidative stress ( $\text{H}_2\text{O}_2$  5  $\mu\text{M}$ , 6 min), to ApoE knockout mouse slices showed ortho. and anti. PS recovery after 1 hr of  $89\% \pm 2$  and  $91\% \pm 2$ , while slices from ApoE4 mice showed ortho. and anti. PS recoveries of  $23\% \pm 2$  and  $19\% \pm 2$  ( $p<0.05$ ). MK-801 reversed the susceptibility of ApoE4 mice to oxidative stress with ortho. and anti. PS recoveries of  $83\% \pm 2$  and  $79\% \pm 2$  ( $p<0.05$ ). These findings suggest that the ApoE4 gene increases susceptibility of CA1 neurons to trauma and oxidative stress through excitotoxic mechanisms. Supported by the VA Research Service.

**E51**

**GROUP I METABOTROPIC GLUTAMATE ANTAGONIST REDUCES NEURONAL DEGENERATION AND BEHAVIORAL DEFICITS AFTER RAT FLUID PERCUSSION INJURY.** Q-Z. Gong\*, S.D. Shields, S. Murphy, R.F. Berman, J.P. Muizelaar, B.G. Lyeth. Dept. Neurological Surgery, U.C. Davis, Davis, CA 95616, USA.

Acute activation of Group I metabotropic glutamate receptors (mGluRs) contributes to traumatic brain injury (TBI) pathophysiology (J Neurosci 16:6012, 1996). We infused over 1 hr, the selective mGluR Group I antagonist, (RS)-1-aminoinidan-1,5- dicarboxylic acid (AIDA) (0, .4, 2, 10 nmol) ( $n=6/\text{group}$ ) into the hippocampus beginning at 5 min after parasagittal fluid percussion TBI. At 24 hrs after TBI, coronal sections were stained with Fluoro-Jade, a fluorescent marker for neuronal degeneration. Positive staining cells were counted in 4 sections/rat. Significantly fewer ( $p<0.05$ ) CA2-3 Fluoro-Jade positive neurons were detected in the 10 nmol AIDA group ( $184 \pm 32$ ) compared to the vehicle group ( $310 \pm 47$ ). In a second experiment, rats were administered 10 nmol AIDA or vehicle ( $n=10/\text{group}$ ) after TBI as above and tested in the Morris water maze for acquisition of a spatial learning/memory task on days 11-15 post-injury. The mean swim distance for the 10 nmol AIDA-treated group ( $1973 \pm 146\text{cm}$ ) was significantly ( $p<0.04$ ) shorter than vehicle controls ( $1493 \pm 142\text{cm}$ ). Post-injury blockade of Group I mGluRs appears to reduce neuronal degeneration and improve functional outcome after TBI. Supported by NIH NS29995



## 216.1

**DELAYED 45-CALCIUM ACCUMULATION FOLLOWING TRAUMATIC BRAIN INJURY IS AGE-DEPENDENT AND REFLECTS SECONDARY CELL DEATH IN THE THALAMUS.** C.L. Osien\*, A.H. Moore, M.L. Prins and D.A. Hovda. Depart. of Surg., Med. & Molec. Pharm., Phys. Sci., Div. of Neurosurg.; UCLA School of Medicine, Los Angeles, CA 90095-7039

Calcium flux is considered an important factor in the pathophysiology of traumatic brain injury. Our previous work has shown diffuse  $^{45}\text{Ca}^{++}$  accumulation in the cortex immediately after lateral fluid percussion (LFP) injury lasting at least 2 days in P17, P28, and adult rats. In addition to this first, acute period of isotope accumulation these studies suggested a second, delayed period of  $^{45}\text{Ca}^{++}$  accumulation in the thalamus. To determine the developmental profile of this delayed  $^{45}\text{Ca}^{++}$  accumulation 36 P17, 34 P28, and 17 adult rats were anesthetized and subjected to a moderate-severe ( $-2.75$  atm) LFP. Immediately, 6 hours, 1 day, 2 days, 4 days, 7 days, and 14 days after injury the rats were injected with  $^{45}\text{Ca}^{++}$  ( $1\mu\text{g}$ , i.v.) and processed for autoradiography. Optical densities were measured in 16 regions of interest, including 6 thalamic nuclei.  $^{45}\text{Ca}^{++}$  accumulation was evident within the ipsilateral cerebral cortex immediately after injury ( $\sim 40\%$  increase), returning to sham levels within 4 days in all three age groups ( $p < 0.03$ ). In contrast, differences existed between the age groups for delayed  $^{45}\text{Ca}^{++}$  accumulation in the thalamus. While P17s showed no delayed  $^{45}\text{Ca}^{++}$  accumulation, P28 and adult rats showed significant accumulation in the thalamus starting at 2 days and increasing out to 14 days ( $\sim 50\%$  increase,  $p < 0.03$ ). Since histological analysis indicated cell death at the time of the delayed  $^{45}\text{Ca}^{++}$  accumulation, this delayed accumulation could represent retrograde degeneration due to diffuse axonal injury and/or  $\text{Ca}^{++}$ -induced apoptosis. The age-dependency of delayed  $^{45}\text{Ca}^{++}$  accumulation in the thalamus may be attributed to differential biomechanical consequences of the LFP and/or greater calcium buffering capacities of younger animals. The results suggest that two temporal patterns of  $^{45}\text{Ca}^{++}$  accumulation exist following LFP: acute calcium flux associated with the injury-induced ionic cascade and delayed calcium accumulation associated with secondary cell death. Supported by NS30308, NS27544.

## 216.3

**IS GUANOSINE BEING MISTAKEN FOR THE PEROXYNITRITE MARKER 3-NITROTYROSINE IN MODELS OF CNS INJURY?** J.S. Althaus, R.L. Roof, A.P. Acharya, S.T. Fountain, W.F. Pool, M.D. Reilly, R.T. Carroll\*, M.D. Davis and E.D. Hall. Parke-Davis Pharm. Research, Ann Arbor, Michigan 48105.

A marker used to identify peroxynitrite activity following CNS injury is the 3-nitrotyrosyl residue of proteins. Recently, a number of studies have purported measurement of 3-nitrotyrosine (3-NT) in brain protein digest by HPLC. These assays vary substantially in processing, chromatographic and detection methodologies. Halliwell and collaborators (J. Neurochem. 70:2220-2223, 1998) reported measurement of an artifactual substance in brain tissue which exhibited chromatographic, electrochemical and chemical properties nearly identical to 3-NT. It was suggested that this artifact might confound the detection of 3-NT in brain tissue. We have developed an HPLC assay for the measurement of 3-NT that circumvents the problem of artifact detection. This was accomplished by using gradient elution, ion pairing and multi-channel electrochemical detection. Using this technology, we were able to measure, in injured brain protein digests, 3-NT as a percentage of tyrosine (3-NT/TYR) at levels much lower (0.004%) than purported (J. Cereb. Blood Flow Metab. 18:123-129, 1998). In fact, at 24 hrs, after impact-acceleration head injury in rat, hippocampal 3-NT/TYR was small did not differ from sham animals. However, in the same model, another peak that eluted very close to 3-NT increased significantly after injury. This same peak was found to increase in microdialysate with rat head injury as well, with no apparent change in 3-NT. The former response was blocked by L-NAME, the non-selective inhibitor of nitric oxide synthase. We suggest that this may be the same artifact reported by Halliwell and collaborators. Isolation of this peak material followed by LC-MS, LC-NMR, LC-EC and LC-UV confirms the identity as guanosine. We recommend including guanosine as an HPLC standard to avoid misidentification with 3-NT. (Supported by Warner-Lambert/Parke-Davis)

## 216.5

**ADENOVIRUS-MEDIATED TRANSFER AND EXPRESSION OF  $\beta$ -GAL IN INJURED HIPPOCAMPUS IS NOT INHIBITED AFTER TRAUMATIC BRAIN INJURY IN MICE.** P.M. Kochanek\*, K.L. Janesko, L.W. Jenkins, P. Robichaud, H.Q. Yan, R.S.B. Clark, C.E. Dixon, D.W. Marion, M.R. Kibbe, T.R. Billiar. Safar Center for Resuscitation Research and Depts. of Anesthesiology/CCM, Pediatrics, Neurosurgery, Surgery, Univ. of Pittsburgh, Pittsburgh, PA, 15260.

In models of focal cerebral ischemia, adenoviral (Ad) gene transfer is attenuated or markedly delayed vs native. After controlled cortical impact (CCI)-induced traumatic brain injury (TBI) in mice, CA1 and CA3 hippocampus both exhibit delayed neuronal death, with DNA damage at 24h, morphological loss of CA3 by 72h, and complete loss of hippocampal parenchyma by 21d. We hypothesized that Ad-mediated expression of  $\beta$ -Gal in hippocampus would be attenuated after CCI in mice.

Isoflurane anesthetized C57BL/6 mice ( $n=16$ ) were subjected to either CCI to left parietal cortex or sham injury (burr hole). Ad-CMV- $\beta$ -Gal ( $10^9$  PFU/ml) was then immediately injected into left dorsal hippocampus. At 24 or 72h, mice were saline perfused and  $\beta$ -Gal expression was quantified (mU/mg protein). Separate mice ( $n=8$ ) were used to examine the distribution of  $\beta$ -Gal staining in vibratome sections.

Robust  $\beta$ -Gal expression in left hippocampus was detected in sham and was similar at 24h ( $48.4 \pm 4.1$ ) vs 72h ( $68.8 \pm 8.8$ , NS). CCI did not reduce  $\beta$ -Gal expression in ipsilateral hippocampus ( $68.8 \pm 8.8$  vs  $88.1 \pm 7.0$  at 72h, sham vs CCI, NS); but, CCI reduced  $\beta$ -Gal expression in contralateral hippocampus ( $14.2 \pm 3.9$  vs  $2.5 \pm 0.2$  at 72h  $p < 0.05$  sham vs CCI).  $\beta$ -Gal was seen in many cell types in ipsilateral hippocampus. Contralateral expression was restricted to periventricular cells and CA3 neurons.

Despite the eventual nearly complete loss of ipsilateral hippocampus by 21 d in this model, Ad-mediated gene transfer is robust in this structure early after TBI. This supports the use of this approach to test novel genes targeting hippocampal neuronal death in this model. Inhibition of gene expression in contralateral hippocampus by injury may reflect reduced CSF circulation or failure of axonal transfer of Ad after CCI. Abe et al, 1997. Whalen et al, 1999. Support: NS30318 and GM44100

## 216.2

**NMDA antagonists enhance delayed neurodegeneration in the hippocampus following traumatic brain injury.** C. Ikonomidou<sup>1</sup> and L. Turski<sup>2</sup>. <sup>1</sup>Dept. Ped. Neurology, Humboldt Univ., Berlin, Germany; <sup>2</sup>Eisai London Res. Laboratories, UK.

Traumatic cortical injury in adult rats causes two types of lesions, an acute local lesion within the cortex and a distant lesion in the ipsilateral CA3 hippocampus which evolves in a delayed fashion. The effect of pre- and posttreatment with the NMDA antagonist CPP on delayed neurodegeneration in the hippocampus was analysed by means of stereological morphometry. Male Fisher 344 rats were anesthetized with tribromoethanol, placed in a stereotaxic apparatus and a craniotomy was performed over the right sensorimotor cortex. A force of 380gxcm produced by a 20g falling weight was selected to produce brain contusion. Rats were sacrificed by perfusion fixation at 3 days after trauma. Pretreatment with CPP (30 mg/kg i.p. at 2, 1 and 0 hrs prior to trauma) resulted in amelioration of hippocampal neuronal loss within the ipsilateral CA3 subfield. When treatment with the same dose-regimen was initiated at 1 hr after trauma no effect on numbers of hippocampal CA3 pyramids was evident, whereas treatment with CPP that was initiated at 4 or 7 hrs after trauma resulted in significantly greater neuronal cell loss compared to vehicle treated rats. CPP demonstrated no effect on neuronal densities and total cell numbers within the ipsilateral CA3 subfield when treatment was initiated at 10 hrs after trauma. These observations reveal that during a critical period following traumatic brain injury, NMDA antagonists enhance slow neurodegeneration and can therefore have neurodestructive properties. Supported by BMBF grant 01K095151TPA3.

## 216.4

**CHANGES IN CORTICAL ENERGY METABOLISM AFTER CLOSED HEAD INJURY IN THE MOUSE.** A.E.M. Mautes\*, B. Cornely, W.-I. Steudel, Y. Yang<sup>1</sup>, E. Shohami<sup>1</sup>. Neurosurg. Res. Lab. Saarland Univ., Homburg, FRG, Dept. of Pharmacol. Hebrew University, School of Pharmacy, Jerusalem, Israel<sup>1</sup>.

The induction of cytokines had been demonstrated after acute stroke and trauma. Temporal pattern of changes in energy balance has been closely linked to cytokine expression. To investigate a possible correlation between cytokine production and energy metabolism after closed head injury (CHI), the present study was designed in a mouse model of CHI. Injury was performed using a weight drop device (Chen et al., 1996) and brains were frozen 4-24h after sham surgery and at 5 min, 4, 12 and 24h ( $n=4$ /group) following CHI. ATP, glucose and lactate contents were determined by computer-assisted bioluminescence imaging in serial tissue sections. Results: i) ATP content was significantly decreased at 4, 12 and 24h on the injured hemisphere as compared to sham. On the contralateral side ATP did not change in comparison to sham. ii) glucose content ipsilaterally significantly decreased at 5min, and remained lower up to 24h. Contralateral glucose content did not change significantly as compared to sham. iii) no changes in lactate could be discerned. Conclusions: CHI in the mouse lead to ipsilateral cortical energy depression. This may be related to the early production of harmful mediators such as cytokines, reactive oxygen species or vasoactive neurotransmitters that locally impair the blood supply.

## 216.6

**FENTANYL VERSUS ISOFLURANE ANESTHESIA: EFFECT ON OUTCOME AFTER TRAUMATIC BRAIN INJURY IN RATS.** K.D. Statler, P.M. Kochanek, C.E. Dixon, H. Alexander, D.S. Warner, R.S.B. Clark, S. Wisniewski, S.H. Graham, L.W. Jenkins, X. Ma\*, D.W. Marion, P. Safar. Safar Center for Resuscitation Research, Univ. of Pittsburgh, Pgh, PA, 15260 and Duke Univ., Raleigh Durham, NC, 27710.

Despite the routine use of fentanyl for initial sedation of patients after severe traumatic brain injury (TBI), it remains to be determined if it is the optimal sedative agent. Isoflurane is the most commonly used anesthetic in experimental models of TBI. Recent studies in experimental cerebral ischemia suggest that isoflurane is neuroprotective (vs fentanyl) in part by increasing cerebral blood flow (CBF) and reducing metabolic demands. To our knowledge, fentanyl has not been directly compared to isoflurane in experimental TBI. We hypothesized that isoflurane would be neuroprotective vs fentanyl when administered early after TBI in rats.

Male Sprague-Dawley rats ( $n=9$ /group) were subjected to controlled cortical impact to the left parietal cortex and randomized to receive either fentanyl (10 mcg/kg bolus followed by a 25 mcg/kg/h iv infusion) or isoflurane (1% by inhalation) for 4 h. Motor (beam balance, beam walking, d 1-5) and cognitive (Morris water maze performance, d 14-20) function were used to assess functional outcome and rats were perfused for the assessment of lesion and hippocampal volumes on d 21.

Rats treated with isoflurane had markedly better motor and cognitive function vs those treated with fentanyl (both  $p < 0.05$  on multiple days); although, there were no differences in either contusion or hippocampal volumes between treatment groups. We speculate that the increase in CBF in concert with metabolic suppression produced by isoflurane may be neuroprotective after TBI. Therefore the use of isoflurane may mask the beneficial effects of novel treatments tested in experimental models of TBI. In addition, fentanyl may not represent the optimal sedative agent, and may even be detrimental in the acute phase after severe TBI. Support: USArmy DAMD17-97-1-7009

**B6**

**FENTANYL VERSUS ISOFLURANE ANESTHESIA: EFFECT ON OUTCOME AFTER TRAUMATIC BRAIN INJURY IN RATS.** K.D. Statler, P.M. Kochanek, C.E. Dixon, H. Alexander, D.S. Warner, R.S.B. Clark, S. Wisniewski, S.H. Graham, L.W. Jenkins, X. Ma, D.W. Marion, P. Safar. Safar Center for Resuscitation Research, Univ. of Pittsburgh, Pgh, PA, 15260 and Duke Univ., Raleigh Durham, NC, 27710.

Despite routine use of fentanyl in patients after traumatic brain injury (TBI), it is unclear if it is the optimal analgesic. Isoflurane is routinely used in TBI models. Studies in cold lesion and ischemia suggest isoflurane is neuroprotective vs. fentanyl. To our knowledge, fentanyl and isoflurane have not been compared in TBI. We hypothesize that isoflurane is neuroprotective vs. fentanyl early after TBI. Male Sprague-Dawley rats (n=18) underwent controlled cortical impact and received 4 h of fentanyl (10 mcg/kg bolus, 50 mcg/kg/h infusion) or isoflurane (1% inhalation). Functional outcome (beam balance, beam walking and Morris water maze [MWM] tasks) and lesion volumes were assessed. Motor and MWM performances were better in rats treated with isoflurane vs. fentanyl ( $p < 0.05$ ). Lesion volumes were not different between groups. We speculate that isoflurane may be neuroprotective after TBI by increasing CBF, suppressing metabolism, and/or modulating excitotoxicity. Isoflurane may mask beneficial effects of novel treatments in experimental TBI. Finally, fentanyl may not be the optimal analgesic agent early after TBI in humans. **Support: USArmy#DAMD17-97-1-7009**

**B8**

**SECONDARY HYPOXIA 24 HOURS AFTER CONTROLLED CORTICAL IMPACT INCREASES INJURY IN CEREBRAL CORTEX BUT NOT HIPPOCAMPUS IN THE MOUSE.** F.A. Welsh\*, J. Keller, R. Raghupathi, G.P. Sinson, T.K. McIntosh. Dept. of Neurosurg., Univ. Penn. Sch. Med., Philadelphia, PA 19104.

The objective of this study was to determine whether an episode of hypoxia 24 hr after brain trauma augments histologic injury. Male C57BL/6 mice (n=10) were subjected to controlled impact injury using a deformation depth of 1 mm and impact velocity of 5 m/sec. After recovery for 24 hr, hypoxia was produced by lowering the percentage O<sub>2</sub> to 9% for 5 min and 7% for an additional 30 min. After an additional recovery period of 5 days, the animals were perfusion-fixed with FAM, and the brains were embedded in paraffin, sectioned, and stained with acid fuchsin/thionin. The stained sections were examined for histologic alteration and the volume of cortical infarction was measured.

Histopathologic alteration was not detected in any region of the contralateral hemisphere. Hypoxia significantly increased the size of the cortical lesion: Sham-hypoxia =  $1.95 \pm 0.42$  mm<sup>3</sup> (mean  $\pm$  SD, n=5) vs. Hypoxia =  $3.15 \pm 0.48$  (p<0.01). The only other histologic alteration detected was in the dentate granule cell layer of the ipsilateral hippocampus. There was both loss of neurons and acidophilic transformation of neurons in this layer. However, the number of acidophilic dentate granule cells was not altered by hypoxia (Sham-hypoxia =  $74 \pm 4$  vs. Hypoxia =  $70 \pm 19$ ). These results indicate that the traumatized cortex remains vulnerable for 24 hr to a level of hypoxia which does not cause histologic injury in the contralateral hemisphere.

Supported by NIH Grant NS-08803.

**B7**

**INFLUENCE OF POST-TRAUMATIC HYPOXIA ON BEHAVIORAL AND HISTOPATHOLOGICAL OUTCOME FOLLOWING MODERATE SPINAL CORD INJURY IN RATS.** Y. Yanagawa\*, A. Marcillo, R. Garcia, K. Loo, W. D. Dietrich. Department of Neurological Surgery and The Miami Project to Cure Paralysis, University of Miami School of Medicine, Miami, FL

Pulmonary dysfunction leading to secondary hypoxia is a common complication of spinal cord injury (SCI). The purpose of this study was to investigate the consequences of an induced post-traumatic hypoxic event following SCI. Forty-five female Sprague-Dawley rats were randomly assigned to 1 of 4 groups, including 1) laminectomy and normoxia, 2) laminectomy and hypoxia, 3) NYU weight-drop and normoxia, and 4) NYU weight-drop and hypoxia. For these studies, a moderate injury was induced by adjusting the height of the weight-drop (10 gm) to 12.5 mm above the exposed spinal cord (T10). Immediately after injury, PaO<sub>2</sub> in hypoxic rats was kept between 30-35 mmHg for 30 min, with 56% nitrous oxide, 31% nitrogen, and 13% oxygen. PaO<sub>2</sub> in the normoxic group was maintained over 100 mmHg, while PaCO<sub>2</sub> in all rats was maintained at 35-40 mmHg. The behavior of the rats was checked every 7 days using the BBB locomotor rating scale. Rats were sacrificed at 8 wks after behavioral testing and perfusion fixed for quantitative histopathological analysis of lesion areas. Although post-traumatic hypoxia tended to improve BBB scores, no significant difference in locomotor performance was demonstrated between the traumatized groups. In contrast, the percent of gray matter sparing at the impact epicenter was significantly reduced in hypoxic vs. normoxic SCI rats ( $p < 0.05$ ). These studies demonstrate that although moderate hypoxia following SCI does not significantly affect locomotive recovery, this secondary insult worsens gray matter pathology.

**B9**

**HYPOXIA EXACERBATES CA3 HIPPOCAMPAL NEURONAL DAMAGE AFTER FLUID PERCUSSION BRAIN INJURY IN RATS.** Namiko Nomura\*, Kojiro Wada, Yoshitaro Matsushita, Hiroshi Nawashiro, Katsuji Shima. Dept. of Neurosurgery, National Defense Medical College, Tokorozawa, Saitama, Japan

We have reported that increased vulnerability of hippocampal CA3 neurons to hypoxia after mild concussion. The present study was designed to determine if a model of moderate fluid-percussion (F-P) brain injury with hypoxia exacerbates hippocampal CA3 lesions, if those lesions are associated with apoptosis using the terminal deoxynucleotidyl transferase-mediated biotin-dUTP nick-end labeling method (TUNEL). Anesthetized Sprague-Dawley rats were injured with a moderate severity fluid percussion pulse (3.5-4.0 atmospheres) administered over the right parietal cortex. The experimental animals were divided into 2 groups, traumatic brain injury (TBI) group (n=6), which was subjected to TBI alone, and TBI + hypoxia group (n=6), which was subjected to TBI followed by 20 min of moderate hypoxia (F<sub>2</sub>O<sub>2</sub>: 10%). Three days following TBI, % neuronal density per 1-mm length of CA3 neurons in the ipsilateral hippocampus was significantly decreased in the TBI + hypoxia group ( $45.2 \pm 29.6$  %;  $p < 0.05$ ) compared to the TBI alone group ( $90.8 \pm 29.1$  %). No significant difference in the number of TUNEL positive cells was observed at 6-h, 24-h and 3-day (n=2) in both groups. These results suggest that TBI with moderate hypoxia induced more hippocampal damage due to not only apoptosis but also necrosis.

## SOLUBLE FAS IS INCREASED IN CSF FROM INFANTS AND CHILDREN AFTER HEAD INJURY

Neal A Seidberg, Acad Hosp of Pittsburgh, Pittsburgh, PA; Robert S B Clark, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA; Patrick M Kochanek, Safar Ctr for Resuscitation Res, Pittsburgh, PA; P David Adelson, Margaret A Satchell, Randall A Ruppel, Acad Hosp of Pittsburgh, Pittsburgh, PA; Keri L Janesko, Safar Ctr for Resuscitation Res, Pittsburgh, PA; Steven H Graham, Univ of Pittsburgh, Pittsburgh, PA

**Introduction:** Fas, a member of the TNF-receptor family, and its ligand FasL, provide a system for regulating intercellular programmed-cell death (PCD), where binding of FasL to Fas receptor triggers apoptosis. Fas has been identified on neurons and astrocytes, and FasL is present on microglia and inflammatory cells, thus, PCD in brain after injury may be regulated in part by Fas-FasL interactions. Accordingly, we examined CSF from infants and children after traumatic brain injury (TBI) for alterations in Fas and FasL. **Methods:** CSF was obtained from 20 patients with severe TBI who required neurointensive care including intraventricular catheter placement. Samples (n = 68) were collected on d 1 - 10 and were immediately centrifuged to remove cells. Control CSF was obtained from 14 children without TBI or meningitis. Fas and FasL were measured by ELISA. **Results:** TBI patients ranged in age from 1 mo - 11 y, 18 survived and 2 died. CSF Fas was increased 3-fold in TBI patients vs control (see table). Post hoc analysis also revealed an association between Fas and age (p = 0.05), and suspected child abuse. **Conclusions:** These data suggest that TBI induces alterations in the Fas/FasL system. Additional patients and multivariate analysis are required to further define associations between Fas and age and suspected child abuse. Therapies targeting cell death receptors, such as Fas, may represent effective strategies aimed at reducing PCD after TBI. **Support:** RO1 NS38620, KO8 NS01946, & P50 NS30318

Group	Fas (mU/ml)	FasL (pg/ml)
Control (n = 14)	269 ± 59	88 ± 14
TBI (n = 20)	759 ± 90*	163 ± 42
Accidental TBI (n = 16)	660 ± 164	168 ± 48
Suspected Child Abuse (n = 4)	1230 ± 94†	146 ± 93

\*mean ± SEM, p < 0.01 vs control. † p = 0.05 vs accidental.

## SYSTEMIC TREATMENT WITH A PAN-CASPASE INHIBITOR IMPROVES HIPPOCAMPAL NEURON SURVIVAL AFTER TRAUMATIC BRAIN INJURY IN MICE

Neal A Seidberg, Acad Hosp of Pittsburgh, Pittsburgh, PA; Steven H Graham, Univ of Pittsburgh, Pittsburgh, PA; Patrick M Kochanek, Safar Ctr for Resuscitation Res, Pittsburgh, PA; C. Edward Dixon, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA; Paula D Nathaniel, John Melick, Safar Ctr for Resuscitation Res, Pittsburgh, PA; Robert S B Clark, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA

**Introduction:** Traumatic brain injury (TBI) produces cell death both immediately after injury as a result of direct mechanical disruption, and in a delayed fashion as a result of secondary injury. Programmed cell death (PCD), or apoptosis, contributes to secondary cell death. The caspase family of cysteine proteases serve as effectors and executioners of PCD, and caspase inhibitors reduce cell death *in vitro* and *in vivo*. We hypothesized that systemic administration of the pan-caspase inhibitor boc-aspartyl(OMe)-fluoro-methylketone (BAF) would reduce hippocampal cell death after controlled cortical impact (CCI) in mice. **Methods:** Anesthetized mice were subjected to severe CCI to the left parietal cortex. Immediately after CCI mice were given 100 nmol BAF or vehicle (DMSO) i.p. in a randomized fashion. In one squadron of mice Caspase-3 activity was measured in injured brain at 24 h (n = 3-4/group). Separate mice underwent motor function tests (beam and round tube balance) at baseline and 24 h after CCI, then were killed for assessment of hippocampal neuron survival and DNA fragmentation using TUNEL (n = 5/group). **Results:** BAF treatment prevented the increase in relative caspase activity typically produced by CCI vs vehicle (85 vs 174% of uninjured hemisphere, p = 0.04). The number of surviving CA1 hippocampal neurons, cells vulnerable to PCD in this model, were increased in BAF treated mice vs vehicle (247 ± 28 vs 149 ± 13, p = 0.02). TUNEL-positive cells in hippocampus were similar between groups. Motor function was worse in BAF treated mice vs vehicle (p < 0.05). **Conclusions:** Pan-caspase inhibition using systemic treatment with BAF completely inhibits caspase-3 activity and enhances survival of vulnerable neurons 24 h after TBI in mice. Surprisingly, motor function at 24 h after TBI is worsened. This may be due to preservation of dysfunctional neurons, or nonspecific effects of BAF. Additional studies evaluating the long-term effects of pan-caspase inhibition after TBI are ongoing. Further investigation to determine the optimal treatment paradigm targeting caspase inhibition after TBI is warranted. **Support:** RO1 NS38620 and P50 NS30318.

## INCREASED ADENOSINE CONCENTRATION IN CEREBROSPINAL FLUID AFTER SEVERE TRAUMATIC BRAIN INJURY IN INFANTS AND CHILDREN: ASSOCIATION WITH SEVERITY OF INJURY

Courtney L Robertson, Michael J Bell, Patrick M Kochanek, P David Adelson, Randall Ruppel, Stephen Wisniewski, Zaichuan Mi, Keri L Janesko, Safar Ctr for Resuscitation Res, Pittsburgh, PA; Robert S B Clark, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA; Edwin K Jackson, Safar Ctr for Resuscitation Res, Pittsburgh, PA

**Introduction:** Traumatic brain injury (TBI) in children results in a myriad of pathophysiologic derangements that contribute to secondary injury, including hypoperfusion, energy failure and excitotoxicity. In addition, a number of endogenous neuroprotectants are produced after TBI, including adenosine, which increases cerebral blood flow and reduces metabolic demands<sup>1,2</sup>. In a prior evaluation of cerebrospinal fluid (CSF) of children following severe TBI<sup>3</sup>, we demonstrated increased peak levels of adenosine after TBI. In the current study, we evaluate the CSF of an expanded sample of infants and children following severe TBI, and examine the contribution of age, GCS, mechanism of injury and time after injury to CSF adenosine levels. **Methods:** Samples (n=304) of ventricular CSF were collected from 27 infants and children (2 mo to 14 y) during the first 7 d after severe TBI (GCS <8). Control CSF samples (n=21) were obtained from infants and children without TBI or meningitis. Adenosine was measured using HPLC. **Results:** Mean adenosine level was markedly increased in CSF of children following TBI vs control (peak 33.5 ± 9.5 and mean 24.3 ± 9.5 vs control mean 3.8 ± 0.5 nmol/L, p<0.001). Using a multiple regression model, the increase in CSF adenosine was independently associated with GCS ≤4 vs >4 and time after injury (both p<0.05). However, increased adenosine was not independently associated with mechanism of injury (abuse vs other) or age (≤4 vs >4y). **Conclusions:** We conclude CSF adenosine concentration is increased in infants and children after severe TBI. This increase was especially pronounced in children with the most severe injuries. Unlike mediators of secondary damage, such as glutamate<sup>4</sup>, adenosine was not associated with child abuse or age ≤4 y. We speculate that adenosine may play an important role in endogenous attempts at neuroprotection after TBI. <sup>1</sup>Cerebrovasc Brain Metab Rev 1:26, 1989; <sup>2</sup>J Cereb Blood Flow Metab 14:853, 1994; <sup>3</sup>Crit Care Med 24:A136, 1996; <sup>4</sup>Pediatrics 102:704, 1998 **Acknowledgment:** Univ of Pittsburgh Center for Injury Research and Control/CDC, Laerdal Foundation, and NS 38087

## ISOFLURANE IMPROVES LONG-TERM NEUROLOGIC OUTCOME COMPARED TO FENTANYL AFTER TRAUMATIC BRAIN INJURY IN RATS

Kimberly D Statler, Acad Hosp of Pittsburgh, Pittsburgh, PA; Patrick M Kochanek, Safar Ctr for Resuscitation Res, Pittsburgh, PA; C. Edward Dixon, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA; Henry L Alexander, Safar Ctr for Resuscitation Res, Pittsburgh, PA; David S Warner, Duke Univ Med Ctr, Durham, NC; Robert S B Clark, Univ Pittsburgh and Safar Ctr for Resuscitation Res, Pittsburgh, PA; Steven R Wisniewski, Univ of Pittsburgh, Pittsburgh, PA; Larry W Jenkins, Safar Ctr for Resuscitation Res, Pittsburgh, PA; Donald W Marion, Univ of Pittsburgh, Pittsburgh, PA; Peter J Safar, Safar Ctr for Resuscitation Res, Pittsburgh, PA

**Introduction:** Despite routine use of fentanyl in patients after traumatic brain injury (TBI), it is unclear if it is the optimal sedative/analgesic agent. Isoflurane is commonly used in models of TBI. Recent studies in cerebral ischemia and focal cryogenic lesion suggest that isoflurane may be neuroprotective vs fentanyl<sup>1,2</sup>. To our knowledge, fentanyl and isoflurane have not been directly compared in TBI. We hypothesized that isoflurane would be neuroprotective vs fentanyl when given early after TBI in rats. **Methods:** Adult rats (n=18) underwent controlled cortical impact (CCI) with physiologic monitoring and then received 4h of N<sub>2</sub>O/O<sub>2</sub> (2:1) and either fentanyl (10 mcg/kg bolus, 50 mcg/kg/h infusion) or isoflurane (1% inhalation). Shams (n=8) underwent identical preparation and anesthesia but no CCI. Functional outcome (beam balance, beam walking, Morris water maze [MWM] tasks) was assessed over 20d in injured and sham rats. Lesion volume was quantified on d21. Additional rats (n=14) underwent CCI and anesthesia as described above with intracranial pressure (ICP) monitoring (Codman intraparenchymal transducer) for 4h. Brain water (wet-dry weight method) was assessed at the end of the anesthetic period. **Results:** After injury, motor and MWM performances were better in isoflurane vs fentanyl treated rats (p<0.05, ANOVA) but did not differ between shams. Lesion volumes were similar between groups. There was increased frequency of ICP>20 mm Hg and higher brain water in rats treated with isoflurane vs fentanyl (p<0.05, ANOVA). **Conclusions:** Rats treated with isoflurane had improved long-term functional outcome after CCI compared to those treated with fentanyl, despite increases in ICP and brain water. We speculate that isoflurane may mediate improved long-term functional outcome after CCI in rats through promotion of cerebral blood flow, suppression of metabolism, and/or modulation of excitotoxicity. Fentanyl may not be the optimal sedative/analgesic agent early after TBI in humans. **Support:** USArmy#DAMD17-97-1-7009. 1. Mira, et al. Anesthesiology 1998; 391-400. 2. Murr, et al. Anesth Analg 1995; 80: 1108-15.